



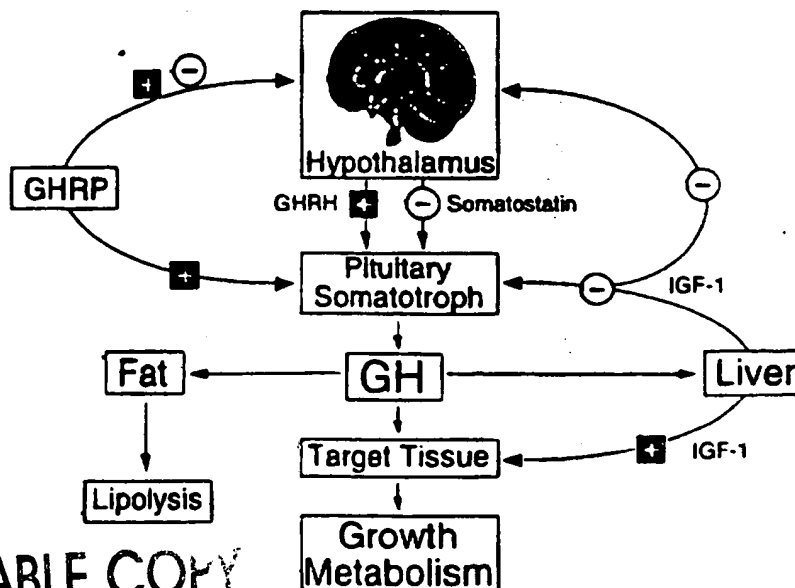
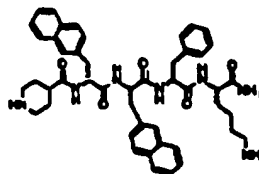
INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6 : C07K 14/60, 5/023, 5/027, 5/03, 5/087, 5/107, 5/117, C07D 211/62, 40/12, A61K 38/12, 38/05, 38/06, 38/07, 31/445		A2	(11) International Publication Number: WO 96/15148
			(43) International Publication Date: 23 May 1996 (23.05.96)
(21) International Application Number: PCT/US95/14968		(81) Designated States: AL, AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TT, UA, UG, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, LS, MW, SD, SZ, UG).	
(22) International Filing Date: 16 November 1995 (16.11.95)			
(30) Priority Data: 08/340,767 16 November 1994 (16.11.94) US			
(71) Applicant: GENENTECH, INC. [US/US]; 460 Point San Bruno Boulevard, South San Francisco, CA 94080-4990 (US).			
(72) Inventors: SOMERS, Todd, C.; 706 Bounty Drive #610, Foster City, CA 94404 (US). ELIAS, Kathleen, A.; 2006 16th Avenue, San Francisco, CA 94116 (US). CLARK, Ross, G.; 711 Ursula Avenue, Pacifica, CA 94044 (US). McDOWELL, Robert, S.; 1264 Church Street, San Francisco, CA 94114 (US). STANLEY, Mark, S.; 284 Lauren Avenue, Pacifica, CA 94044 (US). BURNIER, John, P.; 202 Sterling Avenue, Pacifica, CA 94044 (US). RAWSON, Thomas, E.; 140 Hamilton Avenue, Mountain View, CA 94043 (US).			
(74) Agents: WINTER, Daryl, B. et al.; Genentech, Inc., 460 Point San Bruno Boulevard, South San Francisco, CA 94080-4990 (US).		<p>Published</p> <p><i>Without international search report and to be republished upon receipt of that report.</i></p>	

(54) Title: **LOW MOLECULAR WEIGHT PEPTIDOMIMETIC GROWTH HORMONE SECRETAGOGUES**

(57) Abstract

The present invention comprises growth hormone releasing peptides/peptidomimetics (GHRP) capable of causing release of growth hormone from the pituitary. Compositions containing the GHRP's of this invention are used to promote growth in mammals either alone or in combination with other growth promoting compounds, especially IGF-1. In a method of this invention GHRP's in combination with IGF-1 are used to treat Type II diabetes. An exemplary compound (1) of this invention is provided above.



BEST AVAILABLE COPY

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	GB	United Kingdom	MR	Mauritania
AU	Australia	GE	Georgia	MW	Malawi
BB	Barbados	GN	Guinea	NE	Niger
BE	Belgium	GR	Greece	NL	Netherlands
BF	Burkina Faso	HU	Hungary	NO	Norway
BG	Bulgaria	IE	Ireland	NZ	New Zealand
BJ	Benin	IT	Italy	PL	Poland
BR	Brazil	JP	Japan	PT	Portugal
BY	Belarus	KE	Kenya	RO	Romania
CA	Canada	KG	Kyrgyzstan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo	KR	Republic of Korea	SE	Sweden
CH	Switzerland	KZ	Kazakhstan	SI	Slovenia
CI	Côte d'Ivoire	LI	Liechtenstein	SK	Slovakia
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CN	China	LU	Luxembourg	TD	Chad
CS	Czechoslovakia	LV	Latvia	TG	Togo
CZ	Czech Republic	MC	Monaco	TJ	Tajikistan
DE	Germany	MD	Republic of Moldova	TT	Trinidad and Tobago
DK	Denmark	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	US	United States of America
FI	Finland	MN	Mongolia	UZ	Uzbekistan
FR	France			VN	Viet Nam
GA	Gabon				

LOW MOLECULAR WEIGHT PEPTIDOMIMETIC GROWTH HORMONE SECRETAGOGUES

FIELD OF THE INVENTION

5 The invention relates to synthetic peptidomimetics having growth hormone releasing activity in mammals. The peptidomimetics of this invention are used to stimulate the release of endogenous growth hormone (GH) in mammals needing elevation of serum growth hormone levels.

BACKGROUND OF THE INVENTION

10 GH secretion is known to be inhibited by the hypothalamic hormone somatostatin (SS) and stimulated by GH-releasing hormone (GHRH) in all mammalian species studied including humans. In man, GH is released from the anterior pituitary somatotrophs in pulsatile secretory bursts occurring about 4-8 times in each 24 hour period (Devesa, J., *et al.*, *Trends Endocrinol Metab.* 3:175-183 [1992] and Mason, W. T., *et al.*, *Acta Paediatr Suppl* 388:84-92 [1993]). This
15 episodic release pattern seems to be optimal for inducing the physiological effects of GH since many target tissues appear to be more sensitive to the frequency than the total amount of GH arriving at the target tissue (Robinson and Clark *Growth Hormone : Basic and Clinical Aspects* Isaksson, Binder, Hall and Hokfelt eds., Amsterdam, p109-127 [1987]). It is believed the
20 episodic secretion of GH is caused by the rhythmic alternate release of the excitatory 44-amino acid peptide GHRH and the inhibitory tetradecapeptide SS, regulated through the "pituitary-hypothalamus axis" (see Figure 1). Secreted GH, in turn, both directly and indirectly through IGF-1 appears to maintain this rhythm by stimulating SS and inhibiting GHRH release. Other neurotransmitters also modulate GH release usually by stimulating or inhibiting SS release.
25 Additionally, other factors including exercise, sleep, glucocorticoids, thyroid hormones (e.g. TSH), sex steroids (e.g. testosterone and 17- β estradiol), free fatty acids, amino acids (e.g. arginine and ornithine), and glucose levels further modulate GH release.

 In addition to the two primary endogenous regulators of GH release, SS and GHRH, a number of other peptidyl/nonpeptidyl compounds have been shown to stimulate GH release
30 primarily through the pituitary-hypothalamus axis. These include the peptides galanin, pituitary adenylate cyclase-activation peptide (PACAP), delta sleep-inducing peptide (DSIP), and angiotensin II. These peptides, however, generally lack specificity for GH release. A number of structurally diverse nonpeptidyl GH secretagogues (e.g. Talipexole and Clonidine) are reported to stimulate GH release *in vitro* and *in vivo*, but these compounds are believed to
35 mediate their effect through cholinergic, adrenergic, dopaminergic or serotonergic pathways and thus also lack GH releasing specificity.

 Apart from GHRH, the GH secretagogues having the greatest GH releasing specificity and thus having the greatest therapeutic potential are the growth hormone releasing

peptides/peptidomimetics (GHRP's) (Bowers, *J. Pediatr. Endocrinol.* 6:21-31 [1993]; and Schoen *et al.*, *Annual Reports in Medicinal Chemistry*, 28:177-186 [1993]). These compounds can activate the pituitary-hypothalamus axis (Dickson *et al.*, *Neuroscience* 53:303-306 [1993]) and act directly on the pituitary somatotroph (see Figure 1) by an independent (non-GHRH, non-opiate and non-SS) secretory pathway. Compounds of this class can therefore be characterized by their independent GH releasing pathway. For example, somatotroph cells maximally stimulated with GHRP's can release additional GH when treated with GHRH and *vice versa*. Similarly the inhibitory effects of specific antagonists to GHRH or GHRP's have no effect on stimulation of GH release by the opposite secretagogue *in vitro*. These compounds also exhibit dose-dependent desensitization or attenuation of GH release after continuous exposure with the same or different compounds of the GHRP class. Furthermore, structurally related biologically inactive cognate GHRP compounds capable of inhibiting GH release of a particular GHRP have no effect on GHRH agonism. These effects support the independent pathway model and serve as experimental criteria for compounds belonging to the GHRP class. Surprisingly, while the GHRH receptor has been cloned in a number of species including man (Gaylin *et al.*, *Mol. Endo.* 7:77-84 [1993]), the GHRP receptor has remained elusive.

The paradigm compounds of the GHRP class are the synthetic methionine-enkephalin derived GHRP's identified by Bowers *et al.*, *Endocrinology* 106:663-667 (1980) and Momany *et al.*, *Endocrinology* 108:31-39 (1981). The most widely studied GHRP is referred to as "GHRP-6" (Momany *et al.*, *Endocrinology* 114:1531-1536 [1984]; and Bowers *et al.*, *Endocrinology* 114:1537-1545 [1984]) which has been shown; to be specific for GH release, has no reported long term toxicity, is well tolerated, and can elevate serum GH in a dose-dependent manner in normal humans (Bowers, *J. Pediatr. Endocrinol.* 6:21-31 [1993]). GHRP-6 is active in a dose-dependent manner when administered either iv, intranasally or orally, though it is poorly absorbed orally (~0.3%). More potent second generation hepta- and hexapeptides, "GHRP-1" and "GHRP-2" (also known as KP 102), of this class have been described more recently, though these compounds are also expected to be poorly absorbed orally.

His-D Trp-Ala-Trp-D Phe-Lys-NH₂

30

(HwAWfK-NH₂)

GHRP-6

Ala-His-D βNal-Ala-Trp-D Phe-Lys-NH₂

(AHbAWfK-NH₂)

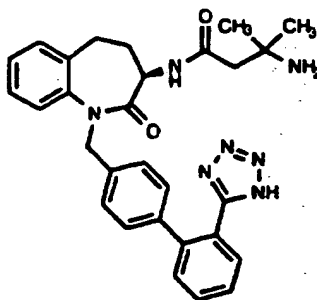
35

GHRP-1

D Ala-D β Nal-Ala-Trp-D Phe-Lys-NH₂(abAWfK-NH₂)

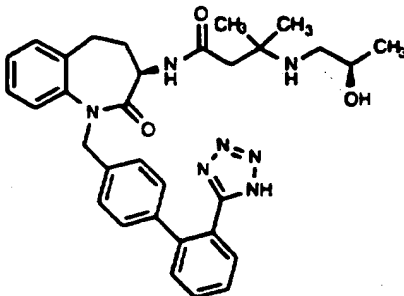
GHRP-2

- 5 More recently, nonpeptidyl benzolactam GH secretagogues that appear to use the same alternative signal transduction pathway as GHRP-6 have been described (Smith, R.G. *et al.*, *Science* 260:1640-1643 [1993] and U.S. Patent No. 5,206,235). The benzolactam L-692,429 in combination with GHRP-6 at concentrations that maximally stimulated GH release produced no additional GH release. Conversely, GHRH and L-692,429 were reported to give a synergistic increase in GH secretion. GHRH and L-692,429 were also reported to effect a common transient desensitization pattern indicating these compounds operate through a common receptor pathway. L-692,429 is reported to be about 6-fold less potent than GHRP-6 and to be specific for GH release, except for some *in vivo* ACTH and cortisol release.
- 10



L-692,429

- 15 A more potent analogue of L-692,429 having a potency in the rat pituitary cell assay slightly greater than GHRP-6 has also been reported (Schoen W. R. *et al.*, *Bioorg. & Medicinal Chem. Lett.* 4:1117-1122 [1994]). This compound, L-692,585, presumably causes GH release by the same alternative pathway as GHRP-6.



L-692,585

A number of these compounds (e.g., "GHRP-6" and L-692,429) are reported to be safe and effective in promoting endogenous GH release in humans, however, there remain problems with oral availability and specificity.

5

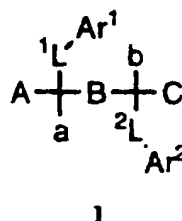
OBJECTS OF THE INVENTION

It is an object of this invention to provide novel GH secretagogues that promote the release of endogenous growth hormone in mammals. It is a further object to provide GH secretagogues that provide a synergistic increase in GH secretion when combined with GHRH. It is still a further object of this invention to provide more potent GH secretagogues than those of the prior art, especially "GHRP-6", "GHRP-1", "GHRP-2", L-692,429 and L-692,585. It is a further object to provide GH secretagogues that are specific for GH release and do not cause significant release of other hormones, especially; LH, FSH, TSH, ACTH, prolactin, vasopressin, oxytocin, insulin and cortisol. These and other objects of the invention will be apparent from the following specification.

15

SUMMARY OF THE INVENTION

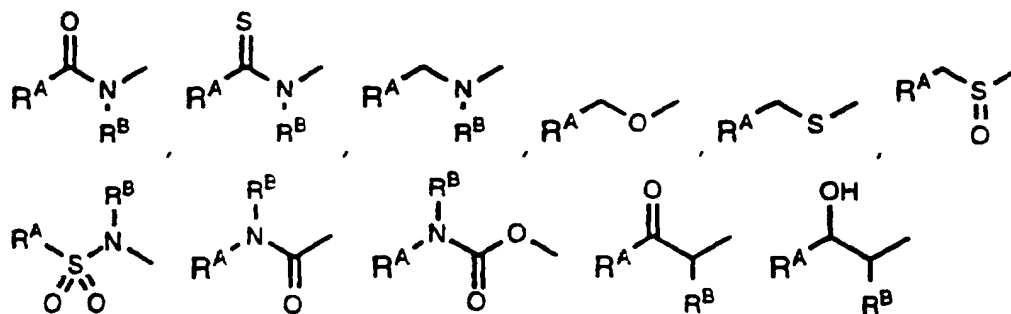
The objects of this invention have been achieved by providing a compound represented by structural formula (I):

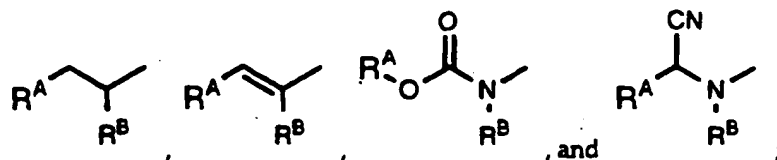


20

where the symbols in formula (I) define the following groups:

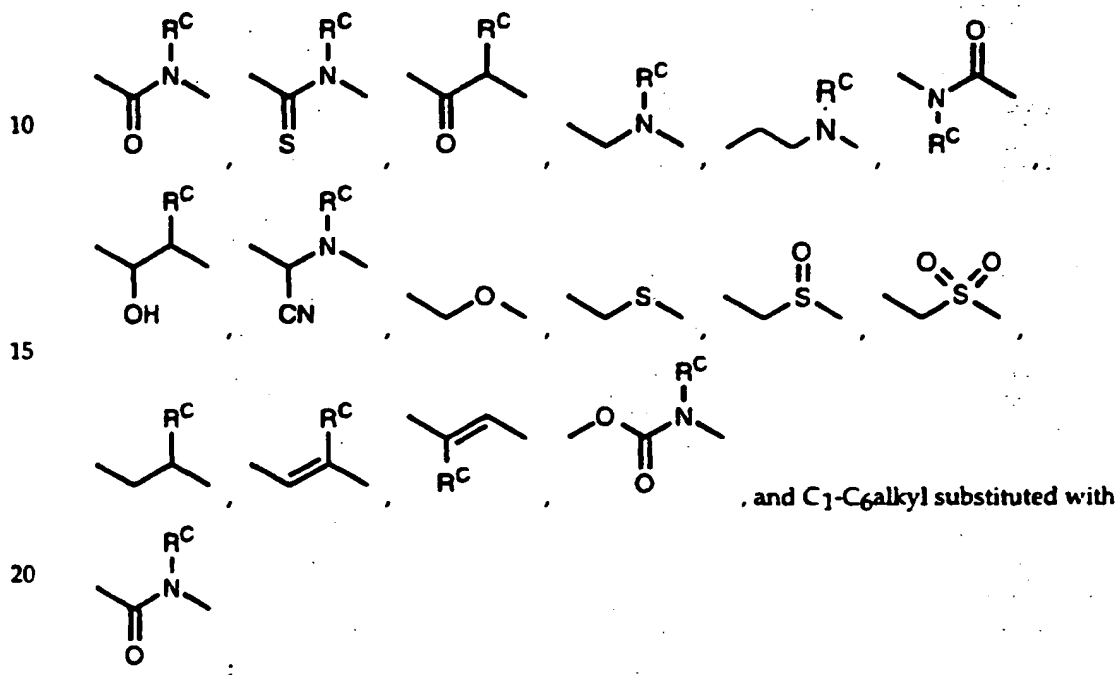
A is selected from





5

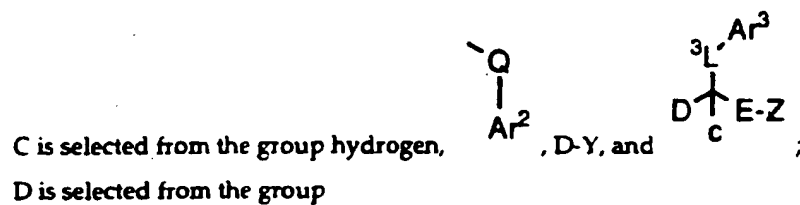
B is selected from



15

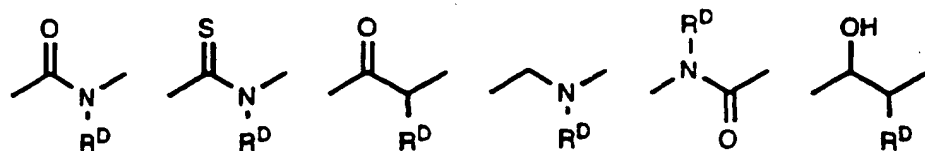
20

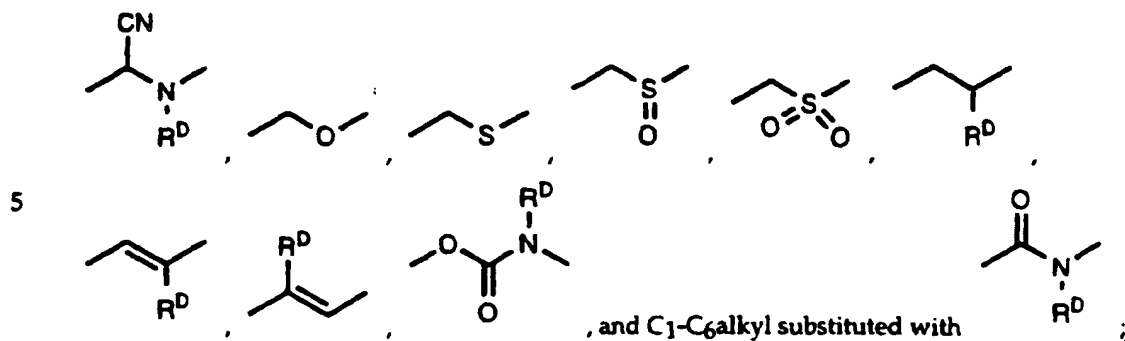
B may optionally be selected from the group a covalent bond, and
 25 C_1-C_3 alkyl, when L^2 is $-N(R^C)-Q$:



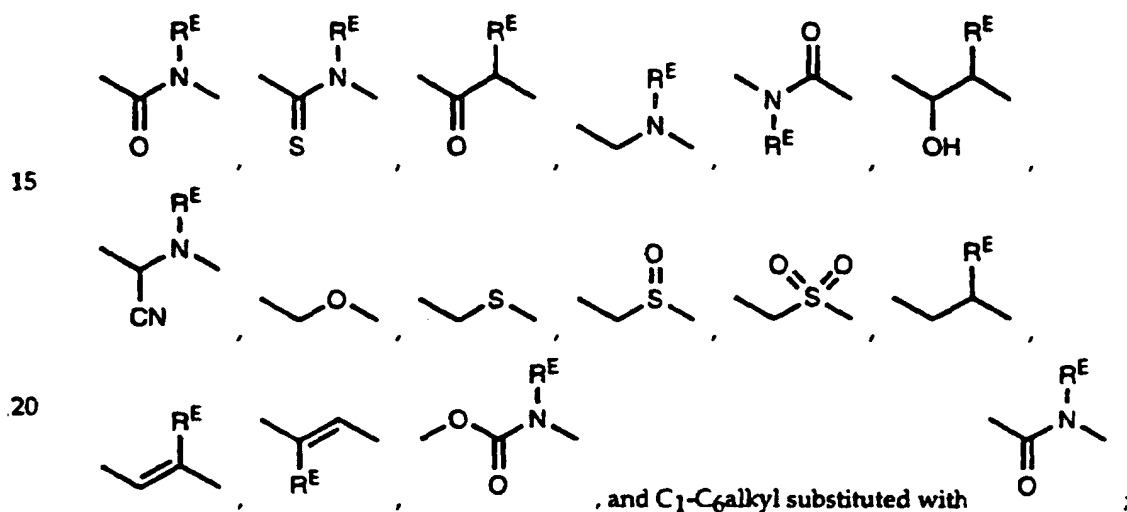
30

35

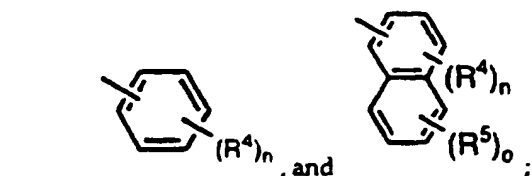




10 E is selected from the group



25 Ar¹ and Ar² are each independently selected from substituted or unsubstituted aryl and substituted or unsubstituted heterocycle, preferably indoyl substituted with (R⁴)_n



Ar¹ and Ar² are independently selected from hydrogen, and C₁-C₆alkyl;
when R^B or R^C are L¹-Ar¹ or L²-Ar²;



Ar³ is selected from hydrogen, and C₁-C₆alkyl;

when R^D is L^3 -Ar³;

Ar¹ together with a, Ar² together with b and Ar³ together with c, each pair together with the carbon to which they are attached may independently form a 5 or 6 member carbocyclic ring;

5 a, b and c are independently selected from hydrogen, and C₁-C₆alkyl;

n and o are independently 1, 2 or 3;

L^1 is selected from -CH₂-O-, -CH₂-CH₂-O-, -CH₂-, -CH₂-CH₂-, and -CH₂-CH₂-CH₂-;

L^2 and L^3 are independently selected from a covalent bond, -O-, -O-CH₂-, -N(R^C)-Q, and L^1 ;

10 Q is selected from the group -L²-, -S(=O)₂-L²-, -C(=O)-, -C(=O)-O-, -CH(X)-, and -CH(X)-CH₂-;

R^A is selected from the group C₀-C₃alkyl-heterocycle where the heterocycle comprises a mono-, bi-, or tricycle containing 5-12 ring atoms, one or two of which are heteroatoms selected from O, S, and N, provided at least one heteroatom is N, where any N atom is
15 optionally substituted with R¹, C₀-C₆alkyl substituted with one or two substituents selected from the group NR²R³, imidazolynyl, pyridynyl, dihydropyridynyl, and piperidynyl;

R^B, R^C and R^D are selected from the group R^A, L¹-Ar¹, L²-Ar², hydrogen, C₁-C₆alkyl, and halo(F, Cl, Br, I)C₁-C₆alkyl;

R^A and R^B together with the N to which they are bonded may form a 5- or 6-member
20 heterocycle, optionally containing one additional heteroatom selected from O, S, and N where any N is optionally substituted with R¹, any carbon is optionally substituted with R⁶ and where the heterocycle is optionally fused to a phenyl ring, optionally substituted with R⁴;

R¹ is selected from hydrogen, C₁-C₆alkyl, C(=O)-C₁-C₆alkyl, C(=O)-NR²R³, C(=NR²)-NR²R³, C(=O)O-C₁-C₆alkyl, and halo(F, Cl, Br, I)C₁-C₆alkyl, hydroxyC₁-C₆alkyl,
25 dihydroxyC₁-C₆alkyl;

R² and R³ are independently selected from R¹ and piperidynyl;

R² and R³ together with the N to which they are bonded may form a 5- or 6-member heterocycle, optionally containing one additional hetero atom selected from O, S, and N where any N is optionally substituted with R¹, any carbon is optionally substituted with R⁶ and
30 where the heterocycle is optionally fused to a phenyl ring, optionally substituted with R⁴;

R⁴ and R⁵ are independently selected from the group hydrogen, halo(F, Cl, Br, and I), cyano, amino, amido, nitro, hydroxy, C₁-C₆alkyl optionally substituted with 1-3 R⁶, C₂-C₆alkynyl optionally substituted with 1-3 R⁶, C₁-C₆alkyloxy optionally substituted with 1-3 R⁶, C₁-C₆acylamino optionally substituted with 1-3 R⁶, C₁-C₆alkylcarbonyl optionally substituted with 1-3 R⁶, C₁-C₆alkyloxycarbonyl optionally substituted with 1-3 R⁶, N-(C₁-C₆alkyl), N-(C₁-C₆acyl)amino optionally substituted with 1-3 R⁶, N-(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶, N,N-di(C₀-C₆alkyl)amino optionally substituted with 1-3
35

R^6 , N,N-di(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R^6 , C₁-C₄perfluoroalkyl, and C₁-C₃perfluoroalkoxy;

R^6 is selected from the group COOR², O(C=O)R², CONR²R³, cyano, NR²R³, NR²COR³, azido, nitro, and hydroxy;

5 R^7 is selected from the group R^6 , C₆-C₁₀aryl optionally substituted with halo(F, Cl, Br, and I), cyano, amino, amido, nitro, hydroxy, C₁-C₄perfluoroalkyl, and C₁-C₃perfluoroalkoxy;

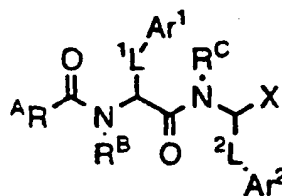
X is selected from the group hydrogen, C₀-C₆alkyl optionally substituted with 1-3 R^6 , C₀-C₆alkyl-O-C₁-C₆alkyl optionally substituted with 1-2 R^6 , and C₁-C₆acyl optionally substituted with a group selected from L²-Ar², R^A, and R^6 ;

10 Y is selected from the group -(C=O)-R^A, C₁-C₆alkyl substituted with 1-2 R^7 , C₂-C₆alkynyl optionally substituted with 1-2 R^7 , C₂-C₆alkylenyl optionally substituted with 1-2 R^7 , and C₁-C₆alkyloxy optionally substituted with 1-2 R^7 ,

Y and R^D together with the N to which they are bonded may form a 5- or 6-member heterocycle, optionally containing one additional hetero atom selected from O, S, and N where
15 any N is optionally substituted with R¹, any carbon is optionally substituted with R^7 and where the heterocycle is optionally fused to a phenyl ring;

Z is selected from the group C₁-C₆alkyl substituted with 1-2 R^7 , C₂-C₆alkynyl optionally substituted with 1-2 R^7 , C₂-C₆alkylenyl optionally substituted with 1-2 R^7 , C₁-C₆alkyloxy optionally substituted with 1-2 R^7 and piperidinyl; and pharmaceutically
20 acceptable salts thereof.

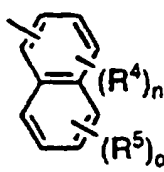
In one embodiment of the invention the compound preferably has a molecular weight between 400-650 da and is represented by formula II



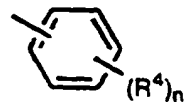
II

where the symbols in formula II are defined as follows:

25 Ar¹ and Ar² are each independently selected from indoyl,



and



n and o are independently 1, 2 or 3;

L¹ is selected from -CH₂-O-, -CH₂-CH₂-O-, -CH₂-, -CH₂-CH₂-, and -CH₂-CH₂-CH₂-;

L² is selected from a covalent bond, -O-, and L¹;

5 R^A is selected from the group C₀-C₃alkyl-heterocycle, -O-C₀-C₃alkyl-heterocycle, and -NR²-C₂-C₆alkyl-heterocycle, where the heterocycle comprises a mono-, bi-, or tricycle containing 5-12 ring atoms, one or two of which are heteroatoms selected from O, S, and N, provided at least one heteroatom is N, where any N atom is optionally substituted with R¹, C₀-C₆alkyl substituted with one or two substituents, O-C₂-C₆alkyl substituted with one or two substituents, and NR²-C₂-C₆alkyl substituted with one or two substituents where the
10 substituents are selected from the group NR²R³, imidazolynyl, pyridynyl, dihydropyridynyl, and piperidynyl;

R^B and R^C are selected from the group hydrogen, C₁-C₆alkyl optionally substituted with a group selected from NR²R³, and phenyl-C₁-C₃-NR²R³, and halo(F, Cl, Br, I)C₁-C₆alkyl;

R¹ is selected from hydrogen, C₁-C₆alkyl, C(=O)-C₁-C₆alkyl, C(=O)-NR²R³, C(=NR²)-NR²R³, C(=O)O-C₁-C₆alkyl, and halo(F, Cl, Br, I)C₁-C₆alkyl, C₁-C₆alkoxyalkyl or
15 (hydroxylalkyl);

R² and R³ are independently selected from hydrogen, C₁-C₆alkyl, piperidynyl, and halo(F, Cl, Br, I)C₁-C₆alkyl;

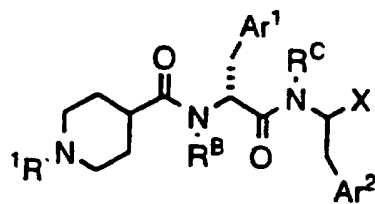
R² and R³ together with the N to which they are bonded may form a 5- or 6-member
20 heterocycle, optionally containing one additional hetero atom selected from O, S, and N where any N is optionally substituted with R¹, any carbon is optionally substituted with R⁶ and where the heterocycle is optionally fused to a phenyl ring, optionally substituted with R⁴;

R⁴ and R⁵ are independently selected from the group hydrogen, halo(F, Cl, Br, and I), cyano, amino, amido, nitro, hydroxy, C₁-C₆alkyl optionally substituted with 1-3 R⁶, C₂-C₆alkynyl optionally substituted with 1-3 R⁶, C₁-C₆alkyloxy optionally substituted with 1-3 R⁶, C₁-C₆acylamino optionally substituted with 1-3 R⁶, C₁-C₆alkylcarbonyl optionally substituted with 1-3 R⁶, C₁-C₆alkyloxycarbonyl optionally substituted with 1-3 R⁶, N-(C₁-C₆alkyl), N-(C₁-C₆acyl)amino optionally substituted with 1-3 R⁶, N-(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶, N,N-di(C₀-C₆alkyl)amino optionally substituted with 1-3 R⁶, N,N-di(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶, C₁-C₄perfluoroalkyl, and C₁-C₃perfluoroalkoxy;
25
30

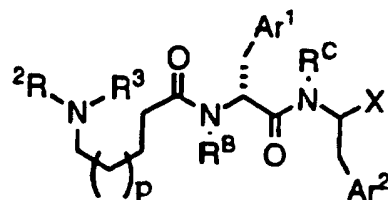
R⁶ is selected from the group COOR², O(C=O)R², CONR²R³, cyano, NR²R³, NR²COR³, azido, nitro, and hydroxy;

X is selected from the group hydrogen, oxo (=O), COOR², CONR²R³, C₀-C₆alkyl-O-C₁-C₆alkyl optionally substituted with 1-2 R⁶, and C₁-C₆alkyl optionally substituted with 1-2 R⁶; and pharmaceutically acceptable salts thereof.
35

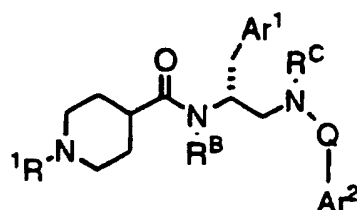
Alternative compounds of this embodiment may be represented by formula IIa-IIg



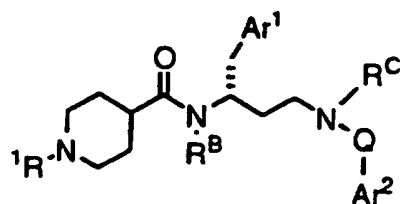
IIa



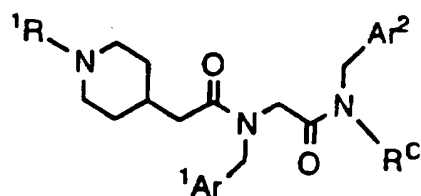
IIb



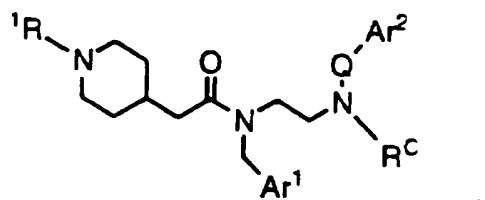
IIc



IIId

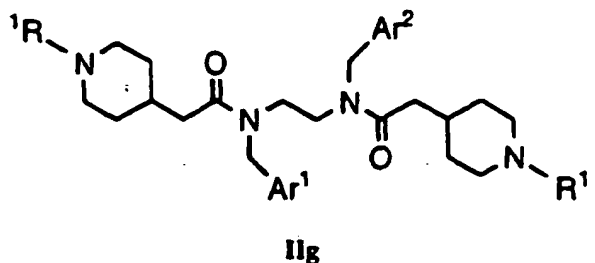


IIe



IIIf

, and



where Ar^1 , Ar^2 , R^B , R^C , R^1 , R^2 , R^3 , R^6 , Q and X are defined above, and p is 0, 1 or 2.

Optionally the Ar^1 , Ar^2 , R^B , R^C , R^1 , R^2 , R^3 , R^6 and X are defined as follows:



Ar^1 and Ar^2 are each independently selected from indoyl, and

5 R^B and R^C are selected from the group hydrogen, and methyl;

R^1 is selected from hydrogen, C_1 - C_6 alkyl, C_2 - C_6 alkyl substituted with 1 or 2 hydroxy groups, $C(=O)$ - C_1 - C_6 alkyl, $C(=O)$ - NR^2R^3 , $C(=NR^2)$ - NR^2R^3 , $C(=O)O$ - C_1 - C_6 alkyl, and halo(F, Cl, Br, I)- C_1 - C_6 alkyl;

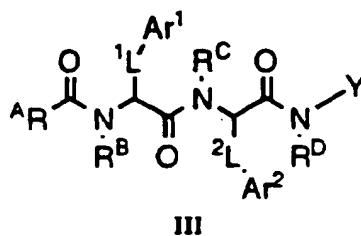
10 R^2 and R^3 are independently selected from hydrogen, C_1 - C_6 alkyl, piperidinyl, and halo(F, Cl, Br, I)- C_1 - C_6 alkyl;

R^2 and R^3 together with the nitrogen to which they are attached may form piperidinyl, pyrrolidinyl, piperazinyl, and morpholinyl;

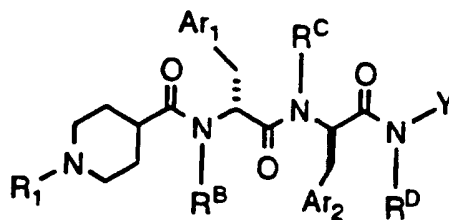
R^6 is selected from the group $COOR^2$, $O(C=O)R^2$, $CONR^2R^3$, cyano, NR^2R^3 , NR^2COR^3 , azido, nitro, and hydroxy;

15 X is selected from the group hydrogen, oxo ($=O$), $COOR^2$, $CONR^2R^3$, C_0 - C_6 alkyl- O - C_1 - C_6 alkyl optionally substituted with 1-2 R^6 , and C_1 - C_6 alkyl optionally substituted with 1-2 R^6 ; and pharmaceutically acceptable salts thereof.

In an alternative embodiment of the invention the compound is represented by structural formula III-IIIi

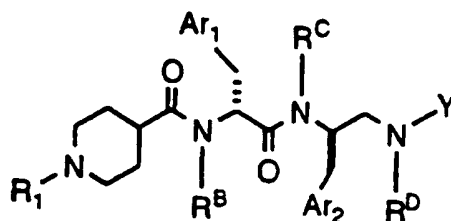


5



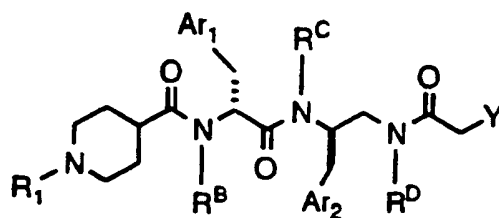
IIIa

10



IIIb

15



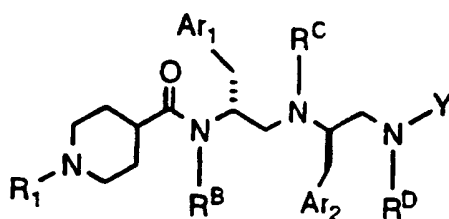
IIIc

20



IIId

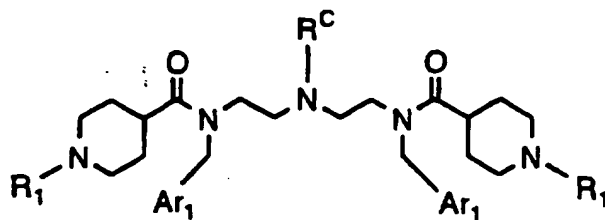
25



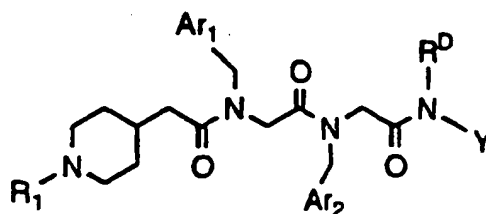
IIId

30

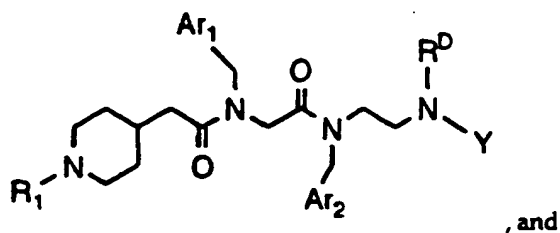
35



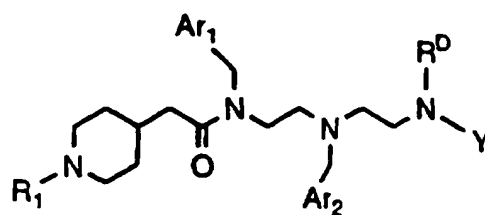
III f



III g



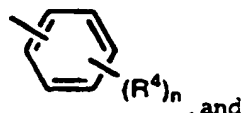
III h



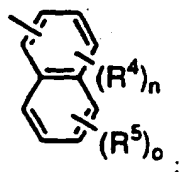
III i

where the symbols in formula III-IIIi are defined as follows:

Ar¹ and Ar² are each independently selected from indolyl,



and



n and o are independently 1, 2 or 3;

L¹ and L² are independently selected from -CH₂-O-, -CH₂-CH₂-CH₂-O-, -CH₂-, -CH₂-CH₂-, and -CH₂-CH₂-CH₂-;

R^A is selected from the group C₀-C₃alkyl-heterocycle, -O-C₀-C₃alkyl-heterocycle, and -NR²-C₂-C₆alkyl -heterocycle, where the heterocycle comprises a mono-, bi-, or tricycle containing 5-12 ring atoms, one or two of which are heteroatoms selected from O, S, and N, provided at least one heteroatom is N, where any N atom is optionally substituted with R¹, C₀-C₆alkyl substituted with one or two substituents, O-C₂-C₆alkyl substituted with one or two substituents, and NR²-C₂-C₆alkyl substituted with one or two substituents where the substituents are selected from the group NR²R³, imidazolynyl, pyridynyl, dihydropyridynyl, and piperidynyl;

R^B , R^C and R^D are selected from the group hydrogen, C₁-C₆alkyl optionally substituted with a group selected from NR²R³, and phenyl-C₁-C₃-NR²R³, and halo(F, Cl, Br, I)C₁-C₆alkyl;

R¹ is selected from hydrogen, C₁-C₆alkyl, C(=O)-C₁-C₆alkyl, hydroxyalkyl C(=O)-NR²R³, C(=NR²)-NR²R³, C(=O)O-C₁-C₆alkyl, and halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ are independently selected from hydrogen, C₁-C₆alkyl, piperidynyl, and halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ together with the N to which they are bonded may form a 5- or 6-member heterocycle, optionally containing one additional hetero atom selected from O, S, and N where any N is optionally substituted with R¹, any carbon is optionally substituted with R⁶ and where the heterocycle is optionally fused to a phenyl ring, optionally substituted with R⁴;

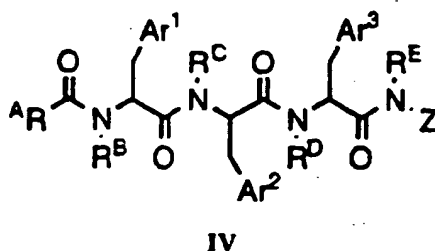
R⁴ and R⁵ are independently selected from the group hydrogen, halo(F, Cl, Br, and I), cyano, amino, amido, nitro, hydroxy, C₁-C₆alkyl optionally substituted with 1-3 R⁶, C₂-C₆alkynyl optionally substituted with 1-3 R⁶, C₁-C₆alkyloxy optionally substituted with 1-3 R⁶, C₁-C₆acylamino optionally substituted with 1-3 R⁶, C₁-C₆alkylcarbonyl optionally substituted with 1-3 R⁶, C₁-C₆alkyloxycarbonyl optionally substituted with 1-3 R⁶, N-(C₁-C₆alkyl), N-(C₁-C₆acyl)amino optionally substituted with 1-3 R⁶, N-(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶, N,N-di(C₀-C₆alkyl)amino optionally substituted with 1-3 R⁶, N,N-di(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶, C₁-C₄perfluoroalkyl, and C₁-C₃perfluoroalkoxy;

R⁶ is selected from the group COOR², O(C=O)R², CONR²R³, cyano, NR²R³, NR²COR³, azido, nitro, and hydroxy;

R⁷ is selected from the group R⁶, and C₆-C₁₀aryl optionally substituted with halo(F, Cl, Br, and I), cyano, amino, amido, nitro, hydroxy, C₁-C₄perfluoroalkyl, and C₁-C₃perfluoroalkoxy;

Y is selected from the group C₁-C₆alkyl substituted with 1-2 R⁷, C₂-C₆alkynyl optionally substituted with 1-2 R⁷, C₂-C₆alkenyl optionally substituted with 1-2 R⁷, and C₁-C₆alkyloxy optionally substituted with 1-2 R⁷, and pharmaceutically acceptable salts thereof.

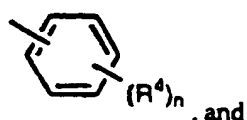
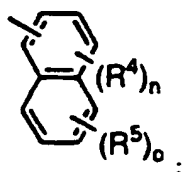
In a further alternative embodiment of this invention the compound is represented by structural formula IV



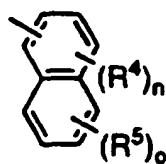
where the symbols of formula IV are defined as follows:

5

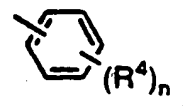
Ar¹ and Ar² are each independently selected from indoyl,



Ar³ is selected from the group



, and



n and o are independently 1, 2 or 3;

Ar^A is selected from the group C₀-C₃alkyl-heterocycle, -O-C₀-C₃alkyl-heterocycle, and
 10 -NR²-C₂-C₆alkyl -heterocycle, where the heterocycle comprises a mono-, bi-, or tricycle containing 5-12 ring atoms, one or two of which are heteroatoms selected from O, S, and N, provided at least one heteroatom is N, where any N atom is optionally substituted with R¹, C₀-C₆alkyl substituted with one or two substituents, O-C₂-C₆alkyl substituted with one or two substituents, and NR²-C₂-C₆alkyl substituted with one or two substituents where the
 15 substituents are selected from the group NR²R³, imidazolyl, pyridinyl, dihydropyridinyl, and piperidinyl;

R^B, R^C, R^D, and R^E are selected from the group hydrogen, C₁-C₆alkyl optionally substituted with a group selected from NR²R³, and phenyl-C₁-C₃-NR²R³, and halo(F, Cl, Br, I)C₁-C₆alkyl;

20 R¹ is selected from hydrogen, C₁-C₆alkyl, C(=O)-C₁-C₆alkyl, C(=O)-hydroxyalkyl, NR²R³, C(=NR²)-NR²R³, C(=O)O-C₁-C₆alkyl, and halo(F, Cl, Br, I)C₁-C₆alkyl;

R^2 and R^3 are independently selected from hydrogen, C_1 - C_6 alkyl, piperidiny, and halo(F, Cl, Br, I) C_1 - C_6 alkyl;

R^2 and R^3 together with the N to which they are bonded may form a 5- or 6-member heterocycle, optionally containing one additional hetero atom selected from O, S, and N where any N is optionally substituted with R^1 , any carbon is optionally substituted with R^6 and where the heterocycle is optionally fused to a phenyl ring, optionally substituted with R^4 ;

R^4 and R^5 are independently selected from the group hydrogen, halo(F, Cl, Br, and I), cyano, amino, amido, nitro, hydroxy, C_1 - C_6 alkyl optionally substituted with 1-3 R^6 , C_2 - C_6 alkynyl optionally substituted with 1-3 R^6 , C_1 - C_6 alkyloxy optionally substituted with 1-3 R^6 , C_1 - C_6 acylamino optionally substituted with 1-3 R^6 , C_1 - C_6 alkylcarbonyl optionally substituted with 1-3 R^6 , C_1 - C_6 alkyloxycarbonyl optionally substituted with 1-3 R^6 , N-(C_1 - C_6 alkyl),N-(C_1 - C_6 acyl)amino optionally substituted with 1-3 R^6 , N-(C_1 - C_6 alkyl)carboxamido optionally substituted with 1-3 R^6 , N,N-di(C_1 - C_6 alkyl)amino optionally substituted with 1-3 R^6 , N,N-di(C_1 - C_6 alkyl)carboxamido optionally substituted with 1-3 R^6 , C_1 - C_4 perfluoroalkyl, and C_1 - C_3 perfluoroalkoxy;

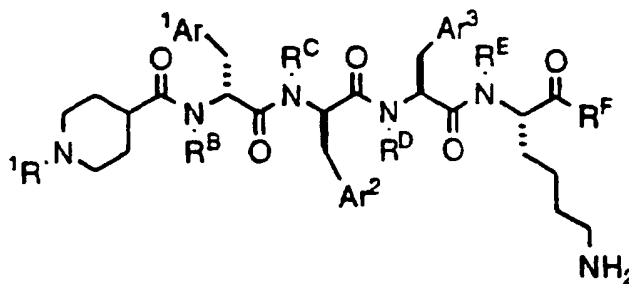
R^6 is selected from the group $COOR^2$, $O(C=O)R^2$, $CONR^2R^3$, cyano, NR^2R^3 , NR^2COR^3 , azido, nitro, and hydroxy;

R^7 is selected from the group R^6 , C_6 - C_{10} aryl optionally substituted with halo(F, Cl, Br, and I), cyano, amino, amido, nitro, hydroxy, C_1 - C_4 perfluoroalkyl, and C_1 - C_3 perfluoroalkoxy;

Z is selected from the group C_1 - C_6 alkyl substituted with 1-2 R^7 , C_2 - C_6 alkynyl optionally substituted with 1-2 R^7 , C_2 - C_6 alkenylyl optionally substituted with 1-2 R^7 , and C_1 - C_6 alkyloxy optionally substituted with 1-2 R^7 ;

Z and R^E together with the N to which they are bonded may form a 5- or 6-member heterocycle, optionally containing one additional hetero atom selected from O, S, and N where any N is optionally substituted with R^1 , any carbon is optionally substituted with R^7 and where the heterocycle is optionally fused to a phenyl ring; and pharmaceutically acceptable salts thereof.

An optional compound of this embodiment is represented by structural Formula (IVa)



IVa

where the symbols in formula IVa are defined as follows:

R^B , R^C , R^D and R^E are selected from the group hydrogen, and C_1 - C_6 alkyl;



Ar^1 and Ar^2 are each independently selected from indoyl, and



Ar^3 is selected from the group

, and

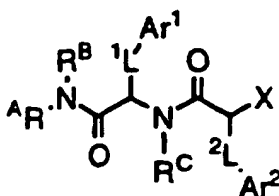
R^F is selected from the group OH, C_1 - C_4 alkoxy, NR^5R^6 , and 1 to 4 α -amino acid residues;

R^4 is selected from hydrogen, halo(F, Cl, Br, and I), cyano, amino, amido, nitro, hydroxy, C_1 - C_4 alkoxy, C_1 - C_4 perfluoroalkyl, and C_1 - C_3 perfluoroalkoxy;

R^5 and R^6 are independently selected from hydrogen, and C_1 - C_6 alkyl; and

pharmaceutically acceptable salts thereof.

In still another embodiment of this invention the compound is referred to as a "retroinverso" of the compound of formula II and is represented by formula V



V

where Ar^1 , Ar^2 , L^1 , L^2 , R^A , R^B , R^C and X are defined above for the compound of formula I.

The invention further comprises a pharmaceutical composition comprising a pharmaceutically acceptable excipient and any of the compounds represented by structural formula I-V. Additionally the invention provides a method for increasing the level of endogenous growth hormone in a mammal comprising administering to the mammal a pharmaceutically effective amount of the forgoing composition to the mammal. The method further comprises administering the composition in combination with a growth factor selected from; growth hormone releasing hormone (GHRH), insulin like growth factor-1 (IGF-1), and insulin like growth factor-2 (IGF-2). In an alternative method of this invention GHRP's represented by formulae I-V, as well as other GHRP's, are used in combination with IGF-1 to

treat diseases in which long term IGF-1 is indicated including but not limited to Type II diabetes.

BRIEF DESCRIPTION OF THE FIGURES

5 **FIGURE 1.** A cartoon showing regulation of growth hormone (GH) release by the "pituitary-hypothalamus axis" and the alternative "GHRP pathway". Some of the stimulatory (+) and inhibitory (-) effects that growth hormone releasing hormone/factor (GHRH/F), growth hormone releasing peptides/peptidomimetics (GHRP's), somatostatin (SS), GH, and insulin-like growth factor 1 (IGF-1) have on selected glands, organs, and tissues are also shown.

10

FIGURE 2. A representative rat "pit" cell assay dose response of GH release over a 15 min. exposure to increasing concentrations of (inip)-bbFK-NH₂. GH release is significantly ($P < 0.05$) increased at 0.3 nM and reaches a plateau by 1 nM with an EC₅₀ of 0.16 nM (see right panel for data points and curve used to calculate EC₅₀). The mean ($n=3$) EC₅₀ for inip-bbFK-NH₂ was 0.18 ± 0.04 nM, over 30-fold more potent than HwAWfK-NH₂ ("GHRP-6") (6.2 ± 1.5 nM; $n=5$).

15

FIGURE 3. Demonstration that (inip)-bbFK-NH₂ acts at the proposed "GHRP receptor". Rat "pit" cell challenges were carried out using combinations of GHRP-6, (inip)-bbFK-NH₂ and GHRH. GHRP-6 and (inip)-bbFK-NH₂ (100 nM) both caused 3-fold increases in GH levels over control (ctrl), but no additional or synergistic increase was observed when used in combination (solid black bars). In contrast, both GHRP-6 and (inip)-bbFK-NH₂ show synergy with 10nM GHRH indicating that neither GHRP-6 nor (inip)-bbFK-NH₂ act via the GHRH receptor (cross hatched bars).

20

25

FIGURE 4. Desensitization effect of the "GHRP receptor" upon challenging rat pituitary cells with three sequential 15 min. incubations with fresh (inip)-bbFK-NH₂. The GH release was decreased after the second 15 min. incubation (total 30 min. exposure to (inip)-bbFK-NH₂) and no significant GH release compared to control occurred during the final challenge with (inip)-bbFK-NH₂. However when GHRH (10 nM) was added to the next 15 min. incubation, a significant GH response occurred, consistent with the separate receptor model.

30

FIGURE 5. Somatostatin suppression of GHRP-stimulated GH release. (Inip)-bbFK-NH₂ at 0.1, 1, 10 and 100 nM significantly elevates GH release (solid black bars). Coincubation of (inip)-bbFK-NH₂ with 20nM somatostatin suppressed this release (crosshatched bars).

35

FIGURE 6. Antagonism of (inip)-bbFK-NH₂ induced GH release in the absence (solid black bars) and presence of 10 μ M of the "GHRP-6" antagonist HwkWfK-NH₂ (crosshatched bars).

FIGURE 7. Specificity for GH release compared to prolactin release induced with (inip)-bbFK-NH₂. GH and prolactin levels in the same media from cells challenged with 100 nM (inip)-bbFK-NH₂. GH release was 3-fold while prolactin levels increased 1.6 fold.

5

FIGURE 8. TSH, FSH and LH release induced with (inip)-bbFK-NH₂. Pituitary cells challenged with (inip)-bbFK-NH₂ had no effect on these hormone concentrations.

FIGURE 9. ACTH release induced with corticotrophin releasing factor (CRF) or (inip)-bbFK-NH₂. ACTH levels rose 3-fold when stimulated with CRF but no significant change was observed with 100 nM (inip)bbFK-NH₂.

FIGURES 10A-10D. Ca⁺⁺ release in pituitary cells by (inip)bbFK-NH₂. Basal Ca⁺⁺ in Indol-1 AM loaded pituitary cells after 4-day monolayer culture (Figure 10A and Figure 10C). Twenty-one (21) seconds after (inip)bbFK-NH₂ addition, the increased Ca⁺⁺ flux in Figure 10D is demonstrated by lighter areas in some of the cells of this heterologous population. Addition of vehicle did not change Ca⁺⁺ levels (Figure 10B).

FIGURE 11. Dose dependent GH release with (inip)bb(feg). GH release by rat pituitary cells to increasing concentrations of (inip)bb(feg) (left panel) over a 15 minute incubation. Right panel shows data points and curve used to calculate the EC₅₀ of 0.09 nM.

FIGURE 12. Demonstration that (inip)bb(feg) acts at the proposed "GHRP receptor". GH response to GHRP-6 (100nM) and (inip)bb(feg) (100 nM) was significantly greater than control but GH release was not synergistic when both were added in combination. GHRH (100 nM) elicited a mild GH response which was synergistic in combination with either GHRP-6 or (inip)bb(feg).

FIGURE 13. Somatostatin suppression of (inip)bb(feg)-stimulated GH release. GH release with 100 nM (inip)bb(feg) was totally suppressed in the presence of 20 nM somatostatin.

FIGURE 14. Desensitization of the "GHRP receptor" upon challenging rat pituitary cells with three sequential 15 min. incubations with fresh (inip)bb(feg). GH release from the same pituitary cells over three sequential 15 minute incubations with (inip)bb(feg) (100 nM). After a total of 45 minutes, GH release was markedly decreased in response to (inip)bb(feg) but these cells were able to release more GH in response to a final 15 minute incubation with GHRH (10 nM).

FIGURE 15. Dose dependent GH release with (inip)b(wol). GH release by rat pituitary cells to increasing concentrations of (inip)b(wol) (left panel) over a 15 minute incubation. Right panel shows the data points and curve used to calculate the EC₅₀ of 3.9 nM.

- 5 **FIGURE 16.** Demonstration that (inip)b(wol) acts at the proposed "GHRP receptor". GH response to GHRP-6 (100nM) and (inip)b(wol) (100 nM) was significantly greater than control but GH release was not synergistic when both were added in combination. GHRH (100 nM) elicited a mild GH response which was synergistic in combination with either GHRP-6 or (inip)b(wol).

10

FIGURE 17. Somatostatin suppression of (inip)b(wol)-stimulated GH release. GH release to 100 nM (inip)b(wol) was totally suppressed in the presence of 20 nM somatostatin.

- 15 **FIGURE 18.** Desensitization effect of the "GHRP receptor" upon challenging rat pituitary cells with three sequential 15 min. incubations with fresh (inip)b(wol). GH release from the same pituitary cells over three sequential 15 minute incubations with (inip)b(wol) (100 nM). After a total of 45 minutes, no significant release of GH was observed in response to (inip)b(wol) but these cells were able to release GH in response to a final 15 minute incubation with GHRH (10 nM).

20

- FIGURE 19.** Body weight gain in normal adult female rats in response to excipient (open squares), three doses of (inip) b b F K-NH₂ (4 µg/d open circles, 20 µg/d open triangles, 100 µg/d half-filled diamonds), or one dose of GHRH (600 µg/d, filled squares); delivery was by subcutaneous minipump infusion for 14 days. There was a dose-related weight gain in response to (inip) b b F K-NH₂, that at 20 µg/d reached the response to GHRH. Means and standard errors are shown.

25

- FIGURE 20.** Body weight gain in normal adult female rats treated for 14 days with excipient (open bar), two doses of (inip) b b F K-NH₂ given subcutaneously by injection (shaded bar, 100 µg/d; solid bar, 20 µg/d) or infusion (lightly hatched bar, 100 µg/d; heavily hatched bar, 20 µg/d). Injections of (inip) b b F K-NH₂ were more effective than infusions. Means and standard errors are shown.

30

- FIGURE 21.** Body weight gain in normal adult female rats treated for 14 days with excipient (open squares), or (inip) b b F K-NH₂ given by subcutaneous injection (100 µg/d; open triangles) or infusion (100 µg/d; open circles). Injections of (inip) b b F K-NH₂ produced a maintained growth response; infusions gave a large initial response that was not maintained. Means and standard errors are shown.

35

FIGURE 22. Body weight gain in lean (open circles) and obese Type II Zucker Diabetic Fatty (ZDF) male rats treated subcutaneously for 24 days with excipient (solid circles), recombinant human growth hormone (rhGH, large open squares, 500 $\mu\text{g/d}$), recombinant human insulin-like growth factor-1 (rhIGF-1, small squares, 758 $\mu\text{g/d}$), (inip) b b F K-NH₂ given by injection (100 $\mu\text{g/d}$; open triangles), the combination of rhGH plus rhIGF-1 (solid squares), or the combination of rhIGF-1 plus (inip) b b F K-NH₂ (solid triangles). The combination of (inip) b b F K-NH₂ plus rhIGF-1 produced a maintained growth response equal to that of rhIGF-1 plus rhGH. Means and standard errors are shown.

10

FIGURE 23. Body weight gain in obese Type II Zucker Diabetic Fatty (ZDF) male rats treated subcutaneously for 7 days with excipient (solid circles), recombinant human insulin-like growth factor-1 (rhIGF-1, open circles, 758 $\mu\text{g/d}$), (inip) b b F K-NH₂ given by injection (100 $\mu\text{g/d}$; open triangles), or the combination of rhIGF-1 and (inip) b b F K-NH₂ (solid triangles). The combination of (inip) b b F K-NH₂ plus rhIGF-1 produced a maintained growth response that was at least additive compared to each agent given alone. Means and standard errors are shown.

FIGURE 24. Basal blood glucose levels obtained weekly in lean (small squares) and obese Type II Zucker Diabetic Fatty (ZDF) male rats treated for 3 weeks subcutaneously with excipient (controls, solid circles), recombinant human growth hormone (rhGH, open squares, 500 $\mu\text{g/d}$), recombinant human insulin-like growth factor-1 (rhIGF-1, open circles, 758 $\mu\text{g/d}$), (inip) b b F K-NH₂ given by injection (100 $\mu\text{g/d}$; open triangles), or the combination of rhGH and rhIGF-1 (solid squares), or the combination of rhIGF-1 and (inip) b b F K-NH₂ (solid triangles). When given alone, or in combination with rhIGF-1, (inip) b b F K-NH₂ had lesser effect on blood glucose (diabetogenic effect) than rhGH at doses with similar somatogenic effects (Fig. 22). Means and standard errors are shown.

FIGURE 25. Blood glucose concentrations following an intravenous insulin challenge (insulin tolerance test) in lean control male rats (open bar), obese Type II Zucker Diabetic Fatty (ZDF) rats treated subcutaneously with excipient (solid bar), recombinant human growth hormone (rhGH, light left-right ascending hatching, 500 $\mu\text{g/d}$), recombinant human insulin-like growth factor-1 (rhIGF-1, light shaded bar, 758 $\mu\text{g/d}$), (inip) b b F K-NH₂ given by injection (100 $\mu\text{g/d}$; light left-right descending hatching), or the combination of rhGH and rhIGF-1 (heavy left-right ascending hatching), or the combination of rhIGF-1 and (inip) b b F K-NH₂ (heavy left-right descending hatching). When given alone, or in combination with rhIGF-1, (inip) b b F K-NH₂ had a greatly reduced effect on insulin sensitivity (diabetogenic effect) than rhGH at doses with similar somatogenic effects (Fig. 22). Means and standard errors are shown.

FIGURE 26. Daily body weight gains in normal adult female rats treated for 7 days with twice daily subcutaneous injections of excipient (open squares), growth hormone releasing hormone (open triangles), H w A W f K (solid squares), (inip) b b F -NH₂ (open circles), (inip) b nmb bam (solid circles), or L-692,585 (solid triangles). Significant weight gain occurred after treatment with all molecules except L-692,585. Means and standard errors are shown.

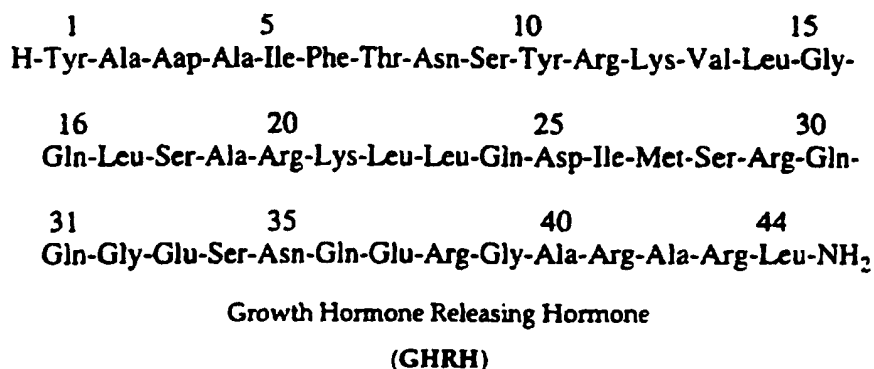
FIGURE 27. Total body weight gain in normal adult rats treated for 7 days with twice daily subcutaneous injections of excipient (solid bar), infusions of recombinant human insulin-like growth factor-1 (rhIGF-1, broad hatching), or the combinations of rhIGF-1 plus growth hormone releasing hormone (light shading), H w A W f K plus rhIGF-1 (narrow hatching), (inip) b b F -NH₂ plus rhIGF-1 (open bar), (inip) b nmb bam plus rhIGF-1 (dark shading), or L-692,585 plus rhIGF-1 (horizontal lines). Weight gains tended to be greater after combination treatment. Means and standard errors are shown.

DETAILED DESCRIPTION OF THE INVENTION

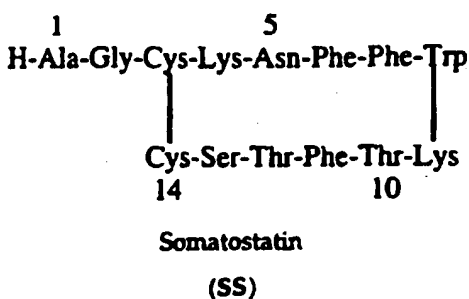
A. Definitions

Terms used in the claims and specification are defined as set forth below unless otherwise specified.

The terms growth hormone releasing hormone (GHRH) or factor (GHRF/GRF) are used interchangeably and refer to the endogenous hypothalamic GH secretagogue, from any species, having the capability of binding to the pituitary somatotroph and inducing a rapid dose-dependent release of GH and biologically active analogs thereof. Included in this definition are; GHRH(1-44), GHRH(1-43), GHRH(1-40), and GHRH(1-29). Other examples of GHRH analogs are described in U. S. patent no. 4,622,312.



The term somatostatin (SS) refers to the inhibitory hypothalamic tetradecapeptide capable of antagonizing in a dose-dependent manner the GH-releasing effect of GHRH.



As used herein, "IGF-1" refers to insulin-like growth factor from any species, including bovine, ovine, porcine, equine, avian, and preferably human, in native-sequence or in variant form, and from any source, whether natural, synthetic, or recombinant. Preferred herein for animal use is that form of IGF-1 from the particular species being treated, such as porcine IGF-1 to treat pigs, ovine IGF-1 to treat sheep, bovine IGF-1 to treat cattle, etc. Preferred herein for human use is human native-sequence, mature IGF-1, more preferably without a N-terminal methionine, prepared, for example, by the process described in EP 230,869 published Aug. 5, 1987; EP 128,733 published Dec. 19, 1984; or EP 288,451 published Oct. 26, 1988. More preferably, this native sequence IGF-1 is recombinantly produced and is available from Genentech, Inc., South San Francisco, Calif. for clinical investigations. Also preferred for use is IGF-1 that has a specific activity greater than about 14,000 units/mg as determined by radioreceptor assay using placenta membranes, such as that available from KabiGen AB, Stockholm, Sweden.

The most preferred IGF-1 variants are those described in U.S. Pat. No. 5,077,276 issued Dec. 31, 1991, in PCT WO 87/01038 published Feb. 26, 1987 and in PCT WO 89/05822 published Jun. 29, 1989, i.e., those wherein at least the glutamic acid residue is absent at position 3 from the N-terminus of the mature molecule or those having a deletion of up to five amino acids at the N-terminus. The most preferred variant has the first three amino acids from the N-terminus deleted (variously designated as brain IGF, tIGF-1, des(1-3) IGF-1, or des-IGF-1).

The term "GHRP" as used herein refers to compounds that cause release of endogenous GH in a dose-dependent manner, where such release is synergized by GHRH but not by other GHRP's such as GHRP-6, and where such release causes a desensitization after continuous exposure to the GHRP while maintaining the ability to respond to GHRH.

The term " C_n-C_m alkyl" means a cyclic or linear, branched or unbranched, saturated aliphatic hydrocarbon radical, having the number of carbon atoms specified, where m and n are zero or integers identifying the range of carbon atoms contained in the alkyl group. When n is zero (0) the term becomes a chemical bond, usually a covalent bond. Examples of alkyl radicals include methyl, ethyl, n-propyl, isopropyl(iPr), n-butyl, iso-butyl, sec-butyl, tert-butyl(tBu), n-

pentyl, 2-methylbutyl, 2,2-dimethylpropyl, n-hexyl, 2-methylpentyl, 2,2-dimethylbutyl, n-heptyl, 2-methylhexyl, cyclohexyl, and the like. The terms "lower alkyl" and "C₁-C₆alkyl" are synonymous and used interchangeably.

The term "C₂-C_malkenyl" means a cyclic or linear, branched or unbranched hydrocarbon radical containing at least one carbon-carbon double bond, having the number of carbon atoms specified, each double bond being independently cis, trans, E or Z, or a non-geometric isomer.

The term "C₂-C_malkynyl" means a cyclic or linear, branched or unbranched hydrocarbon radical containing at least one carbon-carbon triple bond, having the number of carbon atoms specified,

The terms "C₁-C₁₂acyloxy" or "C₁-C₁₂alkanoyloxy" are used interchangeably and denote herein groups of the formula C₀-C₁₂alkyl-C(=O)-O- such as; formyloxy, acetoxy, propionyloxy, butyryloxy, pentanoyloxy, hexanoyloxy, heptanoyloxy, and the like.

The term "N,N-di(C₀-C₆)alkylamino" denotes herein groups of the formula (C₀-C₆alkyl)₂-N- where both, one or none of the hydrogen atoms of H₂N- are substituted with C₁-C₆alkyl.

The term "N-(C₁-C₆alkyl),N-(C₁-C₆acyl)amino" denotes herein an amino group where one hydrogen is substituted with a C₁-C₆alkyl group and the other hydrogen is substituted with a C₁-C₆acyl group.

The terms "C₁-C₆alkyloxycarbonyl" and "C₁-C₆carboalkoxy" are used interchangeably herein and denote groups of the formula C₁-C₆alkyl-O-C(=O)-.

The terms "N-(C₁-C₆alkyl)carboxamido" and "N-(C₁-C₆alkyl)-aminocarbonyl" are used interchangeably herein and denote groups of the formula C₁-C₆alkyl-NH-C(=O)-.

The terms "C₁-C₁₂alkylcarbonyl", "C₁-C₁₂alkanoyl" and "C₁-C₁₂acyl" are used interchangeably herein and denote groups of the formula C₀-C₁₂alkyl-C(=O)- and encompass groups such as formyl, acetyl, propionyl, butyryl, pentanoyl, hexanoyl, heptanoyl, benzoyl and the like.

The term "C₁-C₆acylamino" denotes groups of the formula C₁-C₆alkyl-C(=O)-NH-

The terms "C₁-C₁₂alkyloxy" and "C₁-C₁₂ substituted alkyloxy" denote C₁-C₁₂alkyl and C₁-C₁₂ substituted alkyl groups, respectively, attached to an oxygen which is in turn the point of attachment for the alkyloxy or substituted alkyloxy group to the group or substituent designated (e.g. C₁-C₁₂alkyl-O-). These include groups such as methoxy, ethoxy, n-propoxy, isopropoxy, n-butoxy, t-butoxy, cyclohexyloxy and like groups.

The term "aryl" when used alone means a homocyclic hydrocarbon aromatic radical, whether or not fused, having the number of carbon atoms designated or if none are designated - from 6 to 14. Aromatic radicals may be mononuclear or polynuclear. Examples of aryl groups include phenyl, naphthyl, anthranyl, phenanthranyl, azulyl and the like.

Preferred aryl groups include phenyl, naphthyl, biphenyl, phenanthrenyl, naphthacenyl, and the like (see e.g. *Lang's Handbook of Chemistry* (Dean, J. A., ed) 13th ed. Table 7-2 [1985]).

Optionally the "aryl" is substituted with one or more substituents usually designated by a group "-Rⁿ", where n is any integer. Examples of substituted phenyl groups include

5 mono- or di(halo)phenyl groups such as 4-chlorophenyl, 2,6-dichlorophenyl, 2,5-dichlorophenyl, 3,4-dichlorophenyl, 3-chlorophenyl, 3-bromophenyl, 4-bromophenyl, 3,4-dibromophenyl, 3-chloro-4-fluorophenyl, 2-fluorophenyl and the like; mono- or di(hydroxy)phenyl groups such as 4-hydroxyphenyl, 3-hydroxyphenyl, 2,4-dihydroxyphenyl, the protected-hydroxy derivatives thereof and the like; nitrophenyl groups such as 3- or 4-

10 nitrophenyl; cyanophenyl groups, for example, 4-cyanophenyl; mono- or di(lower alkyl)phenyl groups such as 4-methylphenyl, 2,4-dimethylphenyl, 2-methylphenyl, 4-(iso-propyl)phenyl, 4-ethylphenyl, 3-(n-propyl)phenyl and the like; mono or di(alkoxy)phenyl groups, for example, 2,6-dimethoxyphenyl, 4-methoxyphenyl, 3-ethoxyphenyl, 4-(isopropoxy)phenyl, 4-(t-butoxy)phenyl, 3-ethoxy-4-methoxyphenyl and the like; 3- or 4- trifluoromethylphenyl; mono-

15 or dicarboxyphenyl or (protected carboxy)phenyl groups such as 4-carboxyphenyl; mono- or di(hydroxymethyl)phenyl or (protected hydroxymethyl)phenyl groups such as 3-(protected hydroxymethyl)phenyl or 3,4-di(hydroxymethyl)phenyl, 2,3- and 3,4-methylene dioxy; mono- or di(aminomethyl)phenyl or (protected aminomethyl)phenyl groups such as 2-(aminomethyl)phenyl or 2,4-(protected aminomethyl)phenyl; or mono- or di(N-

20 (methylsulfonylamino))phenyl groups such as 3-(N-methylsulfonylamino))-phenyl. Also, the term "substituted phenyl" represents disubstituted phenyl groups wherein the substituents are different, for example, 3-methyl-4-hydroxyphenyl, 3-chloro-4-hydroxyphenyl, 2-methoxy-4-bromophenyl, 4-ethyl-2-hydroxyphenyl, 3-hydroxy-4-nitrophenyl, 2-hydroxy-4-chlorophenyl and the like. Preferred substituted phenyl groups include the 2- and 3-trifluoromethylphenyl,

25 4-fluoro or chlorophenyl the 4-hydroxyphenyl, the 2-aminomethylphenyl and the 3-(N-(methylsulfonylamino))phenyl groups.

The term "arylalkyl" means one, two, or three aryl groups having the number of carbon atoms designated, appended to an alkyl radical having the number of carbon atoms designated including but not limited to; benzyl, naphthylmethyl, phenethyl, benzyhydryl (diphenylmethyl),

30 trityl, and the like. A preferred arylalkyl group is the benzyl group.

The term "substituted C₆-C₁₂aryl-C₁-C₆alkyl" denotes a C₁-C₆alkyl group substituted at any carbon with a C₆-C₁₂aryl group bonded to the alkyl group through any aryl ring position and substituted on the C₁-C₆alkyl portion with one, two or three groups chosen from halogen(F, Cl, Br, I), hydroxy, protected hydroxy, amino, protected amino, C₁-C₆acyloxy, nitro,

35 carboxy, protected carboxy, carbamoyl, carbamoyloxy, cyano, C₁-C₆alkylthio, N-(methylsulfonylamino) C₁-C₆alkoxy, or other groups specified. Optionally, the aryl group may be substituted with one, two, or three groups chosen from halogen(especially F), cyano, hydroxy, protected hydroxy, nitro, C₁-C₆alkyl, C₁-C₄alkoxy, carboxy, protected carboxy,

carboxymethyl, protected carboxymethyl, hydroxymethyl, protected hydroxymethyl, aminomethyl, protected aminomethyl, or an N-(methylsulfonylamino) group. As before, when either the C₁-C₆alkyl portion or the aryl portion or both are disubstituted, the substituents can be the same or different.

5 Examples of the term "substituted C₆-C₁₀aryl-C₁-C₆alkyl" include groups such as 2-phenyl-1-chloroethyl, 2-(4-methoxyphenyl)ethyl, 2,6-dihydroxy-4-phenyl(n-hexyl), 5-cyano-3-methoxy-2-phenyl(n-pentyl), 3-(2,6-dimethyl-phenyl)n-propyl, 4-chloro-3-aminobenzyl, 6-(4-methoxyphenyl)-3-carboxy(n-hexyl), 5-(4-aminomethyl phenyl)-3-(aminomethyl)(n-pentyl), and the like.

10 Unless otherwise specified, the terms "heterocycle", "heterocyclic group", "heterocyclic" or "heterocyclyl" are used interchangeably herein and refer to any mono-, bi-, or tricyclic saturated, unsaturated, or aromatic ring having the number of ring atoms designated where at least one ring is a 5-, 6- or 7-membered hydrocarbon ring containing a designated number of heteroatoms selected from nitrogen, oxygen, and sulfur, preferably at least one heteroatom is
15 nitrogen (*Lang's Handbook of Chemistry, supra*). The heterocycle is a 5- or 6-member saturated, unsaturated, or aromatic hydrocarbon ring usually containing 1, 2, or 3 heteroatoms, preferably 1 or 2, selected from O, N, and S. Typically, the 5-membered ring has 0 to 2 double bonds and the 6- or 7-membered ring has 0 to 3 double bonds and the nitrogen or sulfur heteroatoms may
20 optionally be oxidized, and any nitrogen heteroatom may optionally be substituted or quarternized. Included in the definition are any bicyclic groups where any of the above heterocyclic rings are fused to a benzene ring. Heterocyclics in which nitrogen is the heteroatom are preferred.

 The following ring systems are examples of the heterocyclic (whether substituted or unsubstituted) radicals denoted by the term "heterocyclic": thienyl, furyl, pyrrolyl, imidazolyl,
25 pyrazolyl, thiazolyl, isothiazolyl, oxazolyl, isoxazolyl, triazolyl, thiadiazolyl, oxadiazolyl, tetrazolyl, thiatriazolyl, oxatriazolyl, pyridyl, pyrimidyl, pyrazinyl, pyridazinyl, thiazinyl, oxazinyl, triazinyl, thiadiazinyl, oxadiazinyl, dithiazinyl, dioxazinyl, oxathiazinyl, tetrazinyl, thiatriazinyl, oxatriazinyl, dithiadiazinyl, imidazoliny, dihydropyrimidyl, tetrahydro-
30 pyrimidyl, tetrazolo[1,5-b]pyridazinyl and purinyl, as well as benzo-fused derivatives, for example benzoxazolyl, benzthiazolyl, benzimidazolyl and indolyl.

 Heterocyclic 5-membered ring systems containing a sulfur or oxygen atom and one to three nitrogen atoms are also suitable for use in the instant invention. Examples of such preferred groups include thiazolyl, in particular thiazol-2-yl and thiazol-2-yl N-oxide, thiadiazolyl, in particular 1,3,4-thiadiazol-5-yl and 1,2,4-thiadiazol-5-yl, oxazolyl, preferably
35 oxazol-2-yl, and oxadiazolyl, such as 1,3,4-oxadiazol-5-yl, and 1,2,4-oxadiazol-5-yl. A group of further preferred examples of 5-membered ring systems with 2 to 4 nitrogen atoms include imidazolyl, preferably imidazol-2-yl; triazolyl, preferably 1,3,4-triazol-5-yl; 1,2,3-triazol-5-yl,

1,2,4-triazol-5-yl, and tetrazolyl, preferably 1H-tetrazol-5-yl. A preferred group of examples of benzo-fused derivatives are benzoxazol-2-yl, benzthiazol-2-yl and benzimidazol-2-yl.

Further suitable specific examples of the above heterocyclic ring systems are 6-membered ring systems containing one to three nitrogen atoms. Such examples include
 5 pyridyl, such as pyrid-2-yl, pyrid-3-yl, and pyrid-4-yl; pyrimidyl, preferably pyrimid-2-yl and pyrimid-4-yl; triazinyl, preferably 1,3,4-triazin-2-yl and 1,3,5-triazin-4-yl; pyridazinyl, in particular pyridazin-3-yl, and pyrazinyl. The pyridine N-oxides and pyridazine N-oxides and the pyridyl, pyrimid-2-yl, pyrimid-4-yl, pyridazinyl and the 1,3,4-triazin-2-yl radicals, are a preferred group. Optionally preferred 6-membered ring heterocycles are;
 10 piperazin-2-yl, piperidyl, piperid-2-yl, piperid-3-yl, piperid-4-yl, morpholino, morpholin-2-yl, and morpholin-3-yl.

An optional group of "heterocyclics" include; 1,3-thiazol-2-yl, 4-(carboxymethyl)-5-methyl-1,3-thiazol-2-yl, 4-(carboxymethyl)-5-methyl-1,3-thiazol-2-yl sodium salt, 1,2,4-thiadiazol-5-yl, 3-methyl-1,2,4-thiadiazol-5-yl, 1,3,4-triazol-5-yl, 2-methyl-1,3,4-triazol-5-yl, 2-
 15 hydroxy-1,3,4-triazol-5-yl, 2-carboxy-4-methyl-1,3,4-triazol-5-yl sodium salt, 2-carboxy-4-methyl-1,3,4-triazol-5-yl, 1,3-oxazol-2-yl, 1,3,4-oxadiazol-5-yl, 2-methyl-1,3,4-oxadiazol-5-yl, 2-(hydroxymethyl)-1,3,4-oxadiazol-5-yl, 1,2,4-oxadiazol-5-yl, 1,3,4-thiadiazol-5-yl, 2-thiol-1,3,4-thiadiazol-5-yl, 2-(methylthio)-1,3,4-thiadiazol-5-yl, 2-amino-1,3,4-thiadiazol-5-yl, 1H-tetrazol-5-yl, 1-methyl-1H-tetrazol-5-yl, 1-(1-(dimethylamino)eth-2-yl)-1H-tetrazol-5-yl, 1-
 20 (carboxymethyl)-1H-tetrazol-5-yl, 1-(carboxymethyl)-1H-tetrazol-5-yl sodium salt, 1-(methylsulfonic acid)-1H-tetrazol-5-yl, 1-(methylsulfonic acid)-1H-tetrazol-5-yl sodium salt, 2-methyl-1H-tetrazol-5-yl, 1,2,3-triazol-5-yl, 1-methyl-1,2,3-triazol-5-yl, 2-methyl-1,2,3-triazol-5-yl, 4-methyl-1,2,3-triazol-5-yl, pyrid-2-yl N-oxide, 6-methoxy-2-(n-oxide)-pyridaz-3-yl, 6-hydroxypyridaz-3-yl, 1-methylpyrid-2-yl, 1-methylpyrid-4-yl, 2-hydroxypyrimid-4-yl, 1,4,5,6-
 25 tetrahydro-5,6-dioxo-4-methyl-as-triazin-3-yl, 1,4,5,6-tetrahydro-4-(formylmethyl)-5,6-dioxo-as-triazin-3-yl, 2,5-dihydro-5-oxo-6-hydroxy-as-triazin-3-yl, 2,5-dihydro-5-oxo-6-hydroxy-as-triazin-3-yl sodium salt, 2,5-dihydro-5-oxo-6-hydroxy-2-methyl-as-triazin-3-yl sodium salt, 2,5-dihydro-5-oxo-6-hydroxy-2-methyl-as-triazin-3-yl, 2,5-dihydro-5-oxo-6-methoxy-2-methyl-as-triazin-3-yl, 2,5-dihydro-5-oxo-as-triazin-3-yl, 2,5-dihydro-5-oxo-2-methyl-as-triazin-3-yl, 2,5-
 30 dihydro-5-oxo-2,6-dimethyl-as-triazin-3-yl, tetrazolo[1,5-b]pyridazin-6-yl and 8-aminotetrazolo[1,5-b]pyridazin-6-yl.

An alternative group of "heterocyclics" includes; 4-(carboxymethyl)-5-methyl-1,3-thiazol-2-yl, 4-(carboxymethyl)-5-methyl-1,3-thiazol-2-yl sodium salt, 1,3,4-triazol-5-yl, 2-methyl-1,3,4-triazol-5-yl, 1H-tetrazol-5-yl, 1-methyl-1H-tetrazol-5-yl, 1-(1-(dimethylamino)eth-
 35 2-yl)-1H-tetrazol-5-yl, 1-(carboxymethyl)-1H-tetrazol-5-yl, 1-(carboxymethyl)-1H-tetrazol-5-yl sodium salt, 1-(methylsulfonic acid)-1H-tetrazol-5-yl, 1-(methylsulfonic acid)-1H-tetrazol-5-yl sodium salt, 1,2,3-triazol-5-yl, 1,4,5,6-tetrahydro-5,6-dioxo-4-methyl-as-triazin-3-yl, 1,4,5,6-tetrahydro-4-(2-formylmethyl)-5,6-dioxo-as-triazin-3-yl, 2,5-dihydro-5-oxo-6-hydroxy-2-

methyl-as-triazin-3-yl sodium salt, 2,5-dihydro-5-oxo-6-hydroxy-2-methyl-as-triazin-3-yl, tetrazolo[1,5-b]pyridazin-6-yl, and 8-aminotetrazolo[1,5-b]pyridazin-6-yl.

The terms "heteroaryl group" or "heteroaryl" are used interchangeably herein and refer to any mono-, bi-, or tricyclic aromatic rings having the number of ring atoms designated where at least one ring is a 5-, 6- or 7-membered hydrocarbon ring containing from one to four heteroatoms selected from nitrogen, oxygen, and sulfur, preferably at least one heteroatom is nitrogen. The aryl portion of the term "heteroaryl" refers to aromaticity, a term known to those skilled in the art and defined in greater detail in *Advanced Organic Chemistry* J. March, 3rd ed., pages 37-69, John Wiley & Sons, New York (1985).

"Optical isomers", "diastereomers", and "geometric isomers" of some of the compounds represented by the formulae described herein are comprehended to be within the scope of the instant invention, as well as racemic and resolved enantiomerically pure forms and pharmaceutically acceptable salts thereof.

"Pharmaceutically acceptable salts" include both acid and base addition salts.

"Pharmaceutically acceptable acid addition salt" refers to those salts which retain the biological effectiveness and properties of the free bases and which are not biologically or otherwise undesirable, formed with inorganic acids including but not limited to hydrochloric acid, hydrobromic acid, sulfuric acid, sulfamic nitric acid, phosphoric acid and the like, and organic acids such as acetic acid, propionic acid, glycolic acid, pyruvic acid, lactic acid, oxalic acid, maleic acid, malic acid, malonic acid, succinic acid, fumaric acid, tartaric acid, citric acid, stearic acid, ascorbic acid, benzoic acid, cinnamic acid, mandelic acid, methanesulfonic acid, ethanesulfonic acid, isethionic acid, p-toluenesulfonic acid, salicylic acid, naturally occurring amino acids and the like.

"Pharmaceutically acceptable base addition salts" include those derived from inorganic bases such as sodium, potassium, lithium, ammonium, calcium, magnesium, iron, zinc, copper, manganese, aluminum salts and the like. Particularly preferred are the ammonium, potassium, sodium, calcium and magnesium salts. Salts derived from pharmaceutically acceptable organic nontoxic bases includes salts of primary, secondary, and tertiary amines, substituted amines including naturally occurring substituted amines, cyclic amines and basic ion exchange resins, such as isopropylamine, trimethylamine, diethylamine, triethylamine, tripropylamine, ethanolamine, 2-diethylaminoethanol, trimethylamine, dicyclohexylamine, lysine, arginine, histidine, caffeine, procaine, hydrabamine, choline, betaine, ethylenediamine, glucosamine, methylglucamine, theobromine, purines, piperazine, piperidine, N-ethylpiperidine, polyamine resins and the like. Particularly preferred organic non-toxic bases are isopropylamine, diethylamine, ethanolamine, trimethylamine, dicyclohexylamine, choline, and caffeine.

In general, unless otherwise specified, the abbreviations used for the designation of amino acids and the protective groups used therefor are based on recommendations of the IUPAC-IUB Commission of Biochemical Nomenclature (*Biochemistry*, 11:1726-1732 (1972)).

Table 1 provides a list of commonly used symbols or abbreviations (abbr.) used to describe the compounds of this invention.

Table I

5	ABBR	(SMILES) STRUCTURE	R/S	COMMON NAME OF COMPOUND
	A	<chem>NC(C)C(=O)</chem>	S	L-Alanine
	a	<chem>NC(C)C(=O)</chem>	R	D-Alanine
	Ab	<chem>NCCCC(=O)</chem>		4-aminobutyric acid
	Ac	<chem>C(C=O)</chem>		Acetyl
10	Abx	<chem>NC2(CCCCC2)C(=O)</chem>		1-amino-1-cyclohexanecarboxylic acid
	Ahp	<chem>NCCCCCCC(=O)</chem>		7-aminoheptanoic acid
	Ahx	<chem>NCCCCCC(=O)</chem>		6-aminohexanoic acid
	amb	<chem>NCc4ccc(cc4)C(=O)</chem>		p-aminomethylbenzoic acid
15	amF	<chem>NC(C)(Cc1ccccc1)C(=O)</chem>	S	alpha-methyl-L-Phenylalanine
	apc	<chem>N3CCC(N)CC3</chem>		1-(4-amino-piperidine)
	api	<chem>NC3CCNCC3</chem>		4-(4-amino-piperidine)
	Ava	<chem>NCCCCC(=O)</chem>		5-aminovaleric acid
	B	<chem>NC(Cc1cc2ccccc2cc1)C(=O)</chem>	S	L-beta-naphthylalanine
20	b	<chem>NC(Cc1cc2ccccc2cc1)C(=O)</chem>	R	D-beta-naphthylalanine
	bA	<chem>NCCC(=O)</chem>		beta-Alanine
	bam	<chem>NCCCCN</chem>		1,4-butanediamine
	Bmn	<chem>NC(Cc1cc2ccccc2cc1)CN</chem>	S	L-2-amino-3-(2-naphthyl)propylamine
	bmn	<chem>NC(Cc1cc2ccccc2cc1)CN</chem>	R	D-2-amino-3-(2-naphthyl)propylamine
25	BOC	<chem>C(C)(C)OC(=O)</chem>		tert-Butoxycarbonyl
	Bol	<chem>NC(Cc1cc2ccccc2cc1)CO</chem>	S	L-beta-naphthylalanol
	bol	<chem>NC(Cc1cc2ccccc2cc1)CO</chem>	R	D-beta-naphthylalanol
	chA	<chem>NC(CC1CCCCC1)C(=O)</chem>		3-cyclohexylalanine
30	chl	<chem>NC(Cc1ccc(Cl)cc1)C(=O)</chem>	R	D-4-chloro-Phenylalanine
	cho	<chem>C4CCC(CC4)C(=O)</chem>		cyclohexane carboxylic acid
	cx	<chem>NC3CCC(N)CC3</chem>		trans-1,4-cyclohexane diamine
	Cxp	<chem>N4CCN(CC4)C(=O)</chem>		1-carboxypiperazine
	dam	<chem>N(C)C</chem>		N,N-dimethylamine
35	F	<chem>NC(Cc1ccccc1)C(=O)</chem>	S	L-Phenylalanine
	f	<chem>NC(Cc1ccccc1)C(=O)</chem>	R	D-Phenylalanine
	fbd	<chem>N(CCCc1ccccc1)CCCCN</chem>		N-(2-phenylethyl) butane diamine
	feb	<chem>N(CCCc1ccccc1)CCC(=O)</chem>		N-(2-phenylethyl) beta-Alanine

Table I (cont.)

	ABBR	(SMILES) STRUCTURE	R/S	COMMON NAME OF COMPOUND
	feg	<chem>N(Cc1ccccc1)CC(=O)</chem>		N-(2-phenylethyl) Glycine
	fem	<chem>NCCc1ccccc1</chem>		1-phenethyl amine
5	G	<chem>NCC(=O)</chem>		Glycine
	H	<chem>NC(Cc1[nH]cnc1)C(=O)</chem>	S	L-Histadine
	h	<chem>NC(Cc1[nH]cnc1)C(=O)</chem>	R	D-Histadine
	hcF	<chem>NC(Cc1ccccc1)CC(=O)</chem>	S	3-(S)-benzyl-beta-alanine
10	hF	<chem>NC(Cc1ccccc1)C(=O)</chem>	S	homo- L-Phenylalanine
	hf	<chem>NC(Cc1ccccc1)C(=O)</chem>	R	homo-D-Phenylalanine
	inip	<chem>N4CCC(CC4)C(=O)</chem>		isonipecotic acid
	isog	<chem>N4CC=C(CC4)C(=O)</chem>		isoguvacine
	K	<chem>NC(CCCCN)C(=O)</chem>	S	L-Lysine
15	k	<chem>NC(CCCCN)C(=O)</chem>	R	D-Lysine
	mab	<chem>N(C)CCCC(=O)</chem>		N-methyl-4-aminobutyric acid
	mam	<chem>NC</chem>		methylamine
	man	<chem>NCCc1cc2ccccc2cc1</chem>		N-(2-naphthylmethyl)amine
20	mBm	<chem>N(C)C(Cc1cc2ccccc2cc1)CN</chem>	S	N-methyl-[2-amino-3-(2-naphthyl)propyl]amine
	mbm	<chem>N(C)C(Cc1cc2ccccc2cc1)CN</chem>	R	N-methyl-[2-amino-3-(2-naphthyl)propyl]amine
	men	<chem>N(C)CCc1cc2ccccc2cc1</chem>		N-Methyl-2-(2-naphthylethylamine)
25	miz	<chem>N(C)C(Cc1cc2ccccc2cc1)CN(=N)N</chem>	R	2(R)-2-(N-Methylamino)-1-azido-3-(2-naphthyl)propane
	mor	<chem>N1CCOCC1</chem>		morpholine
	N	<chem>NC(CC(=O)N)C(=O)</chem>	S	L-Asparagine
	n	<chem>NC(CC(=O)N)C(=O)</chem>	R	D-Asparagine
30	nba	<chem>NC(C)(C)CC(=O)</chem>		3,3-dimethyl-3-aminopropionic acid
	nboI	<chem>N(C)C(Cc1cc2ccccc2cc1)CO</chem>	S	N-Methyl-D-beta-naphthylalanol
	nL	<chem>NC(CCCC)C(=O)</chem>	S	L-Norleucine
	nl	<chem>NC(CCCC)C(=O)</chem>	R	D-Norleucine
	nmb	<chem>N(C)C(Cc1cc2ccccc2cc1)C(=O)</chem>	S	N-Methyl-L-beta-naphthylalanine
35	nmb	<chem>N(C)C(Cc1cc2ccccc2cc1)C(=O)</chem>	R	N-Methyl-D-beta-naphthylalanine
	nmF	<chem>N(C)C(Cc1ccccc1)C(=O)</chem>	S	N-Methyl-L-Phenylalanine
	nmK	<chem>N(C)C(CCCCN)C(=O)</chem>	S	N-Methyl-L-Lysine
	npA	<chem>NC(Cc1c2ccccc2ccc1)C(=O)</chem>	S	L-alpha-Naphthylalanine

Table I (cont.)

	ABBR	(SMILES) STRUCTURE	R/S	COMMON NAME OF COMPOUND
	npa	<chem>NC(Cc1c2ccccc2ccc1)C(=O)</chem>	R	D-alpha-Naphthylalanine
5	npe	<chem>NCCc1cc2ccccc2cc1</chem>		2-(2-naphthyl)ethylamine
	O	<chem>NC(CCCN)C(=O)</chem>	S	L-Omithine
	o	<chem>NC(CCCN)C(=O)</chem>	R	D-Omithine
	P	<chem>N1C(CCC1)C(=O)</chem>	S	L-Proline
	p	<chem>N1C(CCC1)C(=O)</chem>	R	D-Proline
10	pac	<chem>n4ccc(cc4)CC(=O)</chem>		Pyridine-4-acetic acid
	pam	<chem>NCCCCCN</chem>		1,5-pentanediamine
	Pg	<chem>NC(c1ccccc1)C(=O)</chem>	S	L-Phenylglycine
	pG	<chem>NC(c1ccccc1)C(=O)</chem>	R	D-Phenylglycine
	Pol	<chem>NC(Cc1ccccc1)CO</chem>	S	L-Phenylalanol
15	ppc	<chem>N1CCC(CC1)C(=O)</chem>		Piperidine-4-carboxylic acid
	ppz	<chem>N3CCNCC3</chem>		piperazine
	pyc	<chem>n4ccc(cc4)C(=O)</chem>		Pyridine-4-carboxylic acid
	ram	<chem>NCCCN</chem>		1,3-propane diamine
20	S	<chem>NC(CO)C(=O)</chem>	S	L-Serine
	tam	<chem>NCCN</chem>		1,2-ethane diamine
	tbm	<chem>NC(C)(C)C</chem>		tert-butylamine
	tic	<chem>N7C(Cc8ccccc8C7)C(=O)</chem>	S	D-tetrahydroisoquinoline
	Tic	<chem>N7C(Cc8ccccc8C7)C(=O)</chem>	R	L-tetrahydroisoquinoline
25	W	<chem>NC(Cc1[nH]c2ccccc2c1)C(=O)</chem>	S	L-Tryptophane
	w	<chem>NC(Cc1[nH]c2ccccc2c1)C(=O)</chem>	R	D-Tryptophane
	wol	<chem>NC(Cc1[nH]c2ccccc2c1)CO</chem>	R	D-Tryptophanol
	Y	<chem>NC(Cc1ccc(O)cc1)C(=O)</chem>	S	L-Tyrosine
	y	<chem>NC(Cc1ccc(O)cc1)C(=O)</chem>	R	D-Tyrosine
30	Y(I)	<chem>NC(Cc1ccc(O)c(I)c1)C(=O)</chem>	S	3-Iodo-L-Tyrosine

Notes:

- a) The above structures are depicted in the SMILES format ("SMILES, 1. Introduction to Encoding Rules" Weinenger, D. J. Chem. Inf. Comput. Sci. 1988, 28, 31.). They are generally written N- to C-terminal with the points of attachment at the left- and/or right hand atoms depending on sequence position. In cases where attachment would be ambiguous, two different acronyms are used to depict the two modes, if both are used.
- b) It will be understood that when attached at a terminal position, the appropriate C-terminal function indicated in the table (i.e. -OH, -NH₂, OMe) is added to complete the structure (acid,

amide, Me ester, respectively). Also, hydrogen atoms are to be added to the terminal amine functions to fill out the valence.

B. Utility

The compounds of Formula I can be administered to mammals, including man, to release endogenous growth hormone *in vivo*. For example, the compounds can be administered to commercially important mammals such as swine, cattle, sheep and the like to accelerate and increase their rate and extent of growth and the efficiency of their conversion of feed into body tissue, and to increase milk production in such mammals. In addition, these compounds can be administered to humans *in vivo* as a diagnostic tool to determine whether the pituitary is capable of releasing growth hormone. The compounds of Formula I can be administered *in vivo* to adults and children to stimulate growth hormone release.

Accordingly, the present invention includes within its scope pharmaceutical compositions comprising, as an active ingredient, at least one of the compounds of Formula I in association with a pharmaceutical carrier or diluent. Optionally, the active ingredient of pharmaceutical compositions can comprise a growth promoting agent in addition to at least one of the compounds of Formula I.

Growth promoting agents include but are not limited to; TRH, diethylstilbestrol, theophylline, enkephalins, E series prostaglandins, peptides of the VIP-secretin-glucagon-GRF family and other growth hormone secretagogues such as GHRP-6, GHRP-1 as described in U.S. Pat. No. 4,411,890; benzo fused lactams such as those disclosed in U.S. Patent No. 5,206,235; and growth hormone releasing hormone (GHRH) and its analogs or growth hormone (GH) and its analogs or somatomedins including IGF-1 and IGF-2 and their analogs.

The compounds of this invention are shown to induce release of growth hormone and IGF-1. It is known to those skilled in the art that there are many uses for growth hormone and the IGF's. Therefore administration of the compounds of this invention for purposes of stimulating the release of endogenous growth hormone or IGF-1 can have the same effects or uses as growth hormone or the somatomedins themselves. These uses of growth hormone and IGF-1 include the following: stimulating growth hormone release in elderly humans; prevention of catabolic side effects of glucocorticoids, treatment of osteoporosis, stimulation of the immune system, treatment of retardation, acceleration of wound healing, accelerating bone fracture repair, treatment of growth retardation, treating renal failure or insufficiency resulting in growth retardation, treatment of physiological short stature, including growth hormone deficient children, treating short stature associated with chronic illness, treatment of obesity and growth retardation associated with obesity, treating growth retardation associated with Prader-Willi syndrome and Turner's syndrome; accelerating the recovery and reducing hospitalization of burn patients; treatment of intrauterine growth retardation, skeletal dysplasia, hypercortisolism and Cushings syndrome; Induction of pulsatile growth hormone

release; replacement of growth hormone in stressed patients; treatment of osteochondrodysplasias, Noonans syndrome, schizophrenia, depression, Alzheimer's disease, diseases of demyelination, multiple sclerosis, delayed wound healing, and psychosocial deprivation; treatment of pulmonary dysfunction and ventilator dependency; attenuation of
 5 protein catabolic response after a major operation; reducing cachexia and protein loss due to chronic illness such as cancer or AIDS; treatment of hyperinsulinemia including Type II diabetes; adjuvant treatment for ovulation induction; stimulating thymic development and prevent the age-related decline of thymic function; treatment of immunosuppressed patients; treatment of bone marrow transplanted patients, improvement in muscle strength, mobility,
 10 diseases of muscle function, muscular dystrophy's, maintenance of skin thickness, metabolic homeostasis, enhancing renal function and hemeostasis including acute and chronic renal failure, stimulation of osteoblasts, bone remodeling, and cartilage growth; stimulation of the immune system in companion animals; growth promotion in livestock including stimulation of milk production in ruminates and wool or hair growth.

15 An alternative use of the GHRP's of this invention, represented by formulae I-V, as well as other GHRP's as defined herein, including but not limited to GHRP-6 and GHRP-1 as described in U.S. Pat. No. 4,411,890; GHRP-2; benzo fused lactam GHRP's such as those disclosed in U.S. Patent No. 5,206,235; are used in combination with IGF-1 to treat diseases in which long term IGF-1 treatment is indicated. This use of GHRP's is to bring serum GH levels
 20 back to normal when long-term IGF-1 therapy down-regulates the pituitary GH secretion. Such use includes but is not limited to use in the treatment of Type II diabetes.

Other uses of the instant compounds will be apparent from the following references; Amato et al. , *Journal of Clinical Endocrinology and Metabolism* 77(6):1671-1676 (1993), Bengtsson et al. , *Journal of Clinical Endocrinology and Metabolism* 76(2):309-317 (1993), Binnerts et al. , *Clinical*
 25 *Endocrinology* 37:79-87 (1992), Bowers, *Journal of Clinical Endocrinology and Metabolism* 76(4):817-823 (1993), Cuneo et al. , *J. Applied Physiol.* 70(2):688-694 (1991), Cuneo et al. , *J Applied Physiol.* 70(2):695-700 (1991), Degerblad et al. , *Acta Endocrinologica* 126:387-93 (1992), Edén et al. , *Arteriosclerosis and Thrombosis* 13(2):296-301 (1993), Hartman et al. , *Horm Res* 40:37-47 (1993), Ho et al. , *Horm Res* 40:80-86 (1993), Jørgensen et al. , *Acta Endocrinologica* 125:449-453 (1991),
 30 Jørgensen et al. , *The Lancet* June 3:1221-1224 (1989), Lamberts et al. , *Clinical Endocrinology* 37:111-115 (1992), McGauley et al. , *Horm Res* 33(suppl 4):52-54 (1990), Møller et al. , *Clinical Endocrinology* 39:403-408 (1993), O'Halloran et al. , *Journal of Clinical Endocrinology and Metabolism* 76(5):1344-1348 (1993), Orme et al. , *Clinical Endocrinology* 37:453-459 (1992), Rodriguez-Arao et al. , *Horm Res* 39:87-88 (1993), Rosén et al. , *Clinical Endocrinology* 40:111-116 (1994), Rosén et al. ,
 35 *Acta Endocrinologica* 129:195-200 (1993), Rudman et al. , *The New England Journal of Medicine* 323(1):1-6 (1990), Salomon et al. , *The New England Journal of Medicine* 321(26):1797-1803 (1989), Shibasaki et al. , *Journal of Clinical Endocrinology and Metabolism* 58(1):212-214 (1984), Sönksen et al. , *Acta Paediatr Scand [Suppl]* 379:139-146 (1991), Tauber et al. , *Journal of Clinical Endocrinology*

and *Metabolism* 76(5):1135-1139 (1993), Vandeweghe *et al.*, *Clinical Endocrinology* 39:409-415 (1993), Whitehead *et al.*, *Clinical Endocrinology* 36:45-52 (1992), and Bercu *et al.*, U.S. patent no. 5,246,920.

Additionally, the most potent compounds of this invention can be used as GH antagonists. It is known that hypothalamic hormones that are super agonists can also be used as antagonists. For example super agonists of Gonadotrophin Releasing Hormone (GnRH) such as GONADORELIN and LEUPROLIDE act either as agonists or antagonists depending on the method of administration. The actions of the GnRH super agonists are summarized in Goodman and Gilman's, *The Pharmacological Basis of Therapeutics*, 8th Ed., McGraw Hill Inc., p. 1353 (1993). By analogy, it is believed the continuous administration of the compounds of formula I-V will lead to down-regulation of the growth response. These molecules can therefore be used as functional antagonists of pituitary GH secretion, thereby antagonizing GH or IGF-1 action.

The uses of such antagonists of GH secretion include but are not limited to; treatment of excess GH secretion as in acromegaly or gigantism; in cancer of the breast, colon and prostate; in diabetes especially in Type I adolescent patients to counteract the dawn phenomenon; and in Type I and Type II patients to directly control blood glucose, and to control the long-term effects of diabetes, as for example in retinopathy.

The compounds of this invention can be administered by oral, parenteral (*e.g.*, intramuscular, intraperitoneal, intravenous or subcutaneous injection or infusion, or implant), nasal, pulmonary, vaginal, rectal, sublingual, or topical routes of administration and can be formulated in dosage forms appropriate for each route of administration.

C. Methods of Making

1. General Peptide Synthesis

One method of producing GHRP's involves chemical synthesis of the "polypeptide". This can be accomplished using methodologies well known to those skilled in the art (see Stewart, J. M. & Young, J. D. *Solid Phase Peptide Synthesis* Pierce Chemical Co. Rockford, IL[1984]; see also U. S. Patent No.'s 4,105,603; 3,972,859; 3,842,067; and 3,862,925).

"Polypeptides" of the invention may be conveniently prepared using solid phase peptide synthesis (Merrifield, *J. Am. Chem. Soc.*, 85:2149 [1964]; Houghten, *Proc. Natl. Acad. Sci. USA* 82:5132 [1985]). Solid phase synthesis begins at the carboxy-terminus of the putative peptide by coupling a protected amino acid to a suitable resin (*e.g.* chloromethylated polystyrene resin) as shown in Figures 1-1 and 1-2, on pages 2 and 4 of Stewart and Young *supra*. After removal of the α -amino protecting group with, for example, trifluoroacetic acid (TFA) in methylene chloride and neutralizing in, for example TEA, the next α -amino- and sidechain protected amino acid in the synthesis is added. The remaining α -amino- and, if necessary, side-chain-protected amino acids are then coupled sequentially in the desired order

by condensation to obtain an intermediate compound connected to the resin. Alternatively, some amino acids may be coupled to one another forming a peptide prior to addition of the peptide to the growing solid phase polypeptide chain.

The condensation between two amino acids, or an amino acid and a peptide, or a peptide and a peptide can be carried out according to the usual condensation methods such as the azide method, mixed acid anhydride method, DCC (N,N'-dicyclohexylcarbodiimide) or DIPC (N,N'-diisopropylcarbodiimide) methods, active ester method (p-nitrophenyl ester method, BOP [benzotriazole-1-yl-oxy-tris (dimethylamino) phosphonium hexafluorophosphate] method, N-hydroxysuccinic acid imido ester method, etc., and Woodward reagent K method.

Common to chemical syntheses of peptides is the protection of any reactive side-chain groups of the amino acids with suitable protecting groups. Ultimately these protecting groups are removed after the desired polypeptide chain has been sequentially assembled. Also common is the protection of the α -amino group on an amino acid or a fragment while that entity reacts at the carboxyl group followed by the selective removal of the α -amino-protecting group to allow subsequent reaction to take place at that location. Accordingly, it is common in polypeptide synthesis that an intermediate compound is produced which contains each of the amino acid residues located in the desired sequence in the peptide chain with various of these residues having side-chain protecting groups attached. These protecting groups are then commonly removed substantially at the same time so as to produce the desired resultant product following removal from the resin.

Suitable protective groups for protecting the α - and ϵ - amino side chain groups are exemplified by benzyloxycarbonyl (CBZ), isonicotinylloxycarbonyl (iNOC), O-chlorobenzyloxycarbonyl (2-Cl-CBZ), p-nitrobenzyloxycarbonyl [Z(NO₂)], p-methoxybenzyloxycarbonyl [Z(OMe)], t-butoxycarbonyl (BOC), t-amylloxycarbonyl (AOC), isobornylloxycarbonyl, adamantylloxycarbonyl, 2-(4-biphenyl)-2-propyl-oxycarbonyl (BPOC), 9-fluorenylmethoxycarbonyl (Fmoc), methylsulfonyl ethoxycarbonyl (Msc), trifluoroacetyl, phthalyl, formyl, 2-nitrophenylsulphenyl (NPS), diphenylphosphinothioyl (Ppt), dimethylphosphinothioyl (Mpt) and the like.

Protective groups for the carboxy functional group are exemplified by: benzyl ester (OBzl), cyclohexyl ester (Chx), 4-nitrobenzyl ester (ONb), t-butyl ester (OtBu), 4-pyridylmethyl ester (OPic), and the like. It is often desirable that specific amino acids such as arginine, cysteine, and serine possessing a functional group other than amino and carboxyl groups are protected by a suitable protective group. For example, the guanidino group of arginine may be protected with nitro, p-toluenesulfonyl, benzyloxycarbonyl, adamantylloxycarbonyl, p-methoxybenzenesulfonyl, 4-methoxy-2, 6-dimethylbenzenesulfonyl (Mds), 1,3,5-trimethylphenylsulfonyl (Mts), and the like. The thiol group of cysteine may be protected with p-methoxybenzyl, triphenylmethyl, acetylaminoethyl ethylcarbamoyl, 4-methylbenzyl, 2, 4,

6-trimethylbenzyl (Tmb) etc., and the hydroxyl group of serine can be protected with benzyl, t-butyl, acetyl, tetrahydropyranyl and the like.

Stewart and Young *supra* provides detailed information regarding procedures for preparing peptides. Protection of α -amino groups is described on pages 14-18, and side-chain blockage is described on pages 18-28. A table of protecting groups for amine, hydroxyl and
5 sulfhydryl functions is provided on pages 149-151.

After the desired amino acid sequence has been completed, the intermediate peptide is removed from the resin support by treatment with a reagent, such as liquid HF and one or more thio-containing scavengers, which not only cleaves the peptide from the resin, but also
10 cleaves all the remaining side-chain protecting groups. Following HF cleavage, the peptide residue is washed with ether, and extracted from the resin by washing with aqueous acetonitrile and acetic acid.

Preferably in order to avoid alkylation of residues in the polypeptide, (for example, alkylation of methionine, cysteine, and tyrosine residues) a thio-cresol and cresol scavenger
15 mixture is used.

2. Other General Procedures

The peptidomimetic compounds of this invention may also be conveniently prepared by the methods for peptide synthesis described in monographs such as ("Principles of Peptide Synthesis, M. Bodanszky, Springer-Verlag, 2nd Ed., 1993; "Synthetic Peptides: A Users Guide",
20 G. A. Grant, Ed, W. H. Freeman and Co., 1992; and references cited therein), or by other methods generally known to one skilled in the art. The synthesis of compounds of this invention that are peptidomimetic in nature (*i.e.* contain other than standard amide bond linkages) may be prepared by extension of the methods described in the specific Examples 1-37 and the methods laid forth in Schemes I-IV below, by the general synthetic methods described
25 in "Comprehensive Organic Transformations", R. C. Larock, VCH Publishers, 1989, and by methods generally known to one skilled in the art.

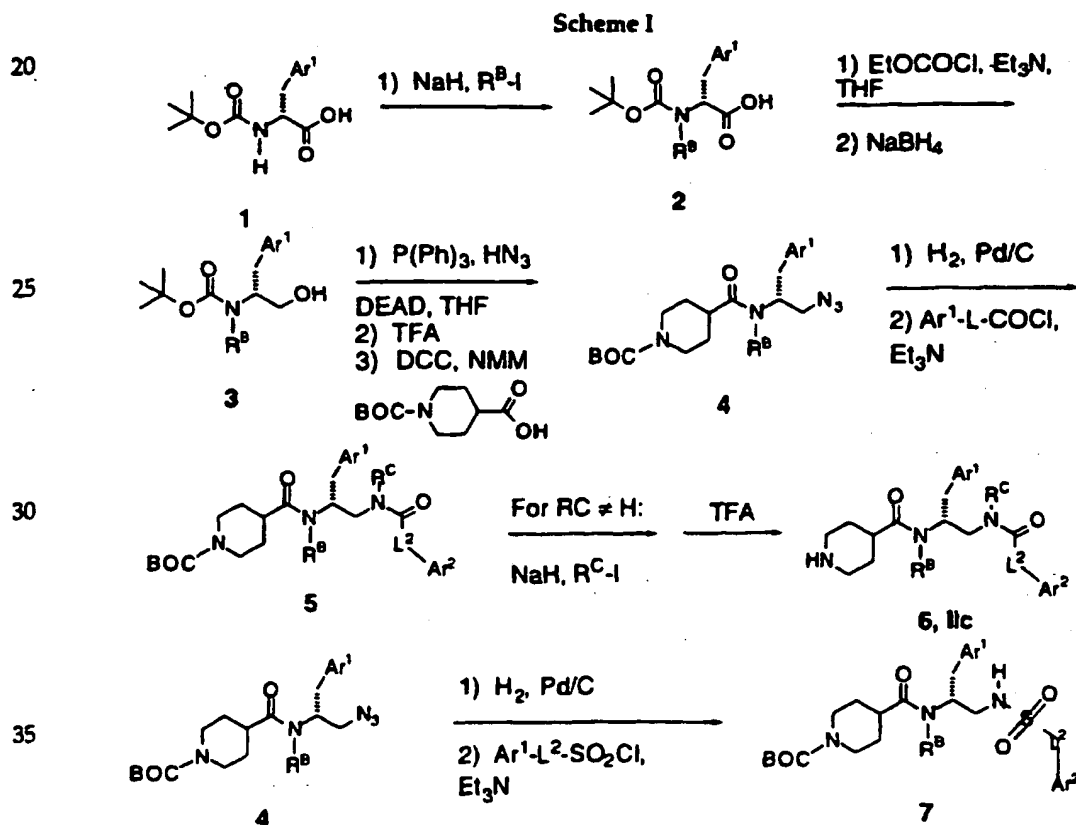
For compounds of Claim 1 where the amide linkages ($-C(=O)-NH-$) are replaced with amide isostere linkages such as; $-CH_2-NH-$, $-CH_2-S-$, $-CH_2-CH_2-$, $-CH=CH-$ (cis and trans), $-C(=O)-CH_2-$, $-CH(OH)-CH_2-$, $-CH(CN)-NH-$, $-O-C(=O)-NH-$ and $-CH_2-SO-$, amide bond
30 replacing methods known in the art are employed. The following references describe preparation of amide isostere linkages which include these alternative-linking moieties: Spatola, A.F., Vega Data 1(3): "Peptide Backbone Modifications" (General Review) (Mar 1983), Spatola, A.F., in "Chemistry and biochemistry of Amino Acids Peptides and Proteins", B. Weinstein, ed., Marcel Dekker, New York, P. 267 (1983); Morley, J.S., *Trends Pharm. Sci.* pp. 463-
35 468; Hudson, D. *et al. Int. J. Pept. Prot. Res.* 14:177-185 (1979) ($-CH_2NH-$, $-CH_2CH_2-$); Spatola, A.F., *et al., Life Sci.* 38:1243-1249 (1986) ($-CH_2-S-$); Hann, M.M., *J. Chem. Soc. Perkin. Trans. I* 307-314 (1982) ($-CH=CH-$, cis and trans); Almquist, R.G., *et al., J. Med. Chem.* 23:1392-1398 (1980) ($-C(=O)-CH_2-$); Jennings-White C., *et al., Tetrahedron Lett* 23:(1982) ($-C(=O)-CH_2-$); Szelke, M., *et*

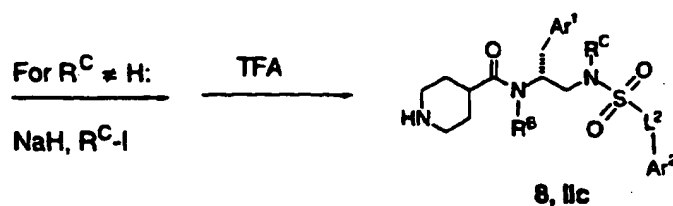
al., EP Application No. 45665 (1982) *Chem Abs* 9739405 (1982) (-CH(OH)-CH₂); Holladay, M.W., *et al.*, *Tetrahedron Lett* 24:4401-4404 (1983) (-C(OH)-CH₂-); Hruby, V.J. *Life Sci* 31:189-199 (1982) (-CH₂S-); and Cho, C.Y. *et al.*, *Science* 261:1303-1305 (1993) (-O-C(=O)-NH-).

The compounds of Claims 2-5 are specifically prepared by the methods described in Schemes I-IV. The N-terminal amino group is shown as isonipecotic acid for clarity, but it is understood that the compounds of this invention with other groups (R^A) at this position are prepared by substitution of the appropriately protected reagent for the protected isonipecotic acid in the scheme. One may in general use a range of methods for the coupling of the components such as preformed active esters, acid chlorides, and coupling reagents. For connections other than amides, alkylation, acylation, and sulfonylation, for example, may be accomplished using the appropriately activated reagent and methods described in ("Comprehensive Organic Transformations", R. C. Larock, VCH Publishers, 1989).

3. Specific Schemes

As shown in Scheme I, protected amino acids of the type 1 may be alkylated according to the procedure of Benoitin (*Can. J. Chem.* 55, 906, 1977) to give a variety of N-substituted compounds (2).





To produce reduced or inverted amide compounds of the Type IIc (Claim 3), (2) may be reduced through the preformed mixed anhydride with sodium borohydride to give protected amino alcohols (3). Conversion of the hydroxyl function to an amine may be accomplished via Mitsunobu coupling of (3) with hydrazoic acid to give an intermediate protected amino azide. Deblocking of the amino function and coupling to the N-terminal group is conveniently performed at this differentiated stage. In this example, the N-BOC is removed with TFA and the resulting free-based amino azide coupled to N-BOC-isonipecotic acid using the reagent DCC to give (4).

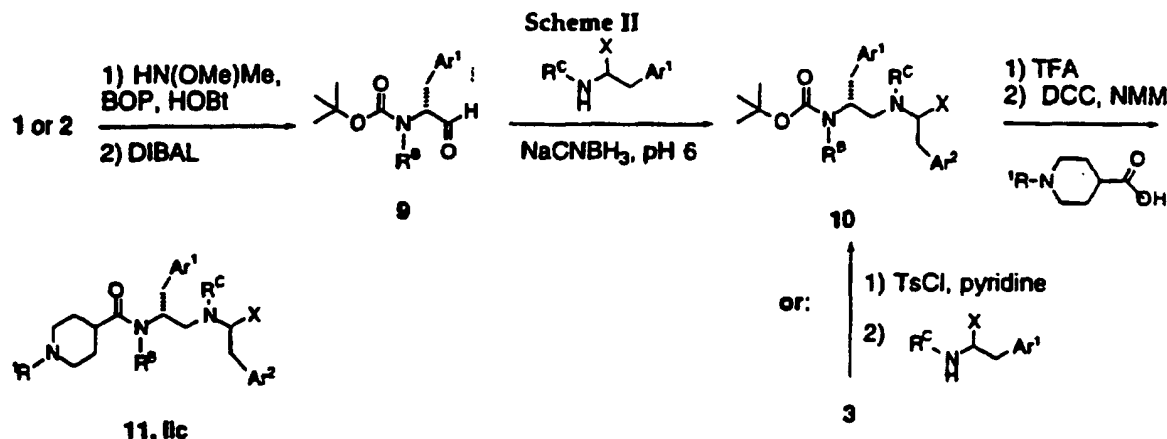
The intermediate azide (4) can be converted to a variety of compounds of this invention. For example, hydrogenation of the azido function gives an amine which can be acylated with a variety of groups, for example, $\text{Ar}^1\text{-L}^2\text{-COCl}$ to give (5) (Scheme I). For the synthesis of the range of L^2 's herein claimed, it is understood that $\text{Ar}^1\text{-L}^2\text{-COCl}$ may be substituted with a variety of different acylating agents like chloro carbonates, activated esters, isocyanates, and the like. For example, 2-naphthoylchloride, benzylchloroformate, phenylacetyl chloride, dihydrocinnamoyl chloride, and phenylisocyanate may be used to give compounds (6) with a range of linkers L^2 . Global deprotection then gives 6, for $R^C = H$. Incorporation of the N-substitution R^C , into (5) can be accomplished via alkylation of (5), for example via deprotonation with sodium hydride and reaction with an alkyl halide. Deprotection gives (6) ($R^C \neq H$).

For synthesis of N-sulfonamido compounds, the amine produced via reduction of (4) can be sulfonylated, alternately alkylated at nitrogen (for $R^C \neq H$), and deprotected to give (8).

For the synthesis of compounds (6) and (8) where $R^B = H$ and $R^C \neq H$, it may be more convenient to incorporate the substituent R^C via reductive amination. For example, using one equivalent of an appropriate aldehyde and sodium cyanoborohydride, R^C can be introduced into the amine from reduction of (4), prior to acylation or sulfonylation to (6) or (8), respectively.

Compounds IIc (Claim 3) in which $X = H$, alkyl, substituted alkyl, and the like, may be synthesized via the route shown in Scheme II.

30



- The intermediate protected amino acids (1) or (2) (from Scheme I) are converted to protected amino aldehydes (9), conveniently via DIBAL reduction of the derived N-methyl-N-methoxyamide. Subsequent reductive amination with an appropriately substituted amine gives (10).
- Alternatively, (10) can be prepared from (3) via conversion of the alcohol to a tosylate or other suitable leaving group, and displacement with an appropriately substituted amine (Scheme II). It should be apparent that a wide variety of amines could be used in these two routes to (10) including tryptamine, N-methyl-(2-naphthyl)ethyl amine, alpha-methylphenethylamine, tryptophanol, and N-methyl-beta-naphthylalanol. The amine may also be part of a heterocycle, i.e. 2-benzyl- or naphthylmethyl-piperidine.

This reductive amination strategy is a general method for incorporation of a reduced amide isostere into a polyamide chain. Thus, substitution of an appropriately protected amino acid derivative or peptide with a free alpha-amine, for the amine component in Scheme II (9) to (10) provides a protected, reduced amide isostere, intermediate. In this manner, compounds of the type exemplified in Claim 8, IIIb-IIIe may be prepared using appropriate orthogonal protecting groups for the reactive functionality. This method may also be employed when the amine component is attached to a solid support, suitable for peptide synthesis, providing a convenient method for the synthesis of longer peptidic compounds.

Completion of the synthesis of compounds of the type 11 (Scheme II) (and by analogy, compounds of Claim 8, IIIb-IIIe), requires deprotection of the amino group and coupling to an appropriately protected N-terminal moiety, shown in Scheme II as N-R¹-isonipecotic acid for clarity. Depending on the particular substituents X, R^B, and R^C in (10), it may be necessary to orthogonally protect reactive functionality prior to removal of the N-terminal blocking group. For example, for (10) (R^C = X = H) the secondary amine in (10) can be acylated with Fmoc-Cl (e.g. R^C = Fmoc) prior to removal of the N-terminal BOC. This ensures that the subsequent acylation occurs only at the terminal amine.

It will be noted that a wide variety of N-terminal groups can be attached to the intermediate deprotected (10). Any suitably protected amino acid, i.e. BOC-4-aminobutyric

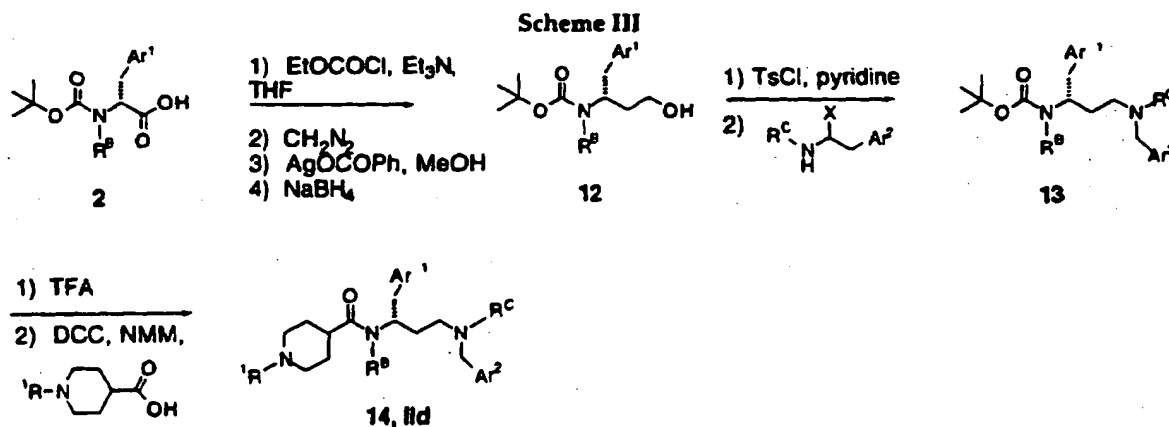
acid, N-alkyl-isonipecotic acid, may be attached using a standard coupling reagent. Also, protected active esters, anhydrides, and acid chlorides, may be used. For the synthesis of urea type linkages, the intermediate deprotected (10) may be reacted with carbonyl diimidazole or phosgene, followed by addition of a suitably protected or symmetrical amine. In particular, reaction with piperazine, propane diamine, or N1,N4-dimethylpropanediamine, give preferred compounds.

For the synthesis of compounds with an N-terminal carbamate linkage, the intermediate deprotected (10) ($R^C \neq H$) may be reacted with carbonyl diimidazole or phosgene, followed by addition of a suitably N-protected amino alcohol such as BOC-aminoethanol, BOC-aminopropanol, and BOC-2- or 3-hydroxypiperidine. Alternatively, the intermediate deprotected (10) can be reacted directly with a preformed N-blocked-chloroformate.

The final step necessary for completion of the synthesis of the compounds of type 11 is removal of the protecting functionality using appropriate conditions (for a general monograph on protecting groups, see Greene, W.T., Wuts P.G.M. *Protective Groups in Organic Synthesis*, 2nd Ed., John Wiley & Sons, NY [1991]).

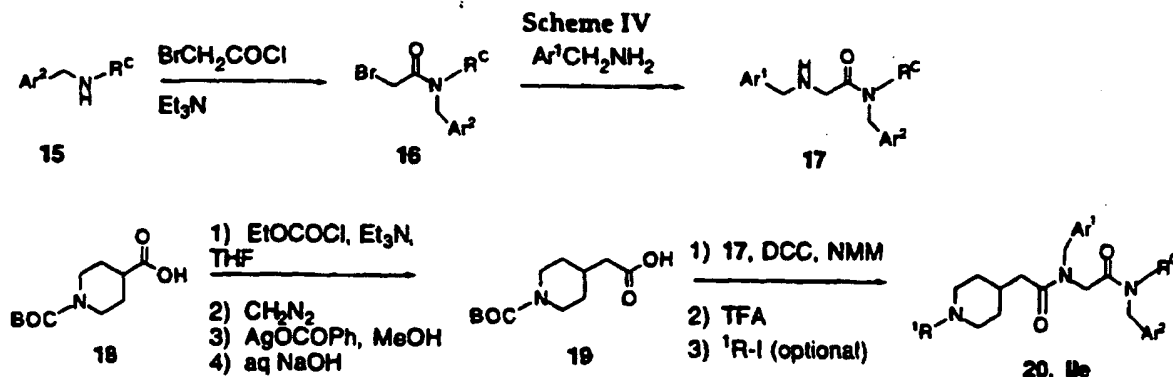
It will be noted that these methods for incorporation of different N-terminal groups (e.g. $RA's$) are generally applicable to the compounds of this invention and not limited to the particular example of Scheme II.

The synthesis of the peptidomimetic compounds of Claim 3, IId-IIg, are shown below in Schemes III-VI. For the synthesis of IId (Scheme III), protected amino acid (2) or (1 for $R^B = H$) is converted to the homologous methyl ester via rearrangement of the diazoketone with Ag(I) in methanol.



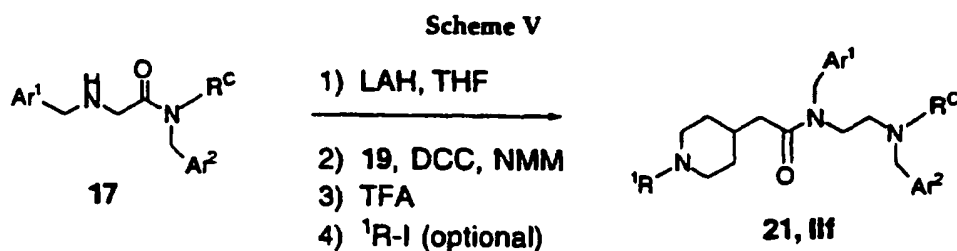
Reduction of the ester provides alcohol (12) which, when converted to a tosylate or similar leaving group, can be displaced by a large range of substituted amines, as exemplified by the conversion of 3 to 10 (Scheme II). Deprotection of the product (13) and acylation provides (14) after deprotection.

Compounds of the type IIe can be prepared as shown in Scheme IV.



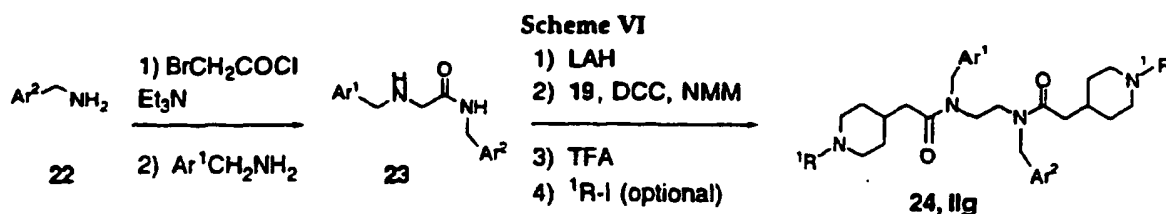
Substituted amine (15) is acylated with bromoacetyl bromide to give (16), which is reacted with a second amine to give (17). Acylation with an appropriate N-terminal moiety gives (20). For example, a preferred N-terminus, 4-carboxymethylpiperidine (19), is prepared via homologation of BOC-isonipetric acid (18), and acylated onto (17) with DCC. Deprotection and optional alkylation of the terminal amine provides (20). Reductive amination with an appropriate aldehyde is an alternate method for the incorporation of R¹ substituents onto the terminal amine. This is a generally applicable method, useful for many compounds of this invention.

Compounds of the type IIf (Scheme V) can be prepared from (17) via LAH reduction of the amide functionality. Acylation with (19), deprotection, and optional N-alkylation provides (21).



The pseudo symmetrical compounds of type IIg are preparable via the route shown in Scheme

VI. Conversion of arylamine (22) to (23) is analogous to the preparation of (17) above.

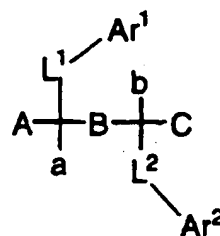


LAH reduction provides a symmetrical or unsymmetrical substituted ethane diamine which is acylated simultaneously at both nitrogens with 19 or another appropriate reagent. Deblocking as above gives (24).

5 D. Preferred Embodiments

The present invention is based on the discovery of several new classes of small peptidomimetics that cause the release of growth hormone in mammals. It is a preferable object of the present invention to provide agents that are selective for GH release and have suitable safety and efficacy for chronic administration to mammals. In a more preferred embodiment, the present invention provides compounds which are suitable for oral, intranasal, or pulmonary delivery. It is an aim of the most preferred embodiments of the present invention to provide compounds that are superior to the prior art by the above criteria. It is further preferred that the compounds be readily synthesizable in optically pure form where necessary.

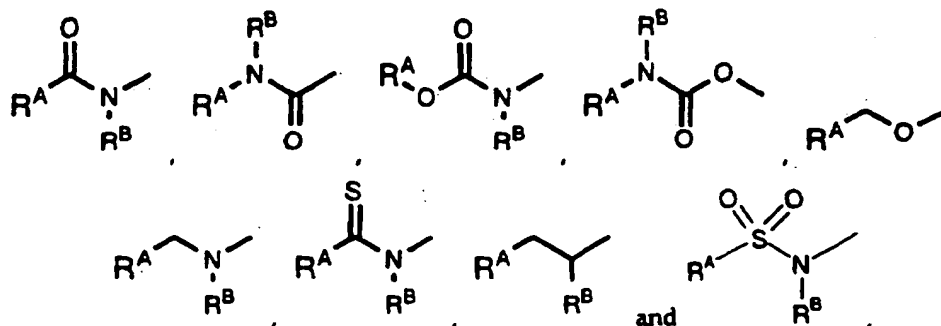
In view of the forgoing, the preferred compounds of this invention have an EC₅₀ in the rat "pit" cell assay of less than about 1.0 nM and most preferably less than about 0.5 nM. Preferred compounds of this invention also have a molecular weight less than 650 da and most preferably less than 600 da. Preferred embodiments of the compounds of this invention are represented by structural Formula (I)



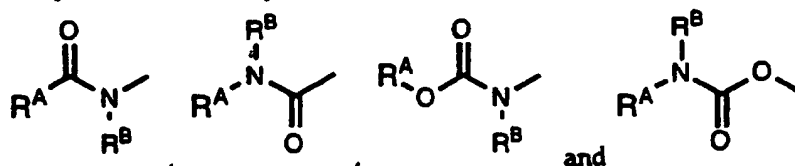
I

20 where the symbols of Formula (I) are selected from the following:

From the substructures shown for group A of Formula (I), those preferred A's are selected from;

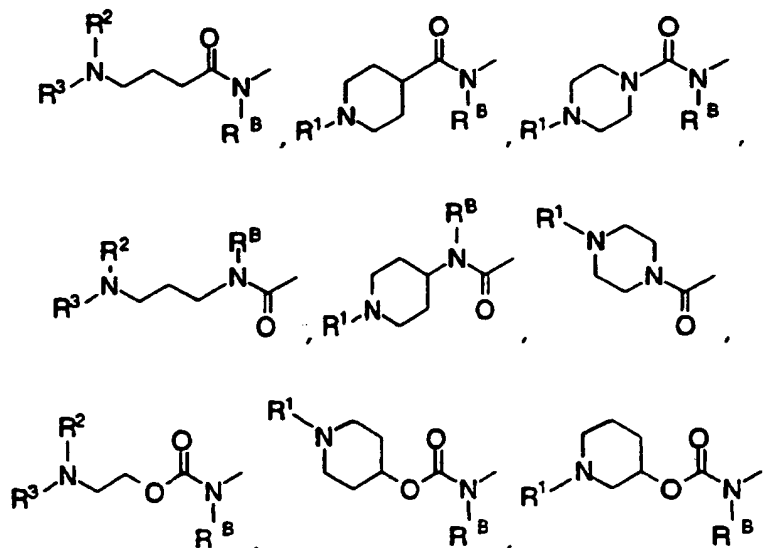


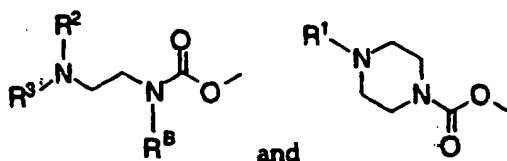
while the most preferred A's incorporate either an amide or carbamate linkage as in;



With respect to certain combinations of $L^1\text{-Ar}^1$ and B, it is preferred to use a commercially available amino acid as a starting material.

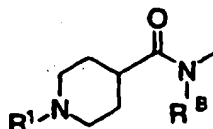
- 5 For the groups R^A , of substructure A, preferred embodiments incorporate functionality that places a basic nitrogen atom (or prodrug form thereof) at a distance of approximately 4 to 8 C-C bonds from the attachment point of $L^1\text{-Ar}^1$, in a through-bond measurement. For example, preferred R^A 's, of the most preferred A substructures above, include alkyl amines $(\text{CH}_2)_n\text{NR}^2\text{R}^3$ (where $n=2$ to 4) and saturated six-membered ring heterocycles containing 1 or 2
- 10 nitrogen atoms, for the amide-linked A's, and $(\text{CH}_2)_n\text{NR}^2\text{R}^3$ (where $n=2$ or 3) and 3- or 4-substituted saturated six-membered ring heterocycles containing 1 or 2 nitrogen atoms, for the carbamate-linked A's. Preferred R's attached at the nitrogen atom of R^A include hydrogen, methyl, ethyl, 2-hydroxyethyl, and 2-hydroxypropyl. More preferred, are those R^A 's that place the amine at approximately 6 C-C bonds from the attachment point of $L^1\text{-Ar}^1$, measured in a
- 15 through-bond manner, as is the case with the most preferred R^A 's $(\text{CH}_2)_3\text{NR}^2\text{R}^3$, 4-piperidinyl, and piperazinyl, for the amide linked A's, and $(\text{CH}_2)_3\text{NR}^2\text{R}^3$ for the carbamate linked A's, where R^2 and R^3 are chosen from the group hydrogen and methyl. In the case of the carbamate where R^A is attached directly to nitrogen, an additionally preferred R^A forms a piperazine, where the carbamate nitrogen is incorporated as a ring atom. Thus, a most preferred
- 20 embodiment of the present invention incorporates a substructure A of the following composition:





where the groups R^B are hydrogen or lower alkyl.

In the most preferred embodiment of the instant invention, the A substructure of Formula (I) is an amide derived from attachment of isonipecotic (inip) acid (piperidine-4-carboxylic acid)



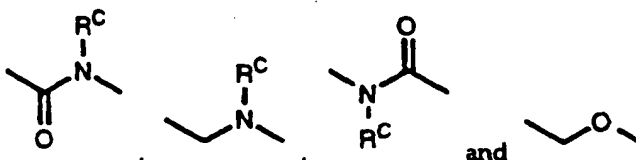
where the groups R^B and R^1 are hydrogen or lower alkyl.

As is taught in the present invention, the above substructure A's display the amine functionality at a near optimal distance from L^1-Ar^1 in Formula (I) and thus it is preferred that the appended R^A 's for other substructure A's of the instant invention mimic this distance as closely as possible, preferably through the incorporation of a rigidifying carbo- or heterocyclic substructure.

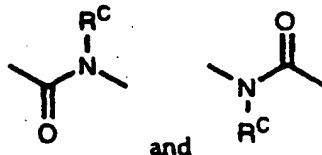
Preferred embodiments of the other substructures of the compound represented in Formula (I) are as follows:

For the groups a and b, hydrogen and methyl, independently selected, are more preferred. In a most preferred embodiment a and b are both hydrogen.

For the substructure B, which links the two aromatic sidechains, the amide, amine, and ether of the following functions are more preferred;



The most preferred B's are selected from:

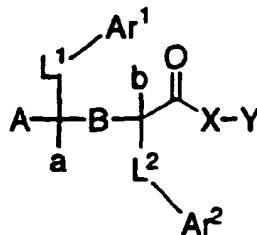


where R^C is additionally preferred to be hydrogen or methyl.

Moreover, when the substructure A, B, and/or C are comprised of the amide function $C(=O)NH$, the present invention teaches that the NH is replacable by NMe with retention of biological activity. It is therefore a most preferred embodiment of the present invention that the groups R^B , R^C , and R^D be independently selected from methyl and hydrogen, a particular

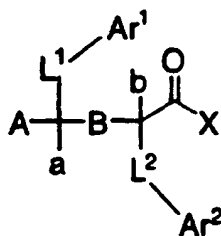
combination chosen so as to optimize for desired properties of the molecule, such as stability and lipophilicity.

- From the preferred list of substructures C, of Formula I, the more preferred embodiments are conveniently discussed by class. For the "pentapeptide, short series", exemplified by the most preferred (inip) b b F K -NH₂ (where C is -C(=O)-Phe - Lys - amide), and Formula Ia below, preferred embodiments include, in addition to the most preferred -C(=O)-Phe - Lys - amide, the substitution for Lys (Y) by the n-alkyl diamines H₂N(CH₂)_nNH₂, where n=2-6, and amino amides selected from the common amino acids.



(Ia)

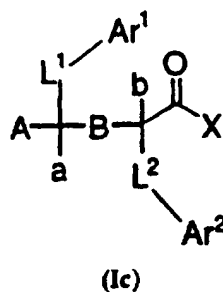
- As is taught in the present invention, a wide range of substitution is allowable at the Lys (Y) and, to a lesser extent, the Phe (X) position. It is therefore preferable to select from all possible C-terminal groups, those that are inexpensive, and improve the overall physical properties of the compound.



(Ib)

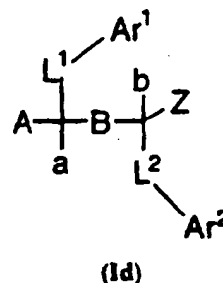
- For the Phe (X) position in the above formula (Ib) and the "tetrapeptide, short series", the Phe is most preferred when a C-terminal amide is included, as are L-alpha-naphthylalanine, L-beta-naphthylalanine, and Tyr. In the short series, exemplified by (inip) b b F -NH₂, the C-terminal carboxamide is a preferred embodiment. Also preferred are the amides N,N-dimethyl, N-methyl and morpholinyl. In a further preferred embodiment, the carboxamide is replaced with the free acid and the reduced congener CH₂OH and hydrogen.

- An additional class of most preferred compounds (Formula Ic) are obtained by the replacement of the Phe in the above structures with a non-aromatic residue. Most preferred among this class are the compounds where X (below) is an amide derived from the lower alkyl diamines and the lower alkyl aminocarboxamides. Most preferable is when X is butane diamine.



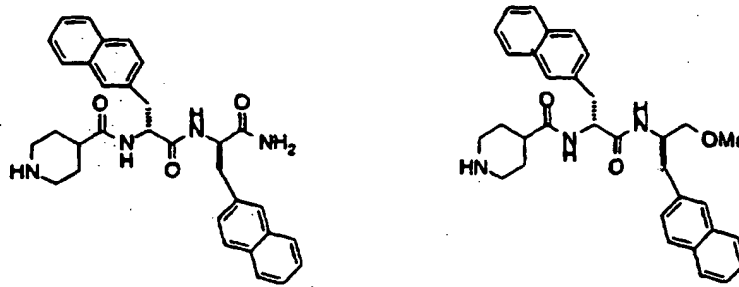
Most preferable is a compound where X is butane diamine and B is a N-methyl amide. Further most preferable compounds include those where X is NH₂, alkyl amides therefrom, OH, and it's lower alkyl esters.

- 5 In the "micro series", exemplified by (inip) b (wol), and depicted below, Z is preferred to be CH₂OH, CH₂OC(=O)R², CH₂NR²R³, CH₂OR, and hydrogen. Most preferred is Z = CH₂OH or hydrogen.

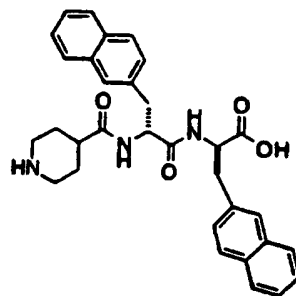
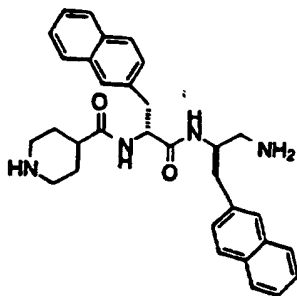


- 10 From the preferred list of L¹-Ar¹'s and L²-Ar²'s detailed in Claim 1, the most preferred are chosen from CH₂Ar, where Ar is preferably 1- or 2-naphthyl, 3-indoyl, or substituted phenyl. In a most highly preferred embodiment, L¹-Ar¹ is CH₂(2-naphthyl) and L²-Ar² is CH₂(3-indoyl) or CH₂(2-naphthyl).

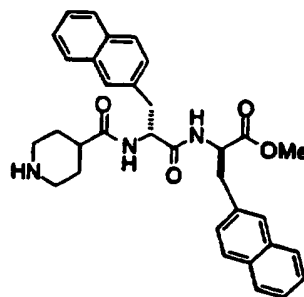
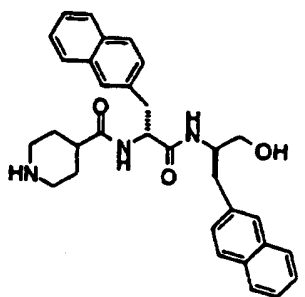
Other most preferred compounds of the present invention include:



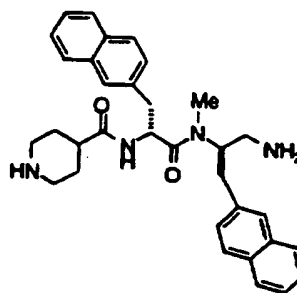
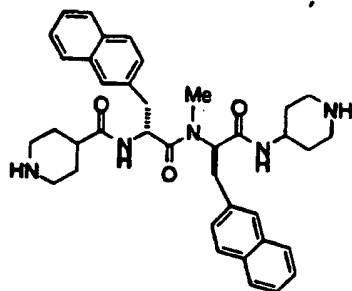
5



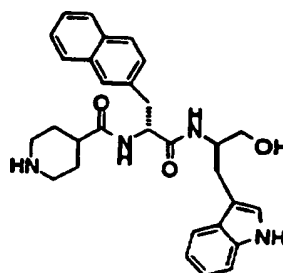
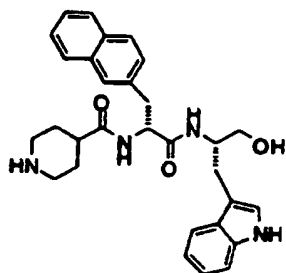
10



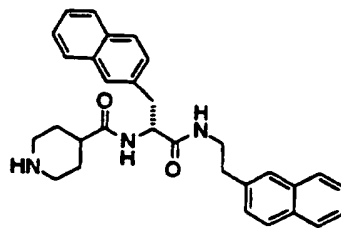
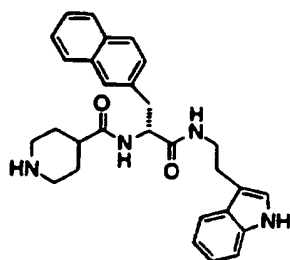
15



20

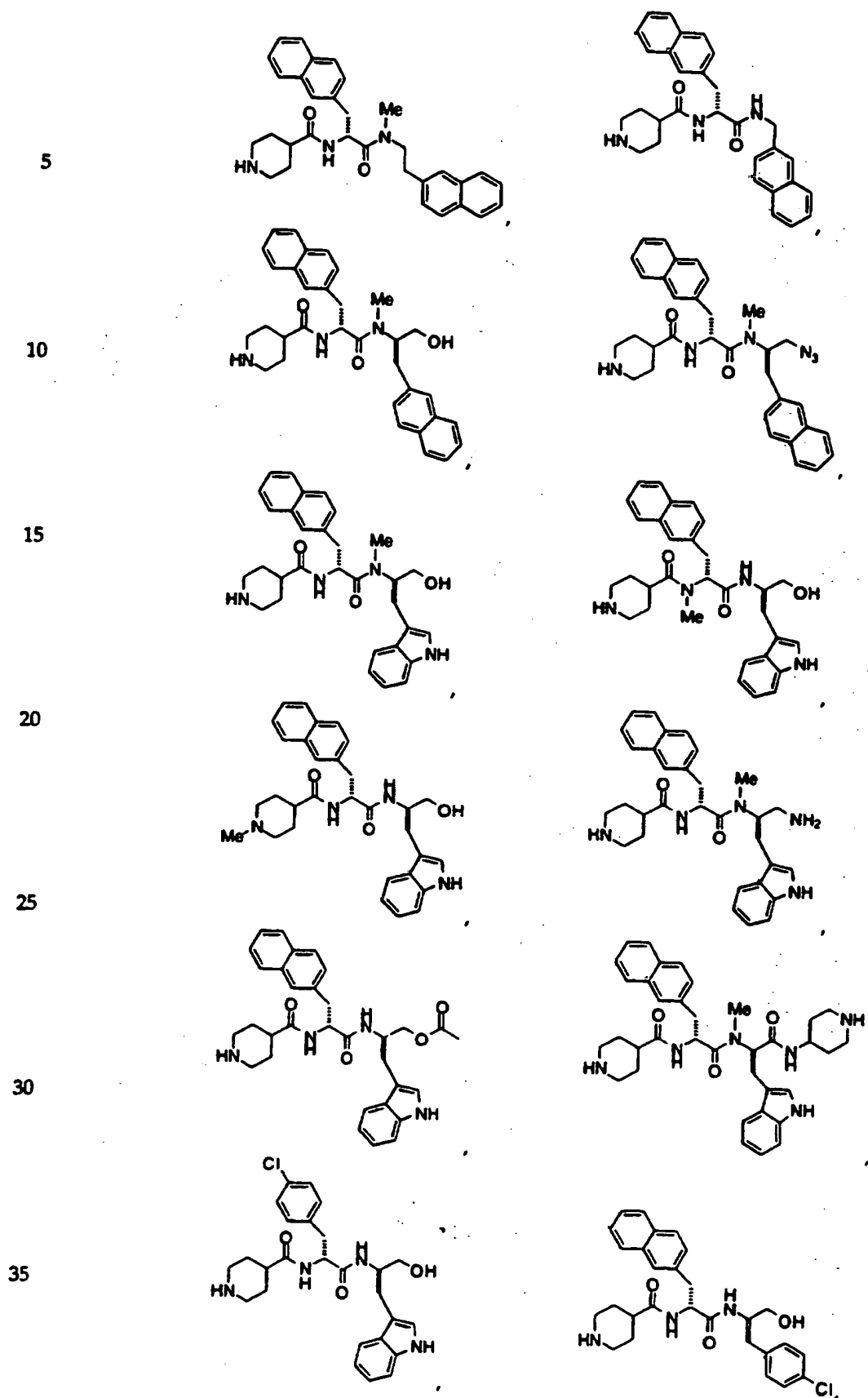


25

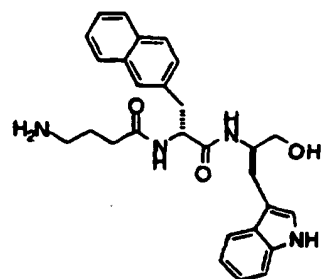
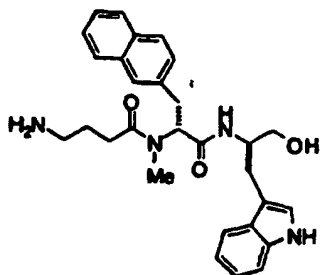


30

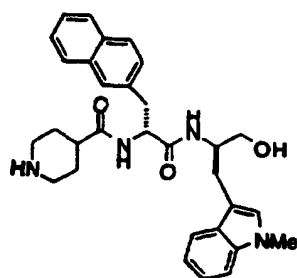
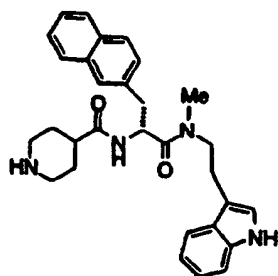
35



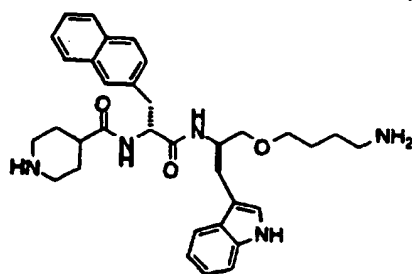
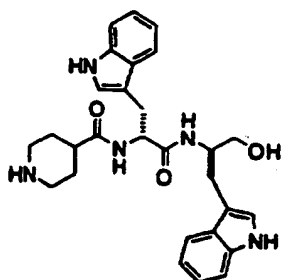
5



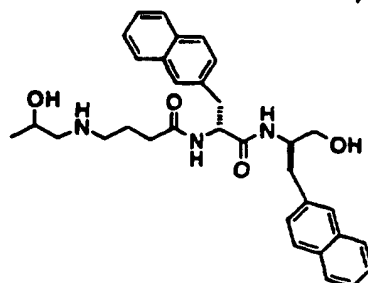
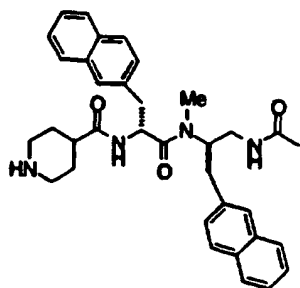
10



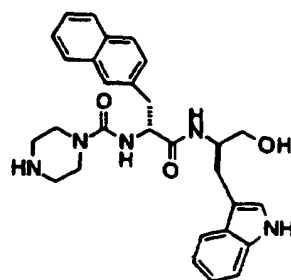
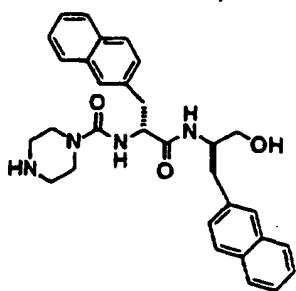
15



20

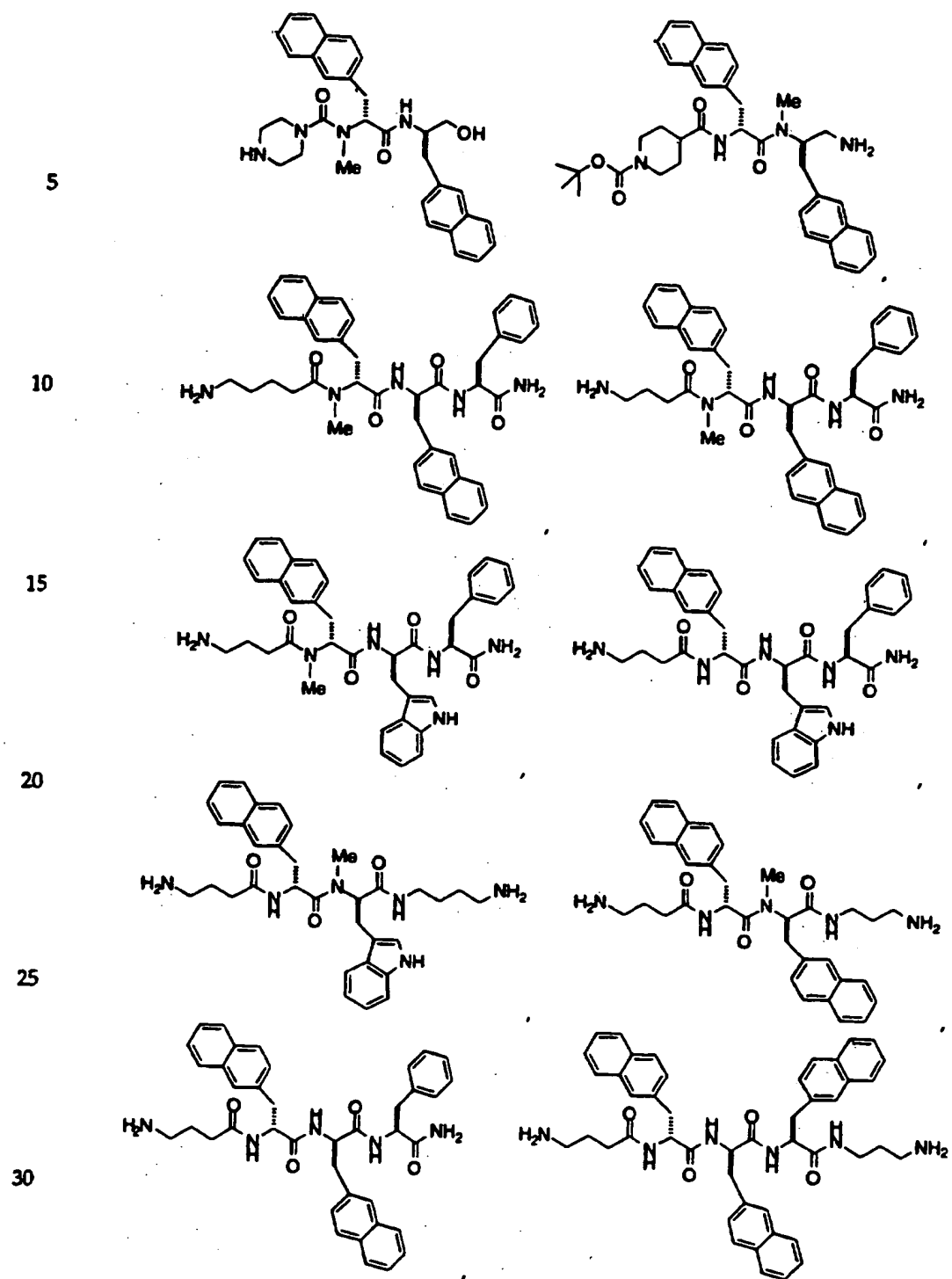


25

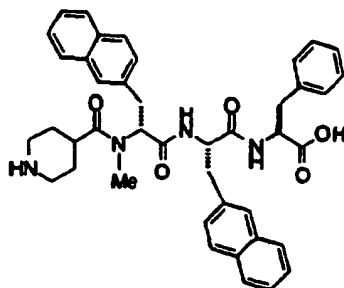
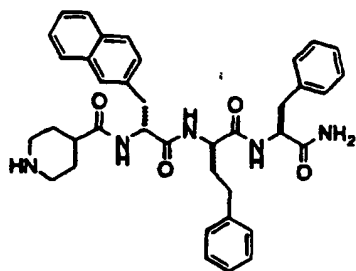


30

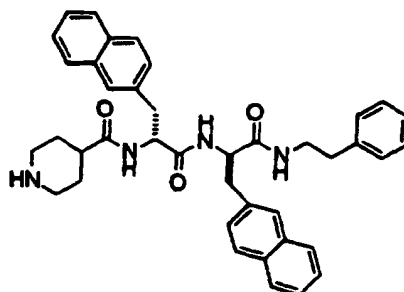
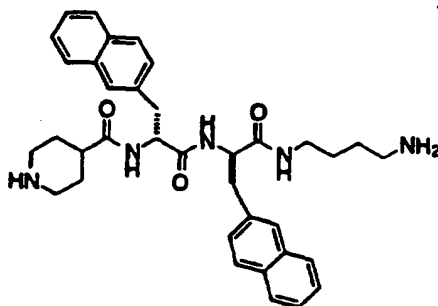
35



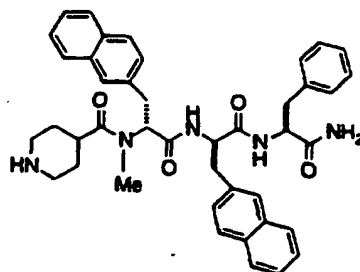
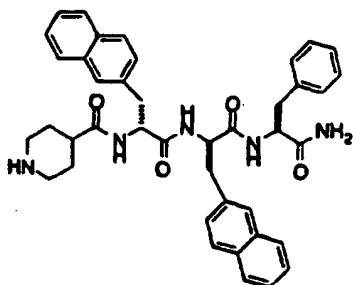
5



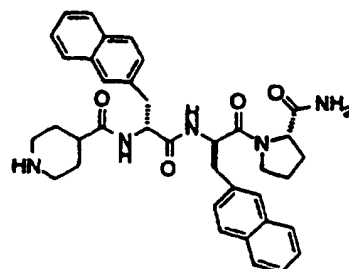
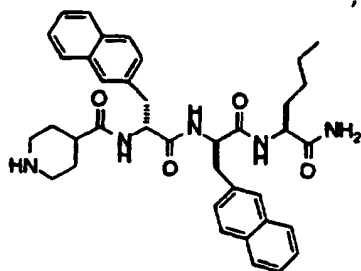
10



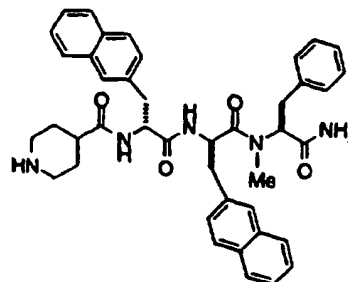
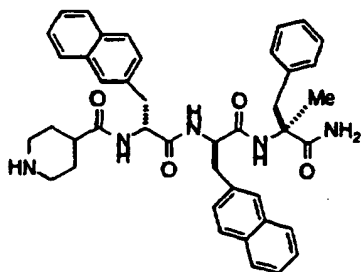
15



20



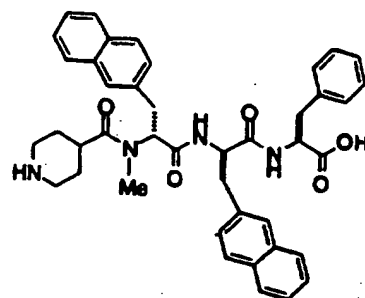
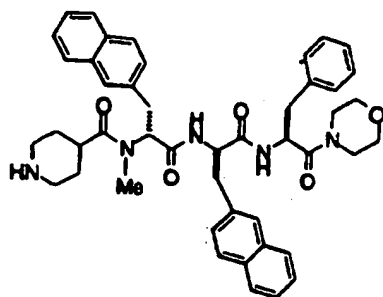
25



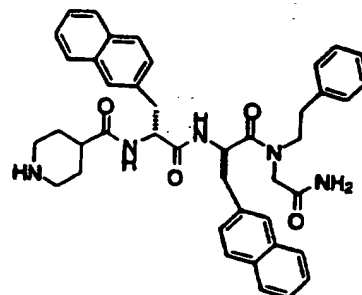
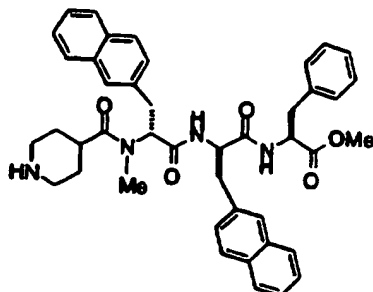
30

35

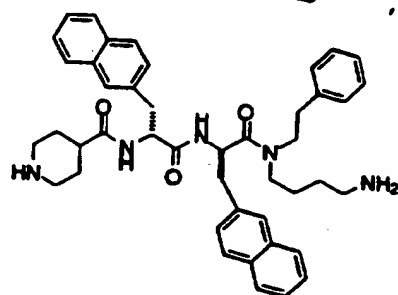
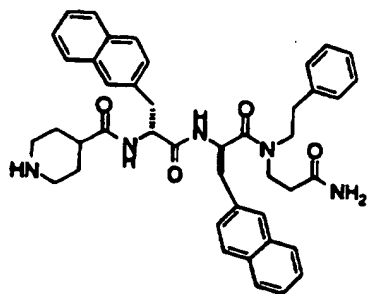
5



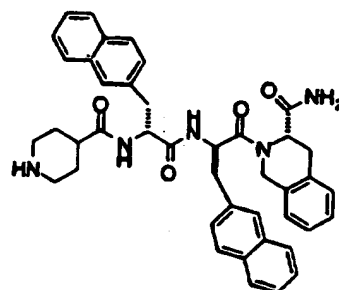
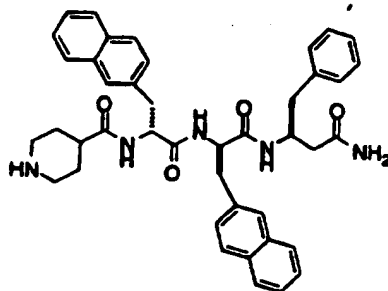
10



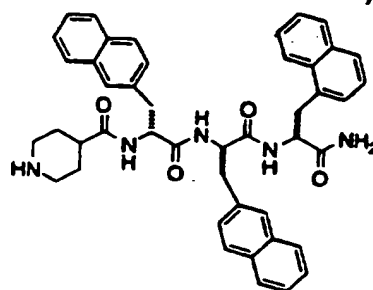
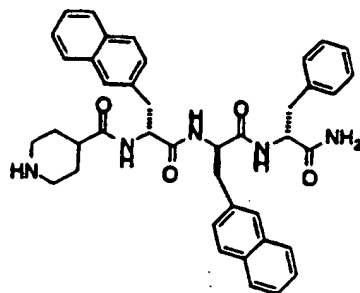
15



20



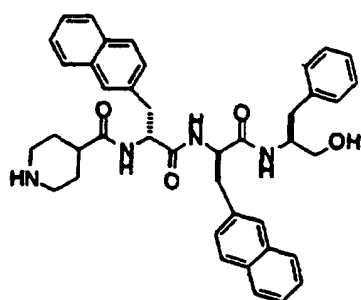
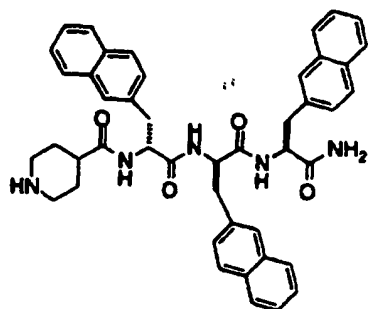
25



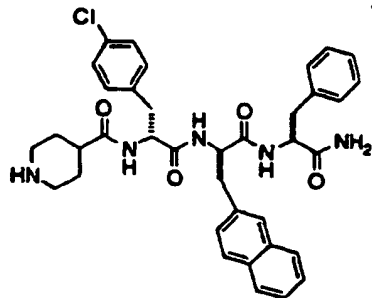
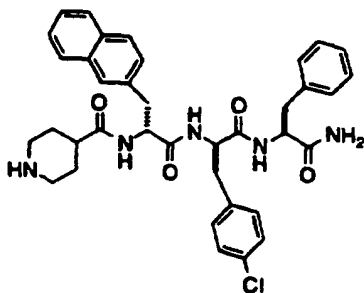
30

35

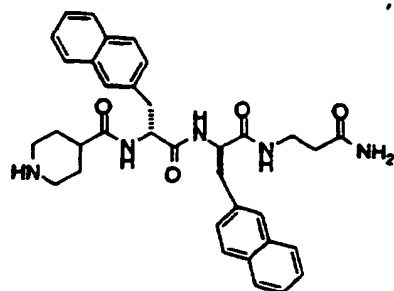
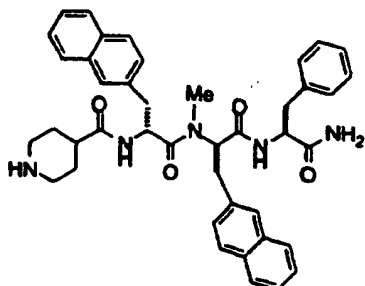
5



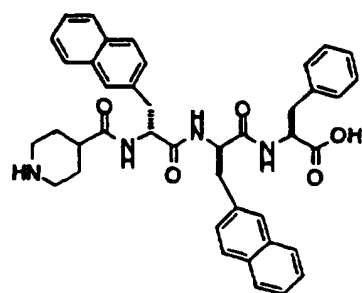
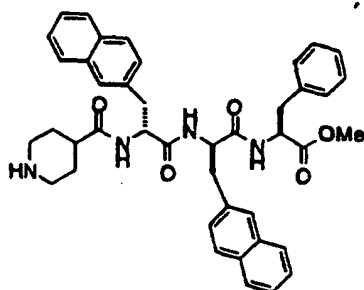
10



15

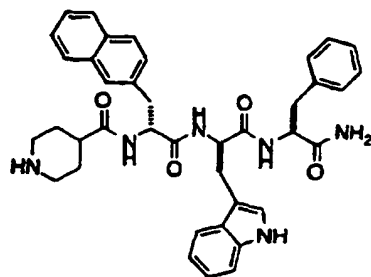
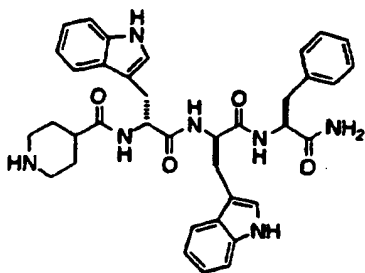


20



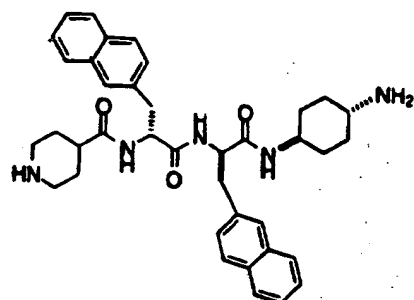
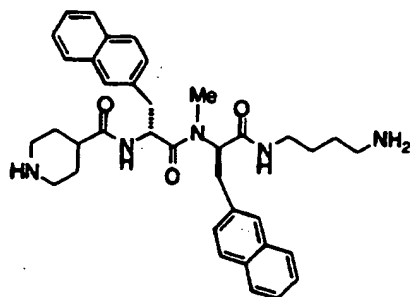
25

30

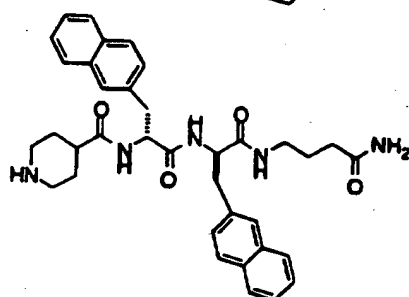
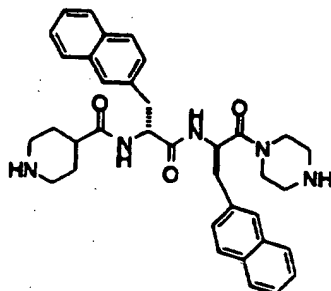


35

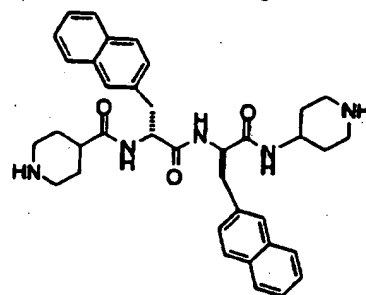
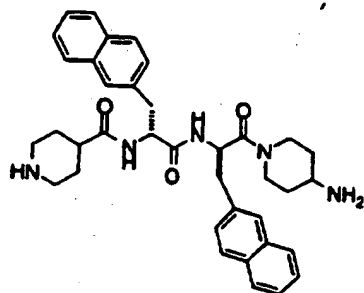
5



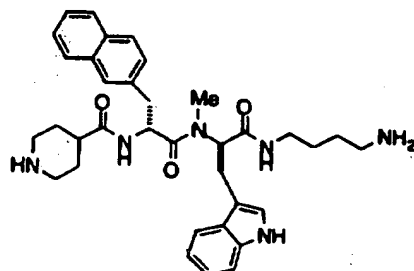
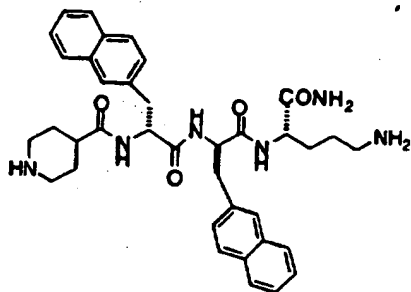
10



15

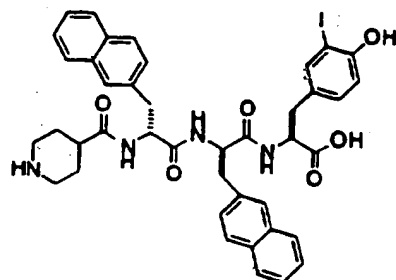
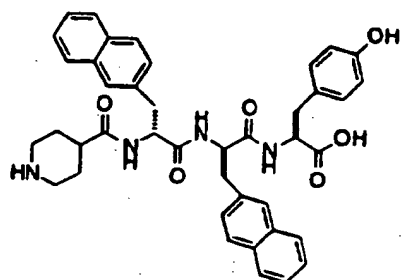


20

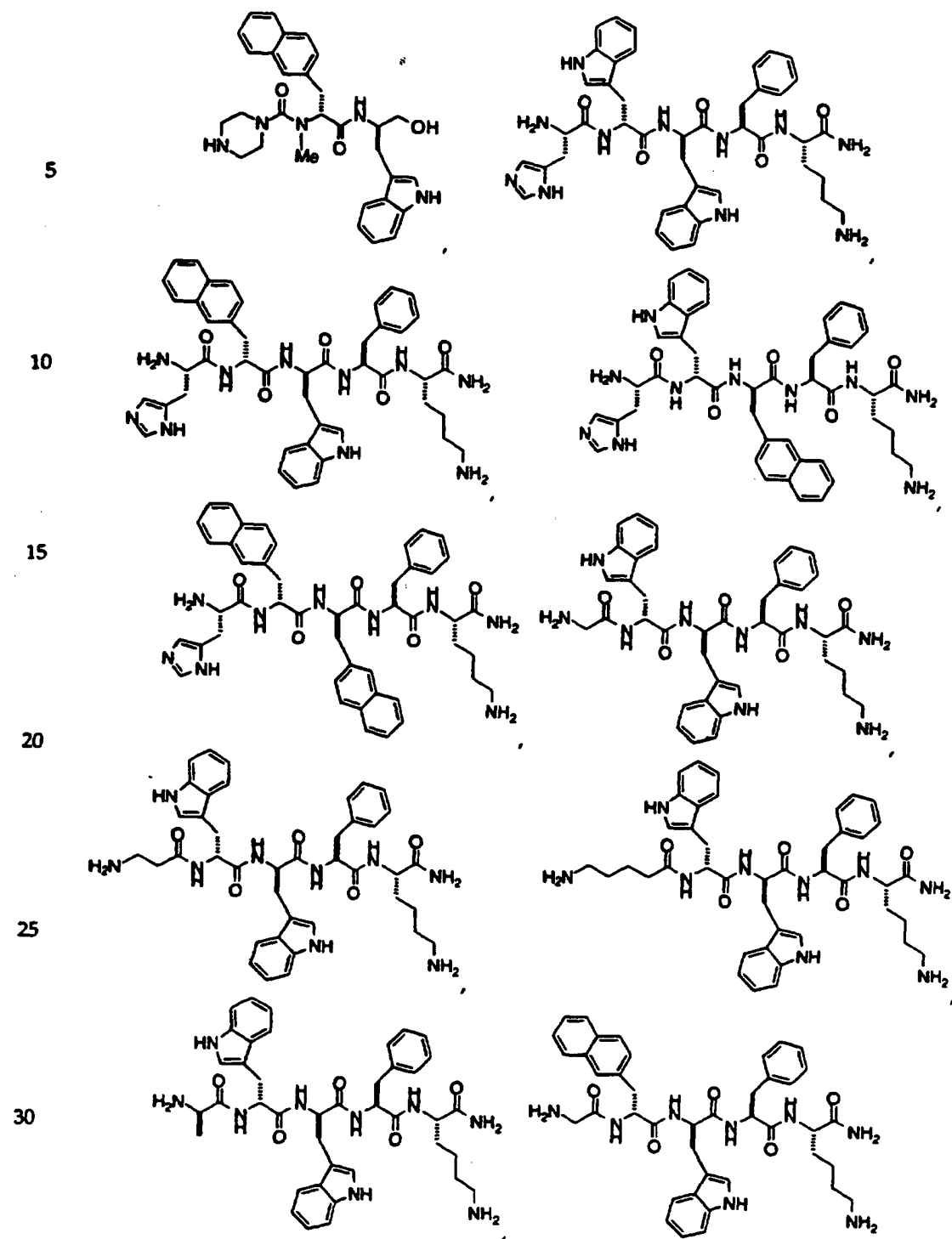


25

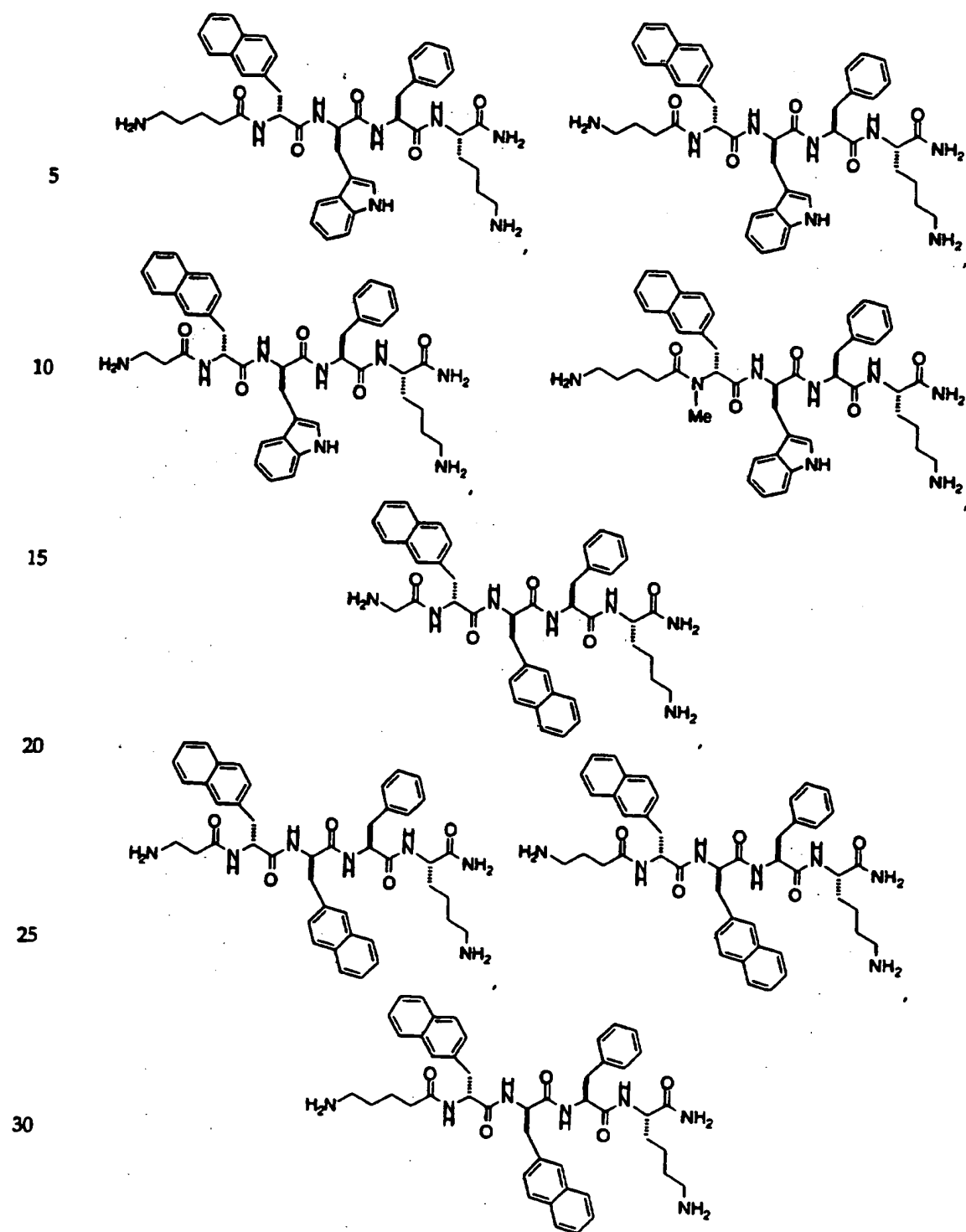
30

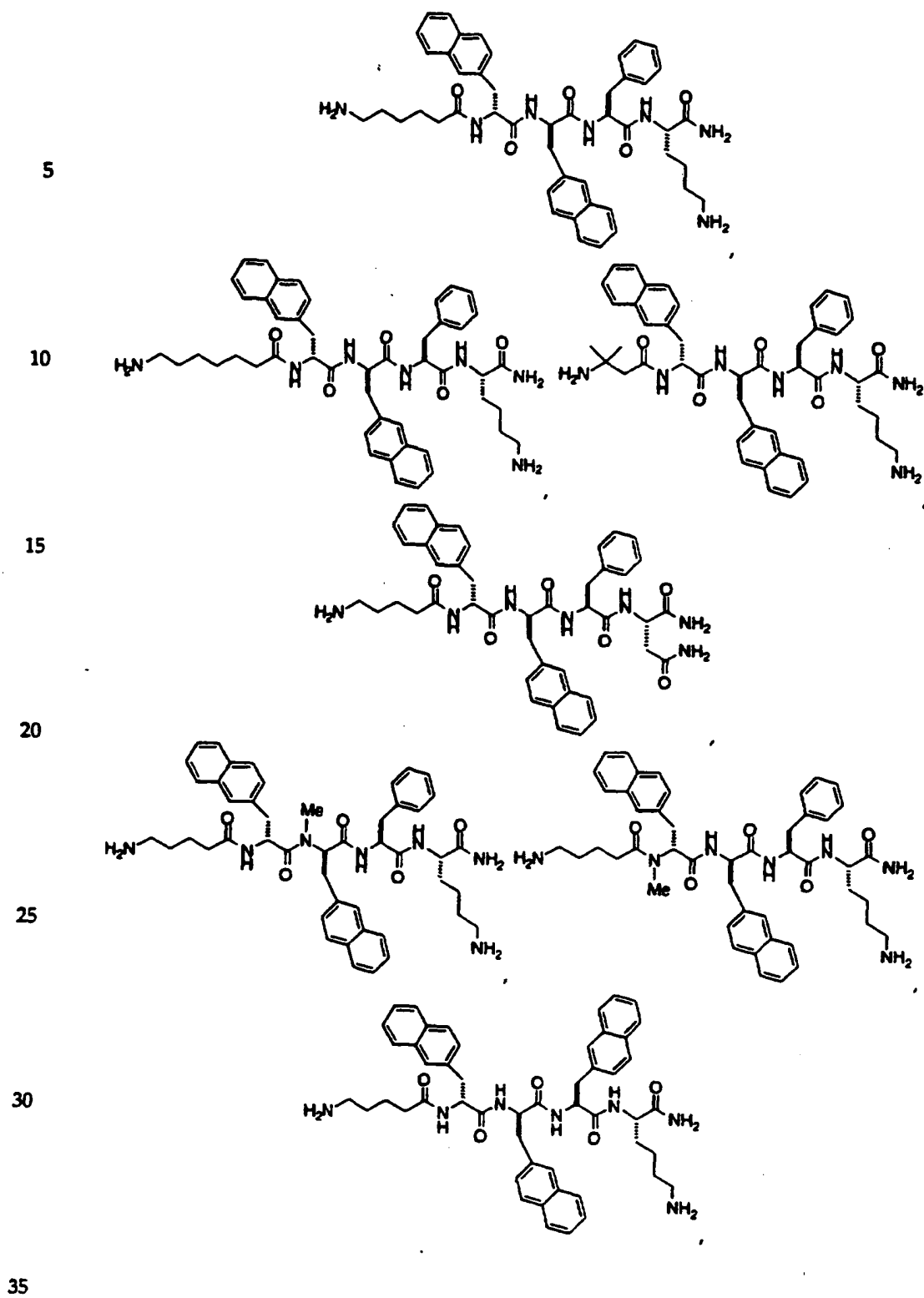


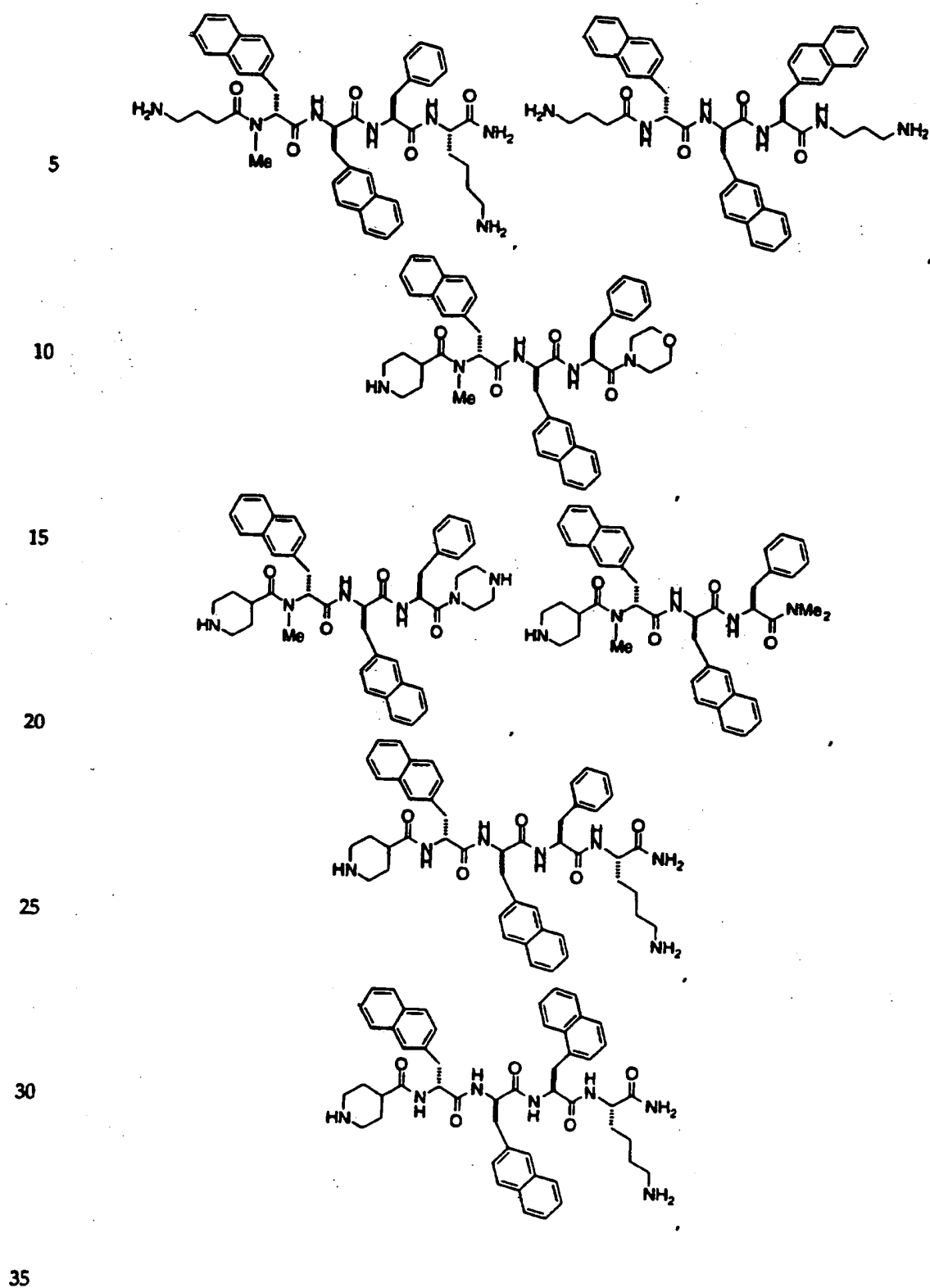
35



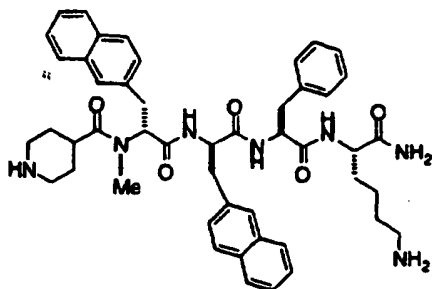
35



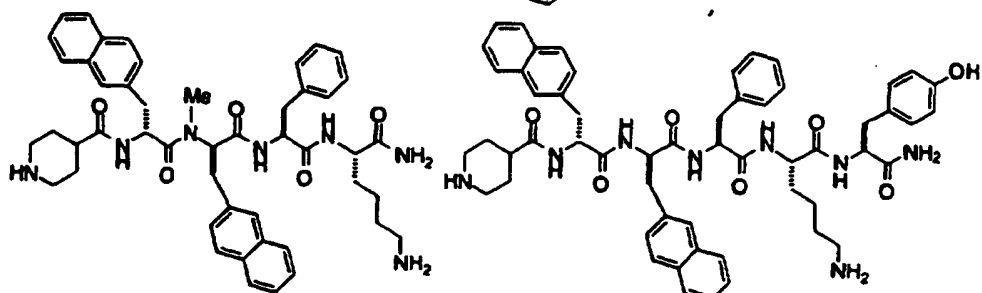




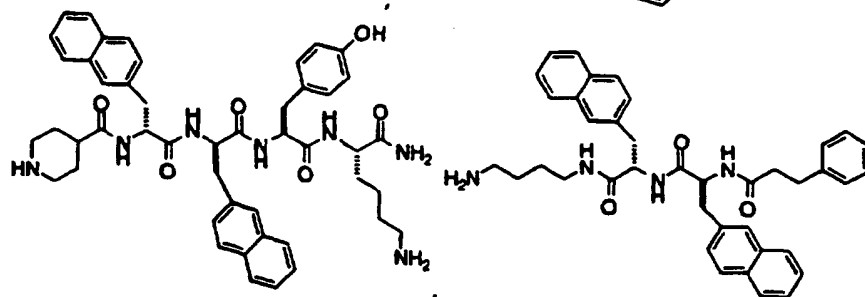
5



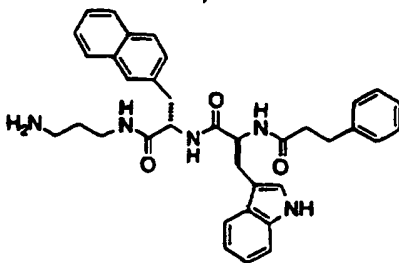
10



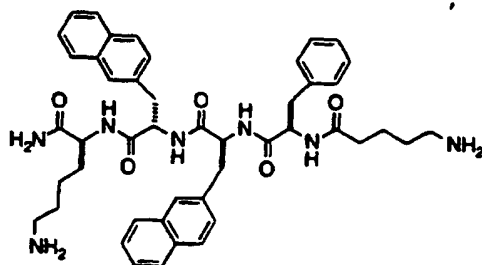
15



20



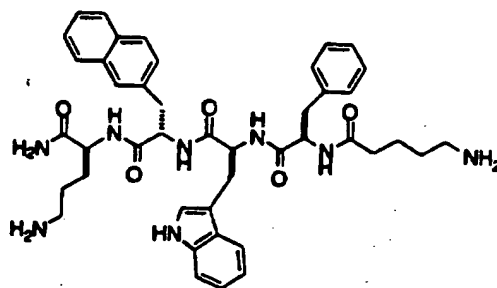
25



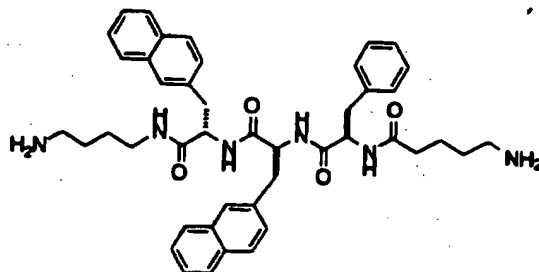
30

35

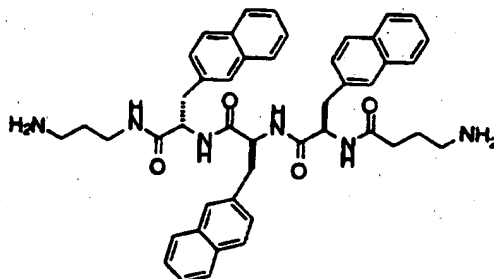
5



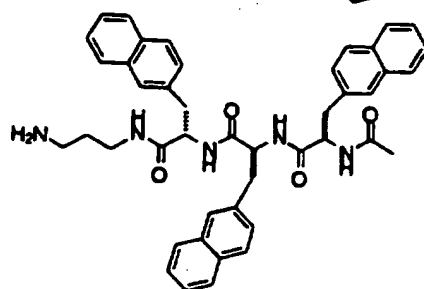
10



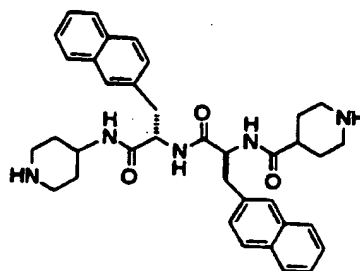
15



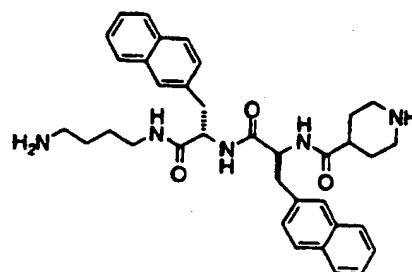
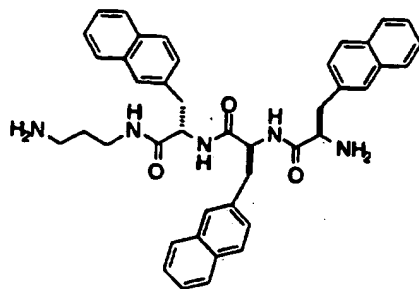
20



25

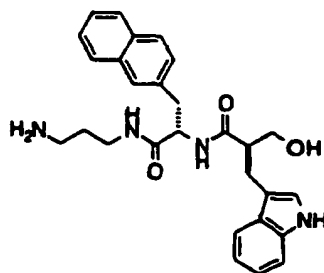
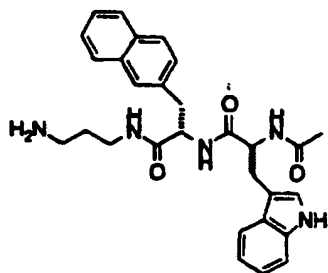


30

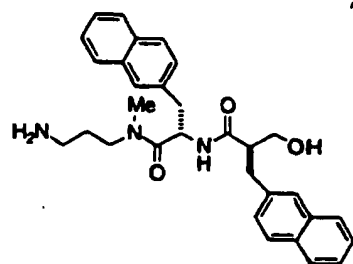
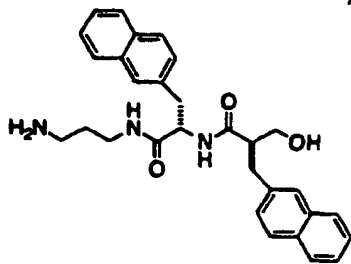


35

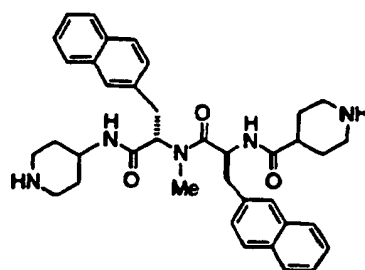
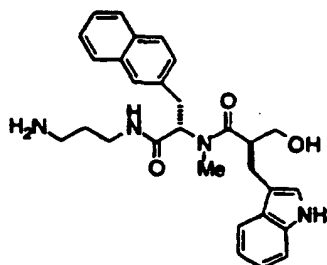
5



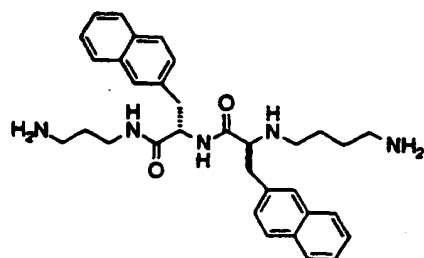
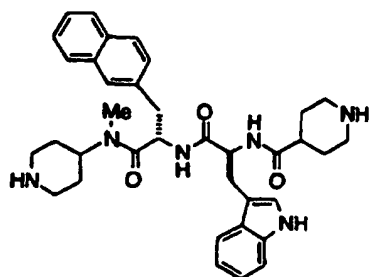
10



15

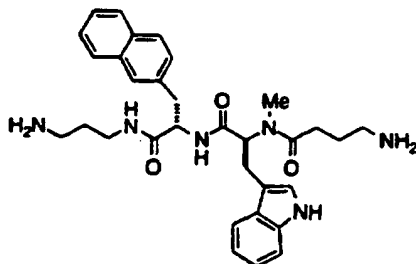


20



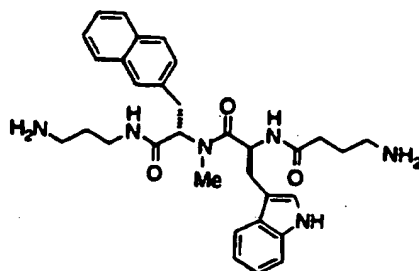
25

30

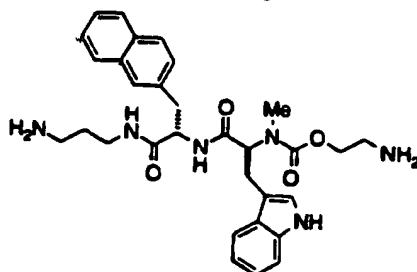


35

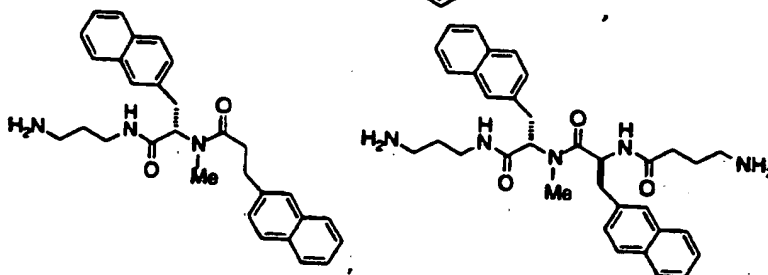
5



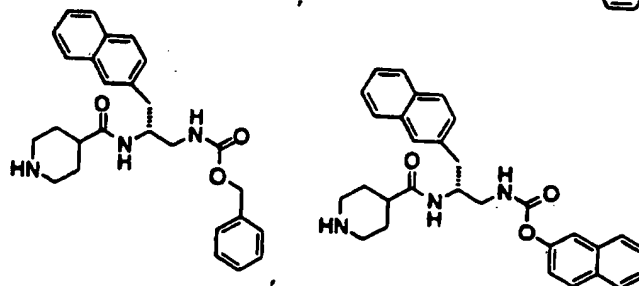
10



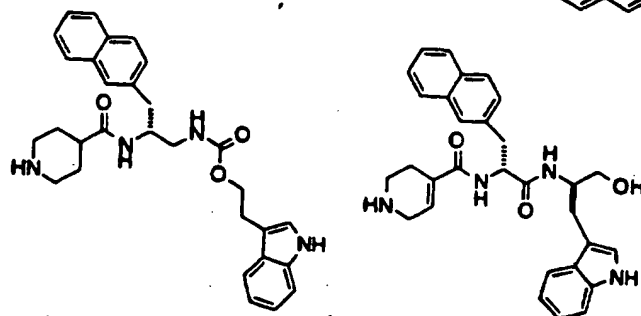
15



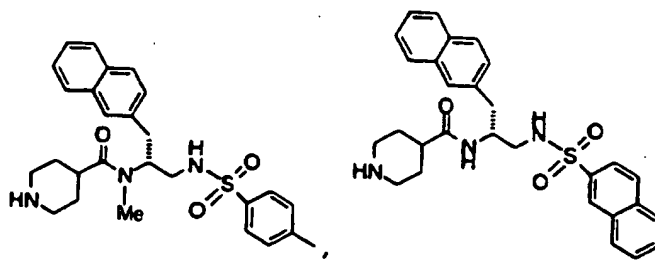
20



25

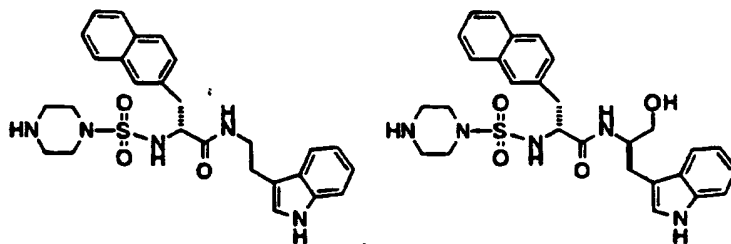


30

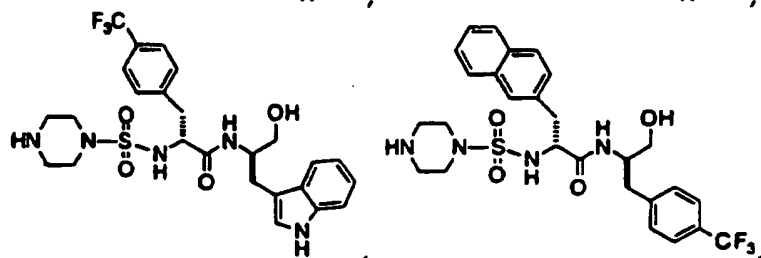


35

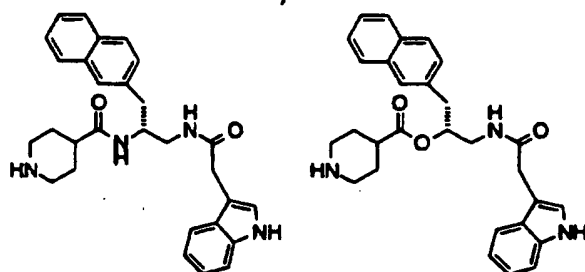
5



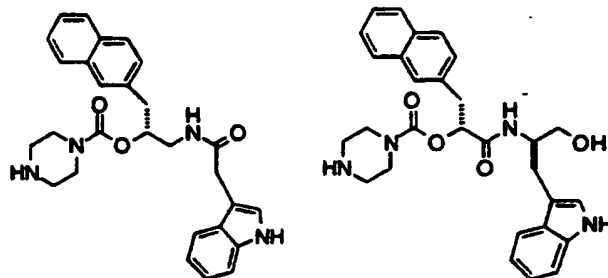
10



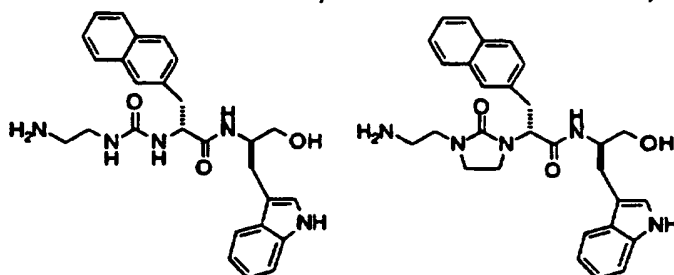
15



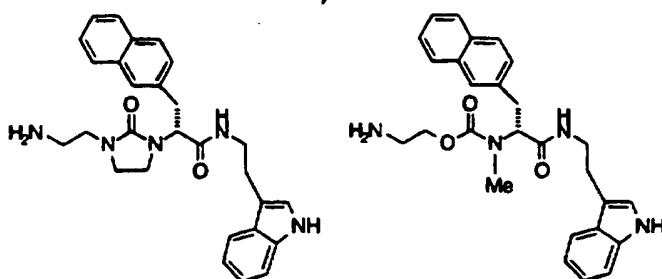
20



25

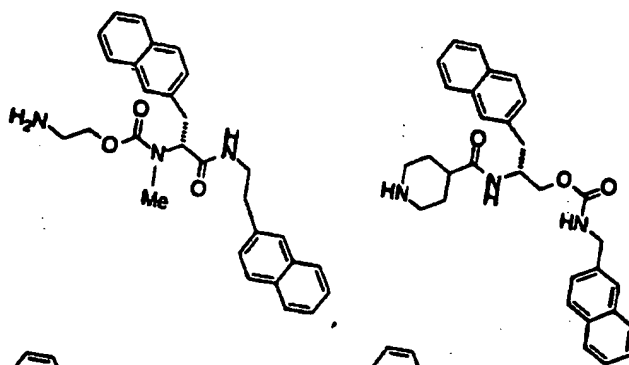


30

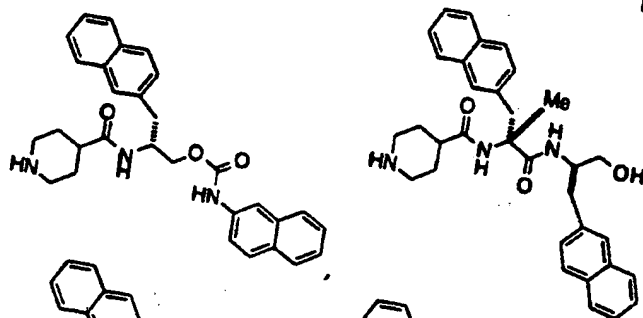


35

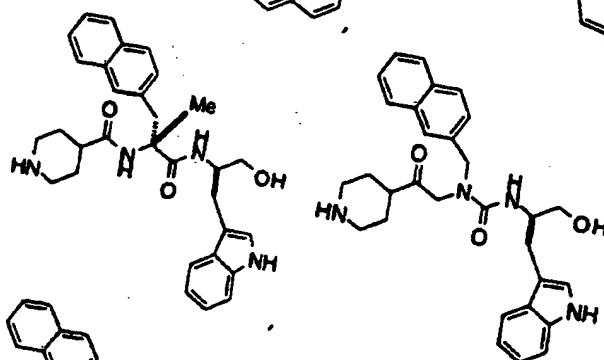
5



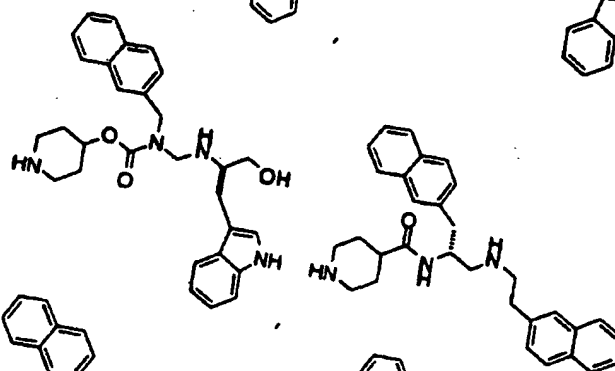
10



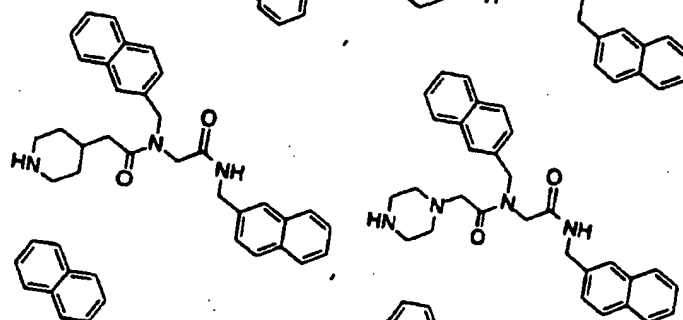
15



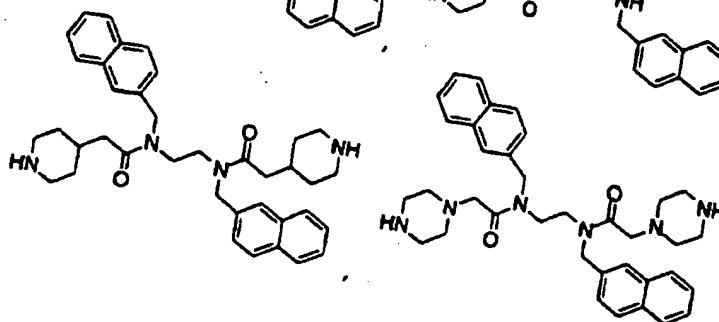
20



25

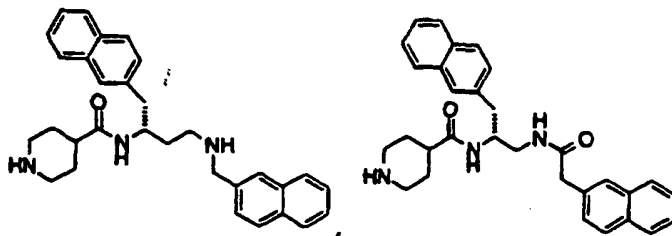


30

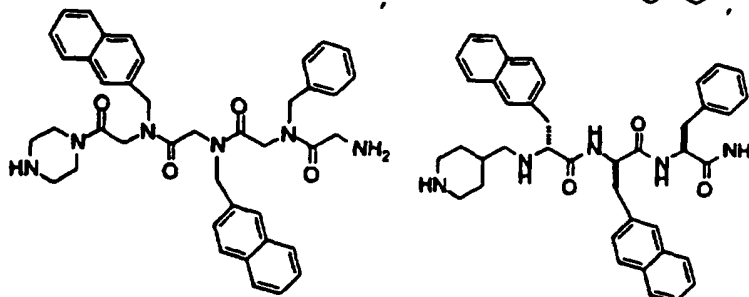


35

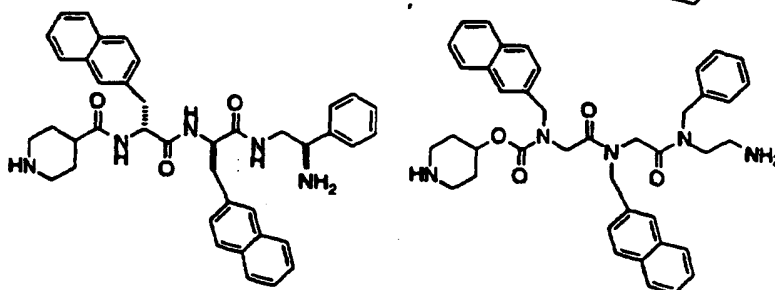
5



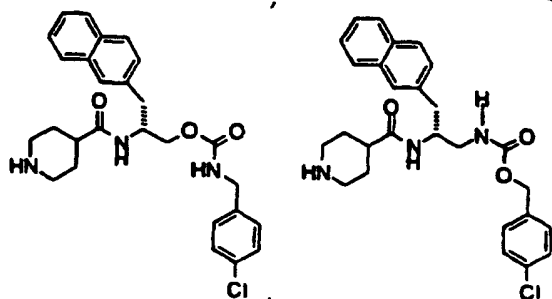
10



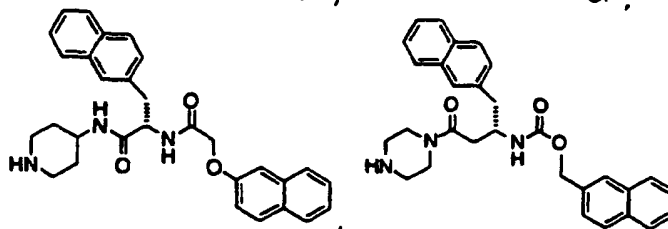
15



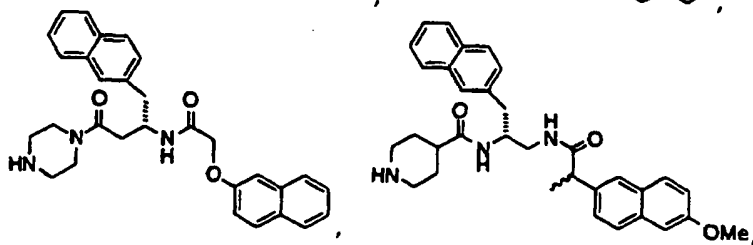
20



25

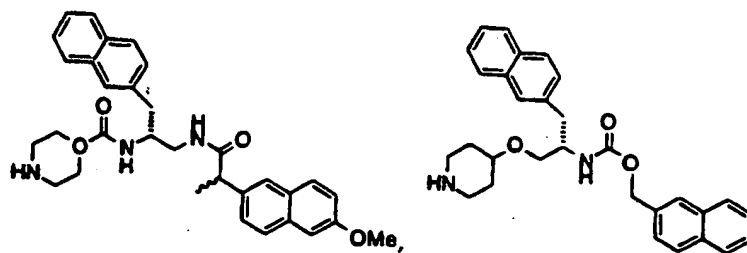


30

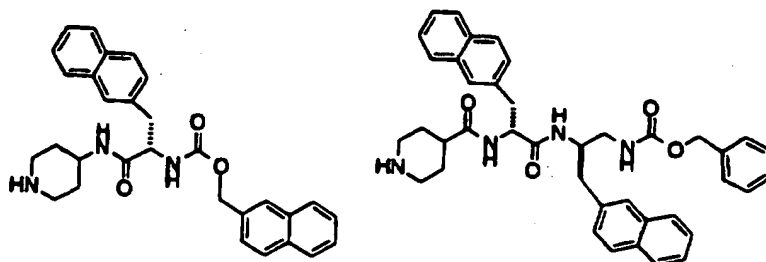


35

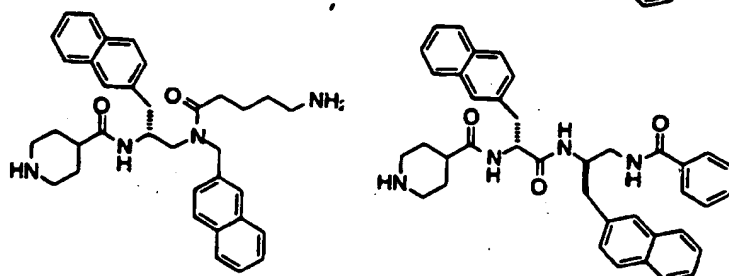
5



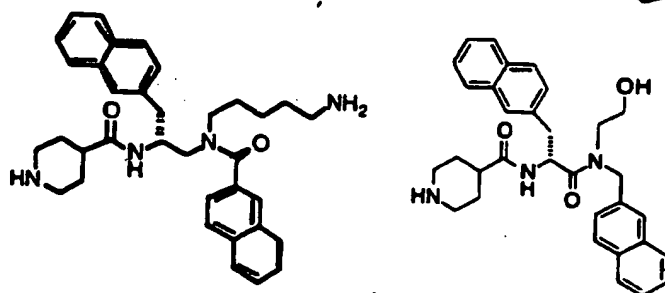
10



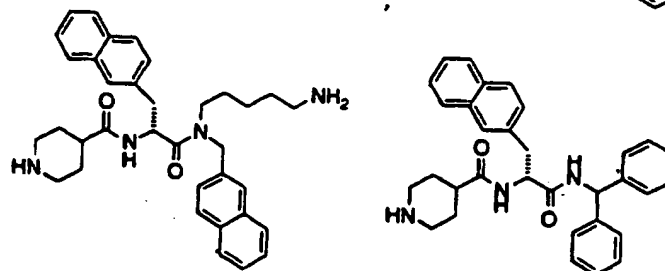
15



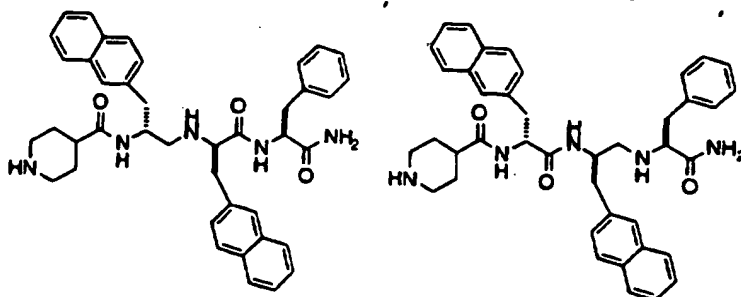
20



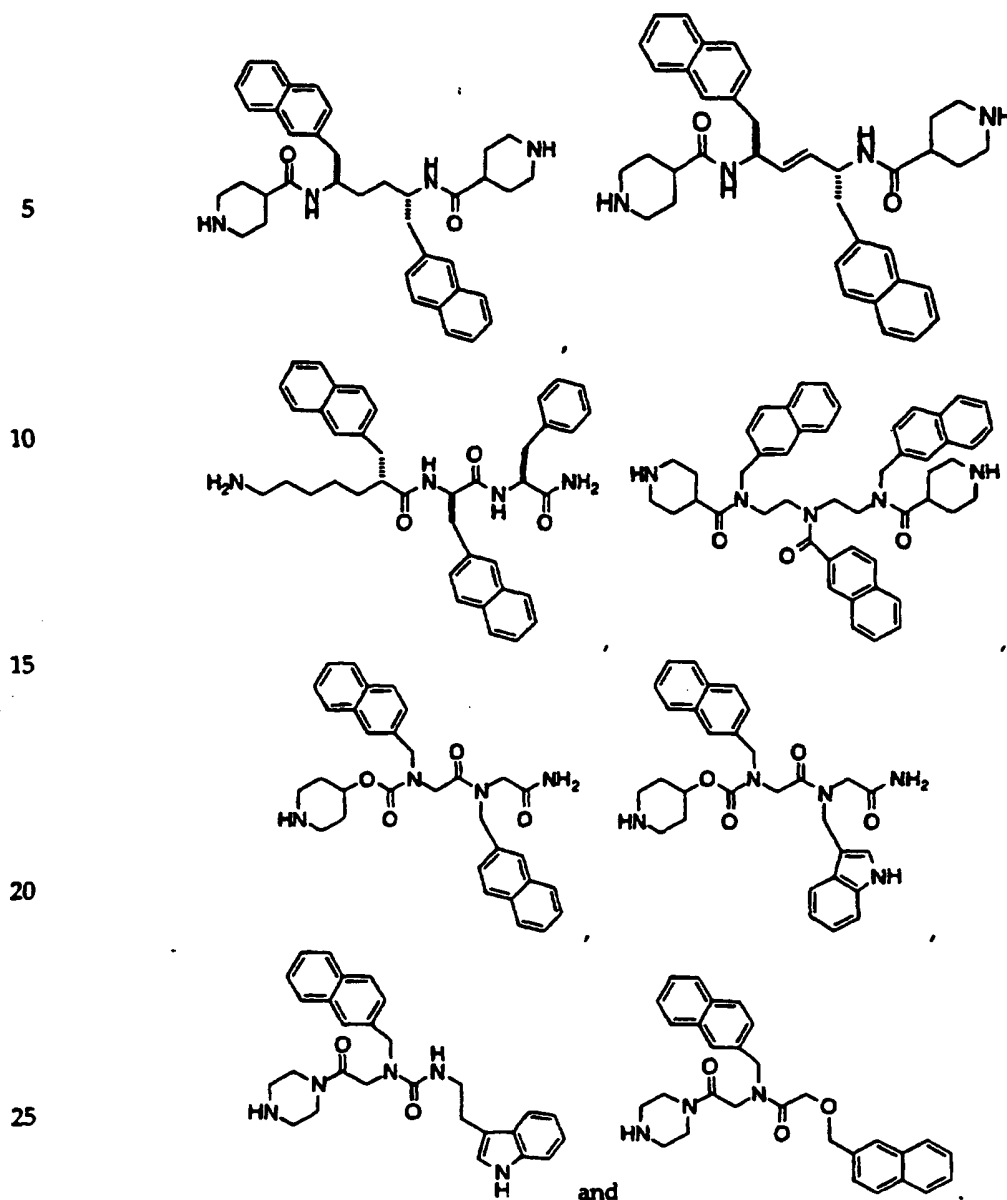
25



30



35



E. Biological Activity

1. *In Vitro* Activity

A. *In Vitro* EC₅₀

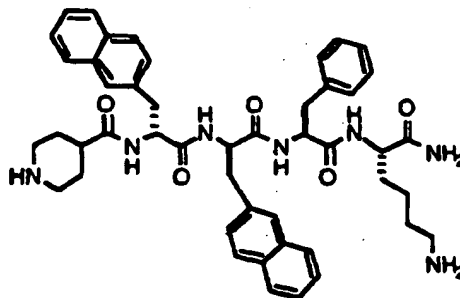
The "pit" EC₅₀ values for all GHRPs were determined by the GH dose-response to the GHRP using the rat pituitary monolayer culture system detailed in Example 38. The results are provided in Tables II - VI in Example 39. Table II details selected biological data for prior art compounds including GHRP-6 with a "pit cell" EC₅₀ of 6.2 ± 1.5 nM (n=5). Table III details selected biological data from 64 compounds from formula IV. Included in this novel class of compound, which is significantly smaller than GHRP-6, is (inip)-bbFK-NH₂ with an EC₅₀ of 0.18 ± 0.04 , over 30-fold more potent than GHRP-6. Table IV details selected biological data

from 63 compounds derived from formula III, including (inip)bb(feg) with an EC_{50} of 0.25 ± 0.19 ($n=3$); almost 25-fold more potent than GHRP-6. Table V details selected biological data from 23 compounds from formula II including (inip)b(wol) ($EC_{50} = 10.6 \pm 6.2$; $n=3$) with a EC_{50} roughly equivalent to GHRP-6. Table VI details selected biological data from "retroinverso" compounds including the most potent, (Ab)bBB(ram), with an EC_{50} of 2 nM ($n=2$).

B. *In Vitro* Characterization

In addition, representatives from novel classes of GHRP were further characterized *in vitro* to determine whether these compounds were acting in a manner analogous to "GHRP-6". The representatives include: from formula IV (inip)-bbFK-NH₂, from formula III (inip)bb(feg) and from formula II (inip)b(wol). The characterization results are detailed below. All experiments had a minimum of three replicates.

1. Representative from Formula IV.



(inip)-bbFK-NH₂

A representative dose response for GH release in the rat "pit" cell assay over a 15 min. exposure to increasing concentrations of (inip)-bbFK-NH₂ is demonstrated in Figure 2 (see Example 38 for all assay protocols). GH release is significantly ($P < 0.05$) increased at 0.3 nM and reaches a plateau by 1 nM with an EC_{50} of 0.16 nM. The mean ($n=3$) EC_{50} for (inip)-bbFK-NH₂ was 0.18 ± 0.04 nM, over 30-fold more potent than (GHRP-6) (6.2 ± 1.5 nM; $n=5$).

To demonstrate that the novel (inip)-bbFK-NH₂ was acting in a manner analogous to "GHRP-6", challenges were carried out using combinations of (GHRP-6), (inip)-bbFK-NH₂ and GHRH (see Figure 3). GHRP-6 and (inip)-bbFK-NH₂ (100 nM) both caused 3-fold increases in GH levels, but no additional increase was observed when these two GHRP's were used in combination. This suggests that both GHRP's may act at the same site. In contrast, both GHRP's showed synergy with GHRH in the rat "pit" cell assay indicating that neither GHRP acts via the GHRH receptor.

A desensitization effect on the putative GHRP receptor was observed when cells were sequentially challenged with fresh (inip)-bbFK-NH₂ every 15 min. (Figure 4). The GH release was decreased after the second 15 min. incubation (total 30 min. exposure to (inip)-bbFK-NH₂) and no significant GH release compared to control occurred during the final (inip)-bbFK-NH₂ challenge. However when GHRH (10 nM) was added to the next 15 min. incubation, a

significant GH response occurred, consistent with the model of separate receptors for GHRH and the GHRP's.

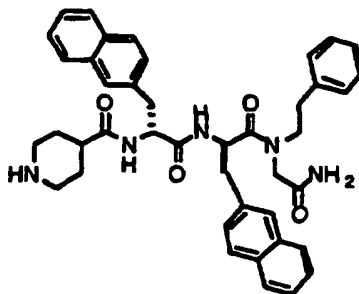
Somatostatin is known to suppress GHRP-stimulated GH release. At 1, 10 and 100 nM (inip)-bbFK-NH₂ significant elevations of GH were observed. Somatostatin (20nM) 5
coincubation with (inip)-bbFK-NH₂ at the same concentrations suppressed this enhanced release (Figure 5).

Other evidence that (inip)-bbFK-NH₂ evokes the GHRP receptor includes the response to the GHRP receptor antagonist HwkWfK. As demonstrated in Figure 6, while high doses (10 μM) were required, HwkWfK did antagonize (inip)-bbFK-NH₂.

10 The specificity of (inip)-bbFK-NH₂ *in vitro* was demonstrated in that LH, FSH, TSH or ACTH release was unchanged by 100 nM (inip)-bbFK-NH₂. Prolactin concentrations were significantly increased but less than 2 fold (Figures 7-9).

Ca⁺⁺ flux determinations are shown in Figure 10. The left panels (A and C) show the basal Ca⁺⁺ pattern immediately prior to addition of vehicle or (inip)-bbFK-NH₂. Twenty-one 15
(21) seconds after (inip)-bbFK-NH₂ application the increased Ca flux in panel D is demonstrated by lighter colors in some of the cells of this heterologous population. As a control, addition of vehicle did not alter the Ca⁺⁺ profile (B), but addition of the calcium secretagogue ionomycin resulted in maximal stimulation of all the cells (data not shown).

2. Representative from Formula III



(inip)bb(feg)

20 This novel GHRP is smaller by about one (lysine) amino acid residue compared to (inip)-bbFK-NH₂ described above. Dose dependent GH release with (inip)bb(feg) is shown in Figure 11. GH release by rat pituitary cells to increasing concentrations of (inip)bb(feg) (left panel) over a 15 minute incubation shows a dose dependent increase. The right panel shows 25
data points and curve used to calculate the EC₅₀ of 0.09 nM for the GHRP (inip)bb(feg). The mean EC₅₀ (n=3) for (inip)bb(feg) was 0.25 ± 0.19 nM.

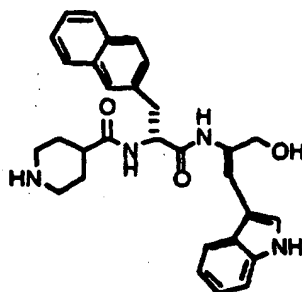
To demonstrate that (inip)bb(feg) also acts at the proposed "GHRP receptor", GH response to GHRP-6 (100nM) and (inip)bb(feg) (100 nM) was measured as shown in Figure 12. The combination of these two GHRP's produced a GH release significantly greater than the 30
control but GH release was not synergistic when these GHRP's were added in combination.

GHRH (100 nM) elicited a mild GH response which was synergistic in combination with either GHRP-6 or (inip)bb(feg) indicating these GHRP's evoke a receptor different from GHRH.

Somatostatin suppression of (inip)bb(feg)-stimulated GH release is shown in Figure 13. GH release with 100 nM (inip)bb(feg) was totally suppressed in the presence of 20 nM somatostatin consistent with the SS suppression response of other GHRP's.

Similarly, desensitization of the "GHRP receptor" upon challenging rat pituitary cells with three sequential 15 min. incubations with fresh (inip)bb(feg) is demonstrated in Figure 14. GH release from the same pituitary cells over three sequential 15 minute incubations with (inip)bb(feg) (100 nM) demonstrated a classic GHRP desensitization pattern. After a total of 45 minutes, GH release was markedly decreased in response to (inip)bb(feg) but these cells were able to release more GH in response to a final 15 minute incubation with GHRH (10 nM).

3. Representative from Formula II



(inip)b(wol)

This novel GHRP is still smaller than (inip)bb(feg), containing only two aromatic residues (b-wol) compared to the three for (inip)-bbFK-NH₂ and (inip)bb(feg). A dose dependent GH release with (inip)b(wol) is demonstrated in Figure 15. GH release by rat pituitary cells to increasing concentrations of (inip)b(wol) (left panel) over a 15 minute incubation shows a dose dependent increase. The right panel shows the data points and curve used to calculate the EC₅₀ of 3.9 nM for the GHRP (inip)b(wol). The mean EC₅₀ (n=3) for (inip)b(wol) was 10.6 ± 6.2 nM.

Again, to demonstrate that (inip)b(wol) acts at the proposed "GHRP receptor", GH response to GHRP-6 (100nM) and (inip)b(wol) (100 nM) was measured as shown in Figure 16. GH release with there two GHRP's was significantly greater than control but not synergistic. In contrast, GHRH (100 nM) elicited a mild GH response which was synergistic in combination with either GHRP-6 or (inip)b(wol).

Somatostatin suppression of (inip)b(wol)-stimulated GH release is demonstrated in Figure 17. GH release to 100 nM (inip)b(wol) was totally suppressed in the presence of 20 nM somatostatin consistent with the SS suppression response of other GHRP's.

Finally, the desensitization effect of the "GHRP receptor" upon challenging rat pituitary cells with three sequential 15 min. incubations with fresh (inip)b(wol) is shown in Figure 18. GH release from the same pituitary cells over three sequential 15 minute incubations with

(inip)b(wol) (100 nM) demonstrated a classic GHRP desensitization pattern.. After a total of 45 minutes, no significant release of GH was observed in response to (inip)b(wol) but these cells were able to release GH in response to a final 15 minute incubation with GHRH (10 nM).

4. Summary of *In Vitro* Characterization

Clearly, by the functional assays demonstrated herein, representatives from each novel class of compound (Formulas II, III, IV and V) elicit GH release in a manner analogous to "GHRP-6". All three classes of compounds released GH in a dose-dependent manner, were synergized by GHRH but not GHRP-6, and had receptor desensitization after continuous exposure to the GHRP while maintaining the ability to respond to GHRH; all consistent with these compounds working through the putative "GHRP receptor". These assays are well-accepted and have been used extensively in the literature. (Cheng *et al.*, *Endocrinology* 132:2727-2731 [1993]; Blake and Smith, *Journal of Endocrinology* 129:11-19 [1991]; Cheng *et al.*, *Endocrinology* 124:2791-2798 [1989]; Smith, *Science* 260:1640-1643 [1993]; and Akman *et al.*, *Endocrinology* 132:1286-1291 [1993]).

Additionally, a representative from Formula IV, (inip)bbFK-NH₂, was further characterized and demonstrated a selective GH release from heterogeneous pituitary cells (excepting a mild, but significant increase in prolactin), ability to release GH via a Ca⁺⁺ flux mechanism, and to be inhibited by a GHRP antagonist. (Bowers *et al.*, *Endocrinology* 128:2027-2035 [1991]).

These data are consistent with the view that all four novel classes of GHRP's elicit GH release in a manner analogous to "GHRP-6", and thus may operate via the same mechanism to release GH *in vitro*.

2. *In Vivo* Activity in Normal Rats

To determine if the new GHRP molecules showed efficacy *in vivo*, young (90 day old) and adult (120 day old) rats were treated with the GHRP's of this invention and GHRH according to the protocols in Examples 41 and 44. Rat GHRH, which has been shown to increase body weight in normal young female rats was used as a positive control in these experiments (Clark and Robinson, *Nature* 314:281-283 [1985]).

A. Body Weight Gain in Normal Rats

Body weight gains plotted against time for the 5 treatment groups are shown in Figure 19. Both (inip) b b F K-NH₂ and rat GHRH induced significant body weight gain compared to the vehicle excipient rats. By Day 5 of treatment the weight gains of all the treated groups were statistically significantly greater than the excipient treated rats. The dose-related nature of the body weight gains to (inip) b b F K-NH₂ is presented in Figure 19. The weight gain in response to the much larger dose of rat GHRH (600 µg/day) was very similar to that induced by the medium dose (20 µg/day) of (inip) b b F K-NH₂. In addition, as little as 4 µg/day (equal to 166 ng/hr) of (inip) b b F K-NH₂ induced a significant weight gain in young normal rats. The great

potency of (inip) b b F K-NH₂ can be seen from this comparison with GHRH and from the very low dose required to induce a significant anabolic effect *in vivo*.

Body weight gains plotted against time for the groups of normal adult female rats treated with other GH secretagogues are shown in Figure 26. The groups treated with GHRH and GHRPs 6, (inip)bbF-NH₂ and (inip)b(nmb)(bam) showed significant body weight gain but the group treated with L-692,585 showed no significant weight gain. Although the response to GHRP (inip)bbF-NH₂ appears to be larger than that to the other secretagogues, this difference was not statistically different. The weight gain in response to GHRP (inip)bbF-NH₂ (36 ± 8 g in 7 days) was similar to that to GHRP (inip) b b F K-NH₂ (36 ± 3 g in 14 days) shown in Figure 20.

These studies show the various classes of GH secretagogues of this invention have significant anabolic effects in normal rats with intact pituitary function. One prior art molecule with minimal activity was L-692,585 a molecule of relatively low potency both *in vitro* and *in vivo*. It is believed, however, that if a larger amount of this molecule had been given significant anabolic effects would have resulted.

B. Organ Weight Gain in Young Normal Rats

Organs were weighed at sacrifice in these experiments to measure the effects of these treatments on the major organ systems. The pituitary and the kidney weight were not affected by treatment. Spleen weight was increased by high dose (inip) b b F K-NH₂ (100 µg/day) and by GHRH (excipient 532 ± 25 mg; high dose (inip) b b F K-NH₂, 628 ± 26 mg; GHRH 624 ± 23 mg). Heart weight was increased by GHRH treatment ($p < 0.05$) and tended to be increased by high dose (inip) b b F K-NH₂ ($p < 0.10$ but > 0.05) compared to excipient treated controls. The thymus was also increased in weight by both (inip) b b F K-NH₂ and GHRH. Thymus weight in excipient treated rats was 485 ± 21 mg, 584 ± 34 mg in high dose (inip) b b F K-NH₂ treated rats, and 575 ± 39 mg in GHRH treated rats. The liver increased in weight in a dose-dependent manner with (inip) b b F K-NH₂. The weight of the liver in the excipient treated rats was 8.75 ± 0.24 g, and with increasing doses of (inip) b b F K-NH₂ liver weight increased from 9.40 ± 0.37 g to 9.70 ± 0.29 g and 10.14 ± 0.29 g for low medium and high doses of (inip) b b F K-NH₂, respectively. Liver weight was also significantly increased by treatment with GHRH. In these experiments there was no statistically significant increase in epiphyseal plate width with either (inip) b b F K-NH₂ or GHRH treatment.

C. Organ and Body Weight Gain Summary

These experiments shows that (inip) b b F K-NH₂ has a range of anabolic effects in normal young female rats. This anabolic effect was seen by increases in body weight, liver weight, spleen weight and thymus weight, with a tendency for heart weight to also increase compared to excipient treated control rats. The effect of the GHRP (inip) b b F K-NH₂ was also dose related with doses of 4, 20 and 100 µg/rat/day all being effective anabolic doses. In these experiments the two highest doses of (inip) b b F K-NH₂ had equivalent effects.

Therefore as little as 166 ng/hr of (inip) b b F K-NH₂ (for 200 g rats; 0.83 µg/kg/hr) was effective at inducing an anabolic effect.

The dose-related effect of (inip) b b F K-NH₂ on the liver is a good indicator of the amount of GH secretion caused by the GHRP (inip) b b F K-NH₂. Liver growth is particularly sensitive to stimulation by GH, and the increased liver weight is the expected response to an increased secretion of GH caused by both (inip) b b F K-NH₂ and GHRH. Kidney shows a relatively poor growth response to GH treatment; the lack of an effect of (inip) b b F K-NH₂ on this organ is therefore the expected result. Wagner and Scow, *Endocrinology* 61:419-425 (1957); Clark *et al.*, *Endocrinology and Metabolism* 1:49-54 (1994).

The effects of (inip) b b F K-NH₂ on the weight of the thymus and spleen indicate that the instant novel GHRPs would be expected to stimulate immune function. Other studies have shown that GH and IGF-1 can significantly stimulate immune function, so it would be expected that GHRPs of this invention, by increasing GH secretion, would also stimulate immune function (Kelley, *Ann. N.Y. Acad. Sci.* 594:95-118 (1990), Clark *et al.*, *J. Clin. Invest* 92:540-548 (1993).

(inip) b b F K-NH₂ tended to increase cardiac weight, indicating a significant anabolic effect of this GHRP on the heart. GH and IGF-1 have been shown to be efficacious in animal models of congestive heart failure, and there is data that GH is effective in humans at improving cardiac function in growth hormone deficient adults. This data suggests that (inip) b b F K-NH₂ would also be effective at improving cardiac function and in the treatment of cardiac congestive heart failure (Sacca *et al.*, *Endocrine Reviews* 15:555-573 [1994]).

The GHRP (inip) b b F K-NH₂ was also effective at stimulating an anabolic response when delivered by continuous infusion. The delivery of (inip) b b F K-NH₂ by SC infusion is an effective treatment and a similar effect could be achieved by any method that maintained a near continuous exposure of the instant GHRP's. For example, oral delivery, transdermal patch, or other delivery systems designed to maintain a continuous exposure to these GHRP's would be appropriate.

D. Comparison of GHRP Infusion Versus Injections in Normal Rats

Body Weight Gain: The GHRP (inip) b b F K-NH₂ at 20 and 100 µg/day, delivered by both injection and infusion, induced significant body weight gain compared to excipient treated rats (see Example 42). By Day 2 of treatment, the weight gains of all the treated groups were statistically significantly greater than the excipient treated rats. The dose-related nature of the body weight gains to injections of (inip) b b F K-NH₂ can be seen in Figure 20. In contrast there were similar weight gains in response to infusions of 20 and 100 µg/day of (inip) b b F K-NH₂. In addition, there were very different patterns of weight gain in response to infusions or injections of (inip) b b F K-NH₂ as can be seen in Figure 21.

Figure 21 also shows an initial rapid weight gain over 2 days in response to the high dose of infused (inip) b b F K-NH₂ (100 µg/d), followed by a marked absence of any further

response for the duration of the experiment. This provides evidence that continuous exposure to high doses of GHRPs can induce tachyphalaxis and thus function as GH antagonists.

In contrast to infusions, periodic injections of (inip) b b F K-NH₂ maintained a significant growth response. Twice daily injections of 10 µg of the GHRP (inip) b b F K-NH₂ produced a large (30 gram) weight gain in adult female rats.

Organ Weights: Organ weights at sacrifice were measured to determine the effects of injection verses infusion of (inip) b b F K-NH₂ on major organ systems. The eviscerated and skinned carcass was significantly heavier in the high dose (152.4 ± 2.5 g) and low dose (152.8 ± 2.8 g) (inip) b b F K-NH₂ injected animals compared to controls (141.8 ± 3 g). The skin was heavier in animals injected with (inip) b b F K-NH₂ and in those infused with low dose (inip) b b F K-NH₂ compared to control animals. When the skin and carcass weight were expressed as a percentage of body weight there was no significant effect due to treatment, indicating that the weight gain was due to a proportional increase in the whole body size of the rats. Soleus muscle, kidney, and liver were also unaffected by treatment. Injections of high dose (inip) b b F K-NH₂ significantly increased heart weight compared to controls (1.15 ± 0.10 g vs. 0.96 ± 0.03 g). Thymus weight was increased by high dose injections of (inip) b b F K-NH₂ (0.33 ± 0.02 g) compared to high dose infusion animals (0.25 ± 0.03 g). Epiphyseal plate width was increased by high dose (inip) b b F K-NH₂ injections (191 ± 8 µm) compared to low dose injections (160 ± 11 µm) whereas (inip) b b F K-NH₂ infusions did not significantly increase cartilage growth. Serum IGF-1 concentrations were not significantly affected by either mode of delivering (inip) b b F K-NH₂.

Serum chemistries were measured in the blood samples obtained at sacrifice. Enzyme levels indicative of cardiac, liver, muscle and kidney function were measured. There were no statistically significant effects of (inip) b b F K-NH₂. In addition, metabolites (glucose, blood urea nitrogen, creatinine, total protein, albumen, cholesterol, bilirubin) and ions (calcium, phosphate, sodium, potassium and chloride) were measured. The only metabolite showing some evidence of changing was the serum triglyceride. There was some evidence that serum triglyceride was increased by high dose injections of the GHRP (inip) b b F K-NH₂ but not by low dose injections (control 126 ± 11 mg%, high dose 183 ± 17 mg%, low dose 128 ± 11 mg%) although this effect failed to reach statistical significance.

This experiment clearly shows that the GHRP (inip) b b F K-NH₂ when given by either injections or infusions has a range of anabolic effects in normal adult female rats. This growth promoting effect was seen by increases in body weight gain, carcass weight, skin weight, heart weight and thymus weight compared to excipient treated control rats. In this experiment two injections per day of 10 µg of (inip) b b F K-NH₂ caused a weight gain nearly equal to that of two injections per day of 50 µg of (inip) b b F K-NH₂. Therefore 10 µg of (inip) b b F K-NH₂ (for 200 g rats; 50 µg/kg/day) appears to be a maximal dose of (inip) b b F K-NH₂ for inducing an anabolic response. Acute intravenous injection experiments with (inip) b

b F K-NH₂ in 80 gram rats (Example 40) over a 25-fold range of doses (e.g. 1.0, 0.2, and 0.04 µg/injection) demonstrates that induction of GH secretion occurs within this dose-response range (ED₅₀ 0.2µg/rat). In 200 g rats this suggests that a 10µg dose of GHRP (inip) b b F K-NH₂ would be well above the effective doses-responses range.

5 There were differences between the anabolic effects of (inip) b b F K-NH₂ in 90 vs. 150 day old rats. In the older rats there was no clear effect on liver weight or spleen weight as seen with GHRP (inip) b b F K-NH₂ infusion in the younger rats. (inip) b b F K-NH₂ injections increased thymus weight, suggesting that these GHRPs can stimulate growth of immune tissue and therefore increase immune function. Other studies have shown that GH and IGF-1 can significantly stimulate immune function, so it would be expected that GHRP's
10 of this invention, by increasing GH secretion, would also stimulate immune function. Kelley, *Ann. N.Y. Acad. Sci.* 594:95-118 (1990), Clark *et al.*, *J. Clin. Invest.* 92:540-548 (1993). GHRP (inip) b b F K-NH₂ tended to increase cardiac weight, indicating an effect on the heart structure and function. GH and IGF-1 have been shown to be efficacious in models of congestive heart
15 failure and this data suggests that (inip) b b F K-NH₂ would also be effective in the treatment of congestive heart failure. Sacca *et al.*, *Endocrine Reviews* 15:555-573 (1994).

 The GHRP (inip) b b F K-NH₂ was clearly effective at stimulating an anabolic response when delivered by both injections and continuous infusion. The delivery of (inip) b b F K-NH₂ by twice daily subcutaneous injection appears to be an effective method of GHRP
20 delivery. Other methods of delivery that would produce a similar blood profile of GHRP, for example oral delivery, delivery to the lung, or other delivery systems designed to maintain an intermittent exposure to GHRP would also be an effective means of inducing GH secretion and thereby the effects of GH.

 The normal serum chemistries indicate that the effects of (inip) b b F K-NH₂ can occur
25 without perturbing the normal balance of blood metabolites and ions. The one possible exception was the tendency for serum triglyceride to be increased by high dose injections, but not by low dose injections of the GHRP (inip) b b F K-NH₂. This may indicate that very high doses of (inip) b b F K-NH₂ can impact the ACTH system and induce corticosterone activity. However 10 µg injections of (inip) b b F K-NH₂ did not seem to affect serum lipids, indicating
30 that this dose, while maximally stimulating GH secretion, has a minimal effect on corticosterone secretion.

E. Combination GHRP and IGF-1 Treatment of Normal Rats

 Normal adult female rats were chosen to study the anabolic effect of GHRP's 6, (inip)bbF-NH₂, (inip)b(nmb)(bam), and L-692,585 when given in combination with IGF-1.
35 Details of the protocols for this study are described in Example 44.

 The body weight gain responses to the GH secretagogues given in combination with IGF-1 (Figure 27) were much greater than those to the GH secretagogues given by themselves

(Figure 26). In addition, the responses to the combination of the GH secretagogues and IGF-1 tended to be greater than to IGF-1 alone.

This study shows for the first time that GHRP has significant anabolic activity when given in combination with chronically administered IGF-1. Furthermore, there is an additional anabolic benefit of administering the combination of GH secretagogues and IGF-1.

3. *In Vivo* Activity in ZDF Rats

A. Combination GHRP and IGF-1 Therapy in Obese Rats

It is known that IGF-1 inhibits GH secretion by a feedback mechanism either acting indirectly on the hypothalamus or directly on the pituitary. Tannenbaum *et al.*, *Science* 220:77-79 (1981). It is also known that GHRH induced GH secretion is suppressed by IGF-1 administration. (Bermann *et al.*, *Program and Abstracts 76th Annual Meeting US Endocr. Soc.*, Abstract 565, (1994). It was however unknown if GHRP could induce GH secretion, or produce effects, in combination with IGF-1 administration. The protocol for IGF-1 administration in combination with GHRP and GH is provided in Example 43. Rat GHRH, which has been shown to increase body weight in normal female rats was used as a positive control in the experiment.

Body Weight Gain: The body weight gains plotted against time for all treatment groups over the entire study (i.e. 24 days) are shown in Figure 22. Body weight gains for the first 7 days for the GHRP (inip) b b F K-NH₂ and IGF-1 treatment groups are shown in Figure 23. Both (inip) b b F K-NH₂ and rhIGF-1 induced significant body weight gain compared to the vehicle treated rats, so that by Day 2 (see Figure 23) of treatment, the weight gains were statistically significantly greater than the excipient treated rats ((inip) b b F K-NH₂: 18.3 ± 1.0 g, rhIGF-1: 21.5 ± 0.7 g, and obese controls: 13.8 ± 0.4 g). Treatment with rhGH had not increased weight gain by this time (13.6 ± 2.5 g). The weight gain in response to the GHRP (inip) b b F K-NH₂ plus rhIGF-1 (26.8 ± 0.8 g) was greater ($p < 0.05$) than that to the combination of rhGH + rhIGF-1 (23.3 ± 1.2 g). At day 7 of treatment these differences in weight gain were maintained. By day 24 of treatment (Figure 22) the weight gain response to (inip) b b F K-NH₂ plus IGF-1 (247.4 ± 7.1 g) was similar to that to rhGH + rhIGF-1 (245.2 ± 4.9 g) and much greater than that of obese controls (169.0 ± 1.6 g) and significantly greater ($p < 0.05$) than for rhIGF-1 treatment alone (232 ± 2 g). It was surprising that GHRP (inip) b b F K-NH₂ could induce a weight gain when given in combination with IGF-1, and that this weight gain was equal to that of a large dose of rhGH and that the effects of the combination of GHRP and rhIGF-1 gave weight gains greater than or equal to those of the combination of rhGH and rhIGF-1.

B. Combination GHRP and IGF-1 Treatment of Diabetic Rats

Obese Zucker Diabetic Fatty (ZDF) rats were chosen to study the diabetogenic effect of GHRP when given in combination with IGF-1. Details of the protocols for this study are described in Example 43.

Blood glucose: High concentrations of blood glucose were used to define the diabetic state of an animal. Rats were started on treatment before diabetes had developed (there was no difference between lean and obese blood glucose values at day 0). Figure 24 shows the changes with time in fasting blood glucose for the 6 obese groups and the lean controls. By day 14 the obese rats were clearly diabetic (obese controls 218 ± 27 mg%, lean controls 140 ± 3 mg%). Treatment with rhGH gave a greater ($p < 0.05$) increase in blood glucose (504 ± 38 mg%) than did the GHRP (inip)bbFK-NH₂ (386 ± 63 mg%). In addition the combination of GHRP (inip)bbFK-NH₂ plus IGF-1 resulted in a blood glucose value of 190 ± 27 mg% comparable to that for rhGH + IGF-1 treatment of 233 ± 49 mg%.

On Day 24 the blood glucose of the obese diabetic rats had risen to more than twice that of the lean controls (147 ± 4 mg% vs. 330 ± 57 mg%) and values were significantly ($p < 0.05$) higher for rhGH treated rats (725 ± 30 mg%) than for GHRP treated rats (542 ± 37 mg%). This difference between rhGH and GHRP was also observed in combination treatment with rhIGF-1. GHRP plus rhIGF-1 resulted in a lower blood glucose measurement (301 ± 53 mg%) than did rhGH + rhIGF-1 treatment (512 ± 55 mg%). However the glucose values in the GHRP + IGF-1 treated group were elevated ($p < 0.05$) compared to animals receiving rhIGF-1 alone (177 ± 4 mg%).

Serum Insulin: At Day 0 the levels in obese rats were elevated compared to lean controls, but there were no differences between the levels in the obese treatment groups. At week 1 IGF-1 treatment significantly reduced serum insulin (obese control, 21 ± 2 ng/ml; IGF-1 treated, 8 ± 1 ng/ml). By itself GHRP (inip)bbFK-NH₂ elevated serum insulin 39 ± 6 ng/ml as did rhGH treatment (48 ± 6 ng/ml). However the combination of GHRP (inip)bbFK-NH₂ plus IGF-1 significantly lowered insulin (to 10 ± 2 ng/ml) compared to the combination of rhGH plus IGF-1 (25 ± 3 ng/ml, $p < 0.05$). Therefore there was evidence that the combination of GHRP (inip)bbFK-NH₂ and IGF-1 stimulated insulin secretion to a lesser extent (was less diabetogenic) than the combination of GH and IGF-1.

Insulin Sensitivity: At Day 24 the sensitivity of the animals to insulin was gauged by measuring blood glucose 30 minutes after an insulin challenge (Figure 25). Following insulin, absolute blood glucose values were again higher in obese diabetic animals (277 ± 42 mg%) than in lean animals (84 ± 8 mg%). However, blood glucose was now reduced in GHRP treated animals to levels (323 ± 67 mg%) not different from those of the obese controls. In contrast, in rhGH treated rats, glucose remained significantly ($p < 0.05$) elevated (533 ± 55 mg%) compared to obese controls or GHRP treated rats. A similar difference between rhGH and GHRP treatment was seen when they were combined with rhIGF-1 treatment. Blood glucose was significantly ($p < 0.05$) lower in GHRP + rhIGF-1 treated rats (238 ± 47 mg%) than in rhGH + rhIGF-1 treated rats (388 ± 48 mg%).

These experiments compare the anabolic and diabetogenic effects of administered GH and GHRP in a rat model of Type II diabetes. These experiments show for the first time that

administered GHRP's have significant anabolic activity when given in combination with IGF-1. This anabolic activity was equivalent to that induced by treatment with GH plus IGF-1. In contrast, administering GH caused significantly greater insulin resistance, as measured by serum glucose and insulin, and by an insulin challenge, than did administered GHRP, even when they were given in combination with rhIGF-1. This study shows that the diabetogenic effect of GHRP (inip)bbFK-NH₂ was significantly less than that of rhGH, at doses that produced similar anabolic effects.

F. Administration

The present invention also provides compositions containing an effective amount of compounds of the present invention, including the nontoxic addition salts, amides and esters thereof, which may, alone, serve to provide the above-recited therapeutic benefits. Such compositions can be provided together with physiologically tolerable liquid, gel or solid diluents, adjuvants and excipients.

The compounds and compositions can be administered to mammals including humans in a manner similar to other therapeutic agents. The dosage to be administered will depend on the usual factors including; age, weight, sex, condition of the patient and route of administration. In general, the dosage required for therapeutic efficacy will range from about 0.001 to 1000 µg/kg, more usually 0.01 to 2.5 µg/kg of the host body weight. Alternatively, dosages within these ranges can be administered by constant infusion over an extended period of time until the desired therapeutic benefits have been obtained.

Typically, such compositions are prepared as injectable liquid solutions or suspensions. Compositions may also be emulsified. The active ingredient is often mixed with diluents or excipients which are physiologically tolerable and compatible with the active ingredient. Suitable diluents and excipients are, for example, water saline, dextrose, glycerol, or the like, and combinations thereof. In addition, if desired the compositions may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, stabilizing or pH-buffering agents, and the like. For a more detailed description of the foregoing see a standard pharmaceutical text such as *Remington's Pharmaceutical Sciences*, Mack Publishing Co. Easton, PA. (1970).

The compositions of this invention are conventionally administered parenterally by injection, either subcutaneously or intravenously. Additional formulations which are suitable for other modes of administration include suppositories, intranasal aerosols, and, in some cases, oral formulations. For suppositories, traditional binders and excipients may include, for example, polyalkylene glycols or triglycerides; such suppositories may be formed from mixtures containing the active ingredient in the range of 0.5% to 10% preferably 1%-2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, cellulose,

magnesium carbonate, and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained-release formulations, or powders, and contain 10%-95% if active ingredient, preferably 25%-70%.

The peptidomimetic compounds may be formulated into the compositions as neutral or salt forms. Pharmaceutically acceptable nontoxic salts include the acid addition salts (formed with the free amino groups) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or organic acids such as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups may be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropyl amine, 2-ethylamino ethanol, histidine, procaine, and the like.

EXAMPLES

General Experimental:

The compounds synthesized via the routes shown in Scheme I-VI followed standard solid-phase methodologies (Barany, G. and Merrifield, R. B. (1980) in "The Peptides", 2, 1-284. Gross, E. and Meienhofer, J. Eds. Academic Press, New York.).

The chemical name abbreviations for common reagents and unusual amino acids used in the examples below are defined as follows (for the definition of acronyms specific to the Tables of Example 39, see Table I):

20

	α Nal	L- α -Naphthylalanine or (L-3-(1-naphthyl)-Alanine)
	BOC	tert-Butyloxycarbonyl
	BOP	benzotriazol-1-yloxy-tris-(dimethylamino)-phosphonium-hexafluorophosphate
25	BOP-Cl	bis(2-oxo-3-oxazolidinyl) phosphinic chloride
	CBZ	Benzylloxycarbonyl
	β Nal	L- β -Naphthylalanine or (L-3-(2-naphthyl)-Alanine)
	D β Nal	D- β -Naphthylalanine or (D-3-(2-naphthyl)-Alanine)
	DCM	Dichloromethane
30	DIPC	Diisopropylcarbodiimide
	DIPEA	Diisopropylethylamine
	DMF	N,N-Dimethylformamide
	DMA	N,N-Dimethylacetamide
	DMSO	Dimethylsulfoxide
35	EDC	1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride
	Fmoc	Fluorenyloxymethylcarbonyl
	HBTU	[2-(1H-benzotriazol-1-yl)-1, 1, 3, 3-tetramethyluronium]hexafluorophosphate

	HOBt	Hydroxybenzotriazole
	inip	Isonipecotic acid (Piperidine-4-carboxylic acid)
	MBHA	p-methylbenzhydrylamine
	MeOH	Methanol
5	MS	Mass Spectrometry
	N-Me	N-Methyl
	NMM	N-Methyl morpholine
	NMP	N-Methyl pyrrolidinone
	TEA	Triethylamine
10	TFA	Trifluoroacetic acid
	THF	Tetrahydrofuran

Note: Standard three letter codes are used to designate the natural amino acids with a "D" placed before it signifying the dextrorotatory enantiomer (i.e. D-Phe is D-Phenylalanine)

15 The generally preferred solid phase chemistry protocols for the synthesis of peptidic compounds of this invention using both BOC- and FMOC- alpha-amine protecting group protocols are shown below. In the absence of a detailed experimental procedure to the contrary, the following chemistry was employed for the synthesis of compounds in the examples:

20

Standard BOC Chemistry Cycle:

- 1) 3 X 1 min. DCM
 - 2) check ninhydrin, recouple if positive
 - 3) 1 X 1 min. 45% TFA*
 - 25 4) 1 X 25 min. 45% TFA
 - 5) 1 X 30 sec DCM
 - 6) 1 X 1 min. MeOH
 - 7) 2 X 1 min. DCM
 - 8) 1 X 1 min. 10% TEA/DCM
 - 30 9) 1 X 8 min. 10% TEA/DCM
 - 10) 3 X 1 min. DCM
 - 11) add preactivated amino acid and couple for 1 h
 - 12) go to step 1
- * 45% TFA = 45% TFA, 45% DCM, 5% anisole, 5% ethane dithiol (by volume)

35 Notes:

- a) For peptide amides, use MBHA resin and start the synthesis at step 7.
- b) For the coupling of standard amino acids, premix 3 equiv. of amino acid, BOP, and HOBt in DMA for 10 min., add 3 equiv. of NMM, then add mixture to the peptide resin. Gentle

bubbling of nitrogen into the slurry by means of a glass frit is a preferred method of agitation during the reaction and washing steps.

c) For coupling to N- α -alkyl-amino acids, use 3 equiv. of amino acid and DIPC, overnight. Alternatively, 3 equiv. of amino acid, BOP-Cl, and DIPEA in DCM, overnight, may be used.

- 5 d) To fully deblock and cleave the peptide from the resin, add (per 1 g resin) 10 mL of HF, 1 mL of anisole, and 0.5 mL of ethylmethanethiol and stir at 0° C for 1 h (for peptides containing Trp, 0.25 g of p-cresol must be added). After the HF is removed, the residue is triturated with ether, collected on a glass frit, and washed several times with ether. The crude peptide is extracted off the resin by washing successively with 10% HOAc/water, HOAc, acetonitrile, 10%
10 HOAc/water, and water. The combined filtrates are frozen and lyophilized. Purification, preferably via reverse phase (C-18) HPLC using an acetonitrile (0.1% TFA)/water (0.1% TFA) gradient, provides the pure peptide.

Standard Fmoc Chemistry Cycle:

- 15 1) 5 X 1 min. DMA
2) check ninhydrin, recouple if positive
3) 1 X 1 min. 20% piperidine/DCM*
4) 1 X 15 min. 20% piperidine/DCM
5) 5 X 1 min. DMA
20 6) 1 X 1 min. DCM
7) add preactivated amino acid and couple for 30 min. to 1 h
8) go to step 1

* For the synthesis of peptide acids, the following protocol may be used after deprotection of the second amino acid to prevent the formation of diketopiperazine. Continue
25 from step 3:

- 4a) 1 X 30 sec DMA
5a) 1 X 30 sec DCM
6a) 1 X 30 sec DMA
7a) 1 X 30 sec DCM
30 8a) Add preactivated amino acid and couple for 30 min. - 1 h
9a) go to step 1 above

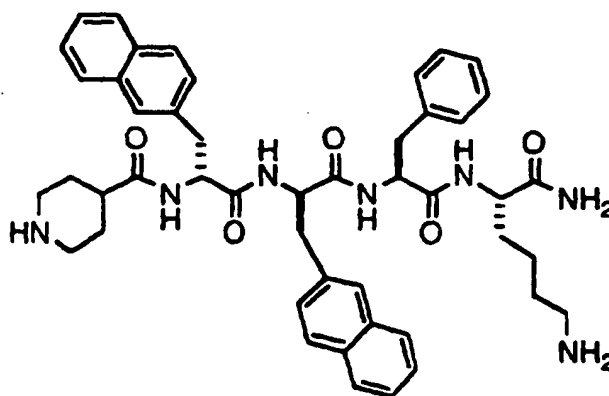
Notes:

- a) For peptide amides, use Fmoc-Am-resin (see below for synthesis), pre-swelled with DCM, and start the synthesis at step 3.
35 b) For coupling of standard amino acids, premix 3 equiv. of the amino acid and BOP in DMA/DCM (1:1) for 10 min., add 3 equiv. of NMM, and add the mixture to the peptide-resin slurry in DMA/DCM (1:1), under gentle nitrogen bubbling.

- c) For coupling to N- α -alkyl-amino acids, use 3 equiv. of amino acid and DIPC in DCM, overnight. Alternatively, 3 equiv. of amino acid, BOP-Cl, and DIPEA in DCM, overnight, may be used.
- d) To fully deblock and cleave the peptide from the resin, add (per 1 g of resin) 10 -15 mL of
 5 95% TFA/triethylsilane (v/v) and shake or stir at room temperature for 1 h. The TFA is removed under vacuum and the residue is triturated with ether, collected on a glass frit, and washed several times with ether. The crude peptide is extracted off the resin by washing successively with 10% HOAc/water, HOAc, acetonitrile, 10% HOAc/water, and water. The combined filtrates are frozen and lyophilized. Purification, preferably via reverse phase (C-18)
 10 HPLC using an acetonitrile (0.1% TFA)/water (0.1% TFA) gradient, provides the pure peptide.
- e) Fmoc-Am-resin was prepared as follows: 60.5 g (0.47mmol/g, 28.4 mmol, Advanced Chemtech #SA5002) of aminomethylated polystyrene resin was placed in a sintered glass funnel reaction vessel and swelled with DCM for 20 min while being agitated with nitrogen bubbling. The solvent was removed by applying a vacuum to the bottom of the funnel and
 15 10% TEA in DCM added. After 20 min., the resin was washed with three portions of DCM and a solution of 23.0 g of p-[(R, S)- α -[1-(9H-Fluoren-9-yl)methoxyformamido]-2,4-dimethoxybenzyl]-PA (42.6 mmol, Novachem) in 100 mL of DMA added. 100 mL of a 1 M solution of DIPC in DCM (100 mmol) was added and the thick suspension agitated for 3.5 h. The resin was washed five times with DMA, twice with DCM, and twice with methanol.
- 20 Drying in vacuo gave 70.2 g of Fmoc-Am-resin at approximately 0.40 mmol/g substitution.

...

Example 1



(inip)-D β Nal-D β Nal-Phe-Lys-amide,
TFA salt

Method A**Step A: (N- ϵ -BOC)Lys-(Am-resin)**

FMOC-Am-resin (10 g, 0.50 mmol/g, 5.0 mmol) was deblocked by agitating with 20% piperidine in DMA for 15 min followed by successive washes with DMA (5X) and DCM (1X).

- 5 The resin gave a positive test with ninhydrin. A solution of 9.37 g (20.0 mmol) of FMOC-(N- ϵ -BOC)-L-Lysine, 8.85 g (20.0 mmol) of BOP, and 3.31 mL (30.0 mmol) of NMM in 30 mL of DMA was added and the solution agitated for 1 h. The resin was washed (5X) with DMA and shown to give a negative ninhydrin test. The N- α -FMOC protecting group was removed with 20% piperidine in DMA for 15 min, followed by successive washings with DMA (5X) and DCM
- 10 (1X) to give (N- ϵ -BOC)Lys-(Am-resin).

Step B: Phe-(N- ϵ -BOC)Lys-(Am-resin)

A solution of FMOC-L-Phenylalanine (7.75 g, 20.0 mmol) and BOP (8.85 g, 20.0 mmol) in 50 mL of DMA/DCM (1:1) was preactivated for 10 min and added to the (N- ϵ -BOC)Lys-(Am-resin)

15 from Step A, followed by NMM (3.31 mL, 30.0 mmol). After 1 h, the resin was washed with DMA (5X, ninhydrin negative), deblocked with 20% piperidine in DMA for 15 min, and washed again with DMA (5X) and DCM (1X) to give Phe-(N- ϵ -BOC)Lys-(Am-resin), displaying a positive ninhydrin test.

20 **Step C: D β Nal-Phe-(N- ϵ -BOC)Lys-(Am-resin)**

FMOC-D- β -naphthylalanine (4.37 g, 10.0 mmol) and 4.42 g (10.0 mmol) of BOP in 50 mL of DMA/DCM (1:1) was preactivated for 10 min, 1.65 mL (15.0 mmol) of NMM was added, and the mixture added to the Phe-(N- ϵ -BOC)Lys-(Am-resin) from step B. After agitation for 2 h, the resin was washed with DMA (5X, ninhydrin negative), deblocked with 20% piperidine in DMA

25 for 15 min, and washed again with DMA (5X) and DCM (1X), to give D β Nal-Phe-(N- ϵ -BOC)Lys-(Am-resin), displaying a positive ninhydrin test.

Step D: FMOC-D β Nal-D β Nal-Phe-(N- ϵ -BOC)Lys-(Am-resin)

FMOC-D β Nal (4.37 g, 10.0 mmol) and 4.42 g (10.0 mmol) of BOP in 50 mL of DMA/DCM (1:1)

30 was preactivated for 10 min and added to the D β Nal-Phe-(N- ϵ -BOC)Lys-(Am-resin) from step C, followed by 1.65 mL (15.0 mmol) of NMM. After agitation for 3 h, the resin was washed with DMA (5X, ninhydrin negative), DCM (2X), and methanol (2X). The resin was dried in vacuo to give 15.6 g of FMOC-D β Nal-D β Nal-Phe-(N- ϵ -BOC)Lys-(Am-resin) with a substitution level of approximately 0.32 mmol/g.

35

Step E: N-FMOC-Isonipecotic acid

To a solution of 10.0 g (77.4 mmol) of isonipecotic acid (Aldrich) in 1 N sodium carbonate/dioxane (1:1) at 0 °C, was added 21.1 g (77.4 mmol) of 9-

fluorenylmethylsuccinimidyl carbonate, portionwise. After 14 h, the dioxane was removed in vacuo, and the suspension diluted with 1200 mL of water. After extraction with 2 portions of ether (discarded), the aqueous solution was cooled in an ice bath and acidified to pH 3 with concentrated hydrochloric acid. The slurry was extracted twice with ethyl acetate and the combined organics washed with water, brine, dried over anhydrous magnesium sulfate, and filtered. The filtrate was concentrated to 500 mL, diluted with 700 mL of hexane, and placed in a refrigerator overnight. The product was collected on a filter and dried in vacuo to give 25.8 g (95%) of N-FMOC-isonipecotic acid as a colorless solid.

10 **Step F: (inip)-D β Nal-D β Nal-Phe-Lys-amide, TFA salt**

FMOC-D β Nal-D β Nal-Phe-(N- ϵ -BOC)-Lys-(Am-resin) (1.0 g, 0.32 mmol) was swelled with DCM for 15 min, deblocked with 20% piperidine in DMA for 15 min, and washed with DMA (5X) and DCM (1X), to give D β Nal-D β Nal-Phe-(N- ϵ -BOC)-Lys-(Am-resin), displaying a positive ninhydrin test. A preactivated solution of N-FMOC-isonipecotic acid (462 mg, 1.32 mmol), 583 mg (1.32 mmol) of BOP, and 0.217 mL (1.98 mmol) of NMM in 10 mL of DMA/DCM (1:1) was added. After agitation for 2 h, the resin was washed with DMA (5X, ninhydrin negative) and deblocked with 20% piperidine in DMA for 15 min. The resin was washed again with DMA (5X) and DCM (3X) then dried in vacuo. The dry resin was suspended in 10 mL of TFA and 0.50 mL of triethylsilane added. The mixture was agitated for 1 h, concentrated in vacuo, and the resin washed 3X with ether. The crude peptide was recovered from the resin by washing with 10% aqueous HOAc, followed by acetonitrile. The combined filtrates were lyophilized to give 140 mg of a solid. A 70 mg aliquot was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt= 30 min) to give 34 mg of (inip)-D β Nal-D β Nal-Phe-Lys-amide, TFA salt as a colorless powder after lyophilization. MS (electrospray, M+H) 798.4.

Method B:

Step A: BOC-(2-Cl-CBZ)Lys-(MBHA-resin)

30 A 20.0 g sample of MBHA-resin (substitution @ 1.04 mmol/g, 20.8 mmol) was washed successively with NMP, DCM (2X), 5% DIPEA/DCM (1X 1 min), 5% DIPEA/DCM (1X 10 min), and DCM (4X). A solution of 25.8 g (3 eq) of BOC-L-(N- ϵ -(2-Cl-CBZ))-Lysine in NMP/DCM (1:1) was added, followed by 8.40 g (3 eq) of HOBT in NMP and 62.4 mL (3 eq) of a 1.0 M solution of DIPC in DCM. After agitation for 1 h, the resin was washed with NMP (1X), DCM (2X), and the coupling judged complete by a ninhydrin test. The resin was neutralized with 5% DIPEA/DCM (1X 1 min), 5% DIPEA/DCM (1X 10 min), DCM (4X), and capped by the addition of 19.7 mL (10 eq) of acetic anhydride and 14.4 mL (4 eq) of DIPEA in DCM for 10 min. Washing the resin with DCM (3X) gave BOC-(2-Cl-CBZ)Lys-(MBHA-resin).

Step B: BOC-Phe-(2-Cl-CBZ)Lys-(MBHA-resin)

Note: The following synthesis cycle was used for all subsequent couplings to this sample of resin:

- 5 1) Deprotect with 50% TFA/DCM for 1 min
- 2) Deprotect with 50% TFA/DCM for 20 min
- 3) Wash resin 4X with DCM
- 4) Neutralize with 5% DIPEA/DCM for 1 min
- 5) Neutralize with 5% DIPEA/DCM for 5-10 min
- 10 6) Wash resin 3X with DCM
- 7) Wash resin 1X with NMP
- 8) Preactivate the BOC-amino acid (3 eq) with BOP (3 eq) and HOBT (3 eq) in NMP for 10 min, add NMM (4.5 eq) and transfer to vessel with resin. Couple for 1 h.
- 9) Wash resin 1X with NMP
- 15 10) Wash resin 2X with DCM
- 11) check ninhydrin for completion of coupling
- 12) Recouple, if necessary (steps 4-11)
- 13) If coupling is complete, proceed with steps 1-11 for coupling of the next residue.
- 20 Specifically, BOC-(2-Cl-CBZ)Lys-(MBHA-resin), vide supra, was deblocked, washed, and coupled with 16.4 g (3 eq) of BOC-L-Phe, 27.6 g of BOP, 8.4 g of HOBT, and 10.3 mL of NMM for 40 min to give BOC-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) (ninhydrin negative).

Step C: BOC-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin)

- 25 The above sample of BOC-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) was deblocked, washed, and coupled with 13.1 g (2 eq) of BOC-D- β -naphthylalanine, 18.3 g of BOP, 5.6 g of HOBT, and 6.85 mL of NMM for 1 h giving an incomplete reaction (ninhydrin positive). The resin was recoupled (c.f. step 12 above) using 6.55 g (1 eq) of BOC-D- β -naphthylalanine, 9.2 g of BOP, 2.8 g of HOBT, and 3.43 mL of NMM for 1 h to give BOC-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin)
- 30 (ninhydrin negative).

Step D: BOC-D β Nal-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin)

- The above sample of BOC-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) was deblocked, washed, and coupled with 13.1 g (2 eq) of BOC-D- β -naphthylalanine, 18.3 g of BOP, 5.6 g of HOBT, and
- 35 6.85 mL of NMM for 3 h giving BOC-D β Nal-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) (ninhydrin negative).

Step E: N-BOC-Isonipecotic acid

To a cold solution of 12.4 g (0.31 mmol, 1.0 eq) of sodium hydroxide in 300 ml of water and 600 ml of dioxane was added 40.0 g (0.31 mmol, 1.0 eq) of isonipecotic acid, followed by 84.0 g (38 mmol, 1.2 eq) of di-*t*-butyl dicarbonate. The mixture was stirred at ambient temperature for 5 h then partitioned between ethyl acetate and 0.5 N citric acid. The organic phase was washed with water, brine, dried over sodium sulfate, and concentrated. The crystalline product was collected by filtration, washed with hexane, and dried under vacuum. Yield: 64.0 g (90%), ¹H NMR (300 MHz, CDCl₃) δ 10.78 (1H, exc), 4.0 (2H, d), 2.85 (2H, t), 2.48 (1H, m), 1.9 (2H, m), 1.65 (2H, m), 1.42 (9H, s). MS (FAB, M+H) 230.1.

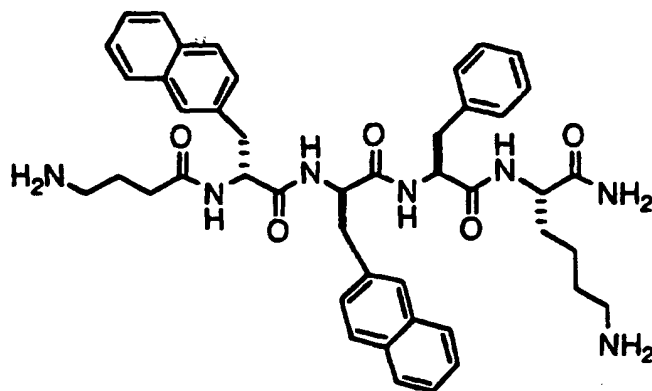
Step F: BOC-(inip)-DβNal-DβNal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin)

The above sample of BOC-DβNal-DβNal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) was deblocked, washed, and coupled with 9.54 g (2 eq) of BOC-isonipecotic acid, 18.3 g of BOP, 5.6 g of HOBT, and 6.85 mL of NMM for 1 h, giving BOC-(inip)-DβNal-DβNal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) (ninhydrin negative).

Step G: (inip)-DβNal-DβNal-Phe-Lys-amide

The above sample of BOC-(inip)-DβNal-DβNal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) was deblocked, washed, neutralized, washed with DCM (4X), ethanol (4X), and dried in vacuo to yield 40.4 g of (inip)-DβNal-DβNal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin). The resin was transferred to a teflon reaction vessel and stirred with a mixture of 200 mL of HF, 20 mL of anisole, and 20 mL of ethyl methyl sulfide at 0 °C for 1 h. The volatiles were removed in vacuo, the resin transferred to a glass fritted funnel and washed repeatedly with ether. The product was extracted from the resin by successive washings with 10% HOAc/water, acetonitrile, and water. The washings were pooled and lyophilized to afford 7.5 g of a powder that was purified by reverse phase HPLC (15-20μ, 300 Å, Vydac C-18, 5x25 cm, gradient: 25 - 45% acetonitrile (0.1% TFA) in water (0.1% TFA) in 200 min at 50 mL/min, rt = 50 min) to give 4.39 g of (inip)-DβNal-DβNal-Phe-Lys-amide, TFA salt as a colorless powder after lyophilization. MS (electrospray, M+H) 798.7.

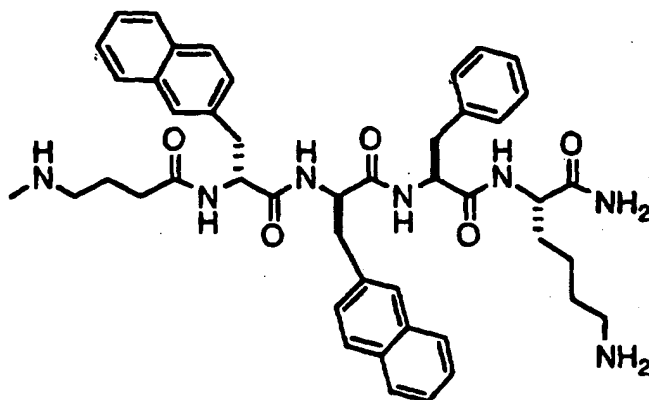
Example 2



(4-aminobutanoyl)-DβNal-DβNal-Phe-Lys-amide,
TFA salt

FMOC-DβNal-DβNal-Phe-(N-ε-BOC)Lys-(Am-resin) (1.0 g, 0.32 mmol), from Example 1, Method A, step D, was swelled with DCM for 15 min, deblocked with 20% piperidine in DMA for 15 min, and washed with DMA (5X) and DCM (1X) to give DβNal-DβNal-Phe-(N-ε-BOC)Lys-(Am-resin), displaying a positive ninhydrin test. A preactivated solution of N-BOC-4-aminobutyric acid (403 mg, 1.98 mmol), 875 mg (1.98 mmol) of BOP, and 0.33 mL (2.97 mmol) of NMM in 15 mL of DMA/DCM (1:1) was added. After agitation for 1 h, the resin was washed with DMA (5X, ninhydrin negative), DCM (3X), MeOH (2X) and dried in vacuo. The dry resin was suspended in 10 mL of TFA and 0.50 mL of triethylsilane added. The mixture was agitated for 1 h, concentrated in vacuo, and the resin washed with ether. The crude peptide was recovered from the resin by washing with 10% aq HOAc, followed by acetonitrile. The combined filtrates were lyophilized to give 245 mg of a solid. A 50 mg aliquot was purified by reverse phase HPLC (15-20μ, 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 25 min) to give 28 mg of (4-aminobutanoyl)-DβNal-DβNal-Phe-Lys-amide, TFA salt as a colorless powder after lyophilization. MS (electrospray, M+H) 772.4.

Example 3



[4-(N-Methylamino)butanoyl]-D β Nal-D β Nal-Phe-Lys-amide,
TFA salt

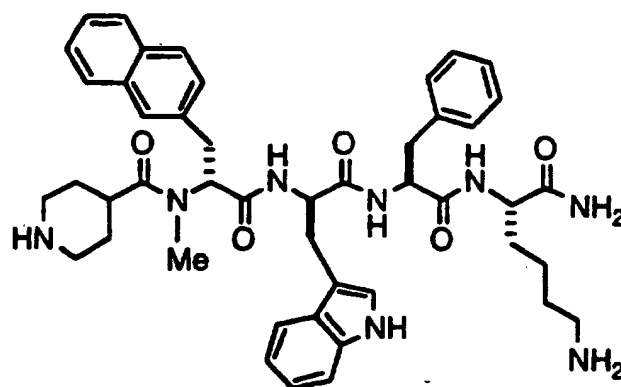
Step A: BOC-4-(N-Methylamino)butyric Acid

To a 0 °C solution of 5.00 g (24.6 mmol) of BOC-4-aminobutyric acid in 75 mL of dry THF, was added 12.2 mL (197 mmol) of methyl iodide followed by 2.95 g (73.8 mmol, 60% dispersion in mineral oil) of sodium hydride, portionwise. The reaction was rapidly stirred at room temperature for 12 h and quenched by the careful addition of water. The mixture was partitioned between ether and water, and the organic phase extracted with 1 N aq sodium bicarbonate. The combined aqueous phases were chilled and acidified to pH 3 with 1 N sodium hydrogen sulfate, then extracted with two portions of ethyl acetate. The combined organics were washed successively with water, 5% aq sodium thiosulfate, water, brine, and then dried over anhydrous magnesium sulfate. Concentration in vacuo afforded 5.00 g (94%) of BOC-4-(N-methylamino)butyric acid. ¹H NMR: (300 MHz, CDCl₃) δ 3.28 (2H, bt, J = 7 Hz), 2.84 (3H, s), 2.36 (2H, t, J = 7.5 Hz), 1.85 (2H, m), 1.45 (9H, s).

Step B: [4-(N-Methylamino)butanoyl]-D β Nal-D β Nal-Phe-Lys-amide, TFA salt

Following the procedure of Example 2, FMOC-D β Nal-D β Nal-Phe-(N- ϵ -BOC)Lys-(Am-resin) (0.50 g, 0.17 mmol, (from Example 1, Method A, Step D) was deblocked and coupled to BOC-4-(N-methylamino)butyric acid (Step A) (147 mg, 0.68 mmol) using 300 mg (0.68 mmol) of BOP and 0.11 mL (1.02 mmol) of NMM. Cleavage from the resin afforded 120 mg of a solid after lyophilization. A 62 mg aliquot was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 23 min) to give 27 mg of pure [4-(N-methylamino)butanoyl]-D β Nal-D β Nal-Phe-Lys-amide, TFA salt. MS (electrospray, M+H) 786.4.

Example 4



(inip)-(N-Me-DβNal)-DTrp-Phe-Lys-amide,
TFA salt

Step A: DTrp-Phe-(N-ε-BOC)Lys-(Am-resin)

A 1 g (approx. 0.4 mmol) sample of Phe-(N-ε-BOC)Lys-(Am-resin) (Example 1, Method A, Step B) was reacted with a preactivated solution of 0.85 g (2.0 mmol) of FMOC-D-Tryptophan, 0.88 g of BOP, and 0.33 mL of NMM for 1 h. Washing and deprotection as per the general protocol above gave DTrp-Phe-(N-ε-BOC)Lys-(Am-resin), displaying a positive ninhydrin test.

Step B: BOC-N-Methyl-D-β-naphthylalanine (BOC-N-Me-DβNal)

To a cold (0° C), stirred, THF solution of N-BOC-D-β-naphthylalanine (15.0 g, 47.6 mmol) and methyl iodide (14.8 mL, 238 mmol) was added sodium hydride (5.70 g of 60% dispersion in mineral oil, 143 mmol) in portions over 45 min. The mixture was allowed to warm slowly to ambient temperature over 16 h then partially concentrated and poured into 1 L of dilute aqueous sodium bicarbonate. Neutral species were extracted into ethyl acetate and discarded. The aqueous phase was acidified with citric acid and the separated product extracted into ethyl acetate, washed with dilute sodium bisulfite, brine, dried over magnesium sulfate, and concentrated. The crude product was crystallized from DCM/hexane to yield 14.9 g (95%) of the title compound. ¹H NMR (300 MHz, CDCl₃, rotational isomers evident) δ 1.31 (9H, 2s, BOC), 2.70 (3H, 2s, N-Me), 3.32 (2H, m, CH₂Ar), 4.8 (1H, m, CH), 7.54 (7H, m, Ar). MS (FAB, M+H) 330.2.

Step C: FMOC-N-Methyl-D-β-naphthylalanine (FMOC-N-Me-DβNal)

BOC-N-methyl-D-β-naphthylalanine (10.0 g, 30.4 mmol) was dissolved in TFA (100 mL) and stirred for 1 h. The TFA was removed under vacuum and the residue combined with 9-fluorenylmethyl-succinimidylcarbonate (13.3 g, 40.0 mmol), potassium carbonate (6.2 g, 45 mmol), THF (300 mL), water (124 mL), and stirred at ambient temperature for 18 h. The reaction mixture was partitioned between ethyl acetate and dilute aq HCl, and the organic phase

washed with brine, dried over magnesium sulfate, and concentrated. The crystalline product was collected by filtration and washed with hexane to yield 12.9 g (94%) of the title compound. ¹H NMR (300 MHz, DMSO-d₆, rotational isomers evident) δ 2.70 (3H, 2s, NMe), 3.20 (2H, m, CH₂Ar), 4.18 (3H, m, OCH₂ and CHAr), 4.90 (1H, m, COCHN), 7.0 to 8.0 (15H, m, Ar), 13.0
 5 (1H, s, COOH). [α]_D²⁰ = +48.0° (c=1.625 in MeOH/DCM 1:1). MS (FAB, M+H) 452.3.

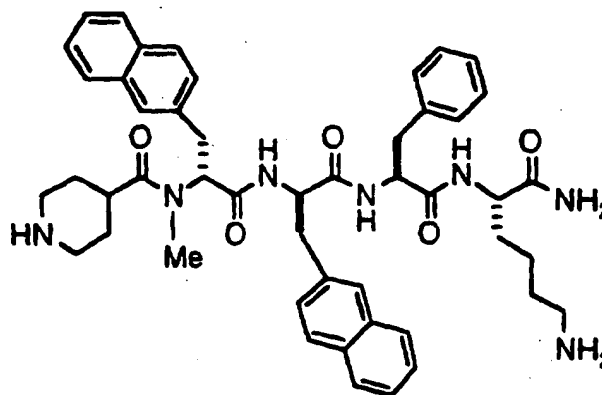
Step D: (N-Me-DβNal)-DTrp-Phe-(N-ε-BOC)Lys-(Am-resin)

The DTrp-Phe-(N-ε-BOC)Lys-(Am-resin) from Step A was agitated with a preactivated solution of 0.68 g (1.5 mmol) of Fmoc-(N-Me-DβNal), 0.88 g of BOP, and 0.33 mL of NMM for 3 h.
 10 Washing and deprotection as per the general Fmoc protocol above gave (N-Me-DβNal)-DTrp-Phe-(N-ε-BOC)Lys-(Am-resin) displaying a faint orange ninhydrin test.

Step E: (inip)-(N-Me-DβNal)-DTrp-Phe-Lys-amide, TFA salt

(N-Me-DβNal)-DTrp-Phe-(N-ε-BOC)Lys-(Am-resin) (1.0 g, 0.32 mmol, Step D) was treated with
 15 a preactivated solution of 0.88 g (2.5 mmol) of N-Fmoc-isonipecotic acid, 1.10 g of BOP, and 0.66 mL of NMM for 4 h. After washing with DMA (5X), a ninhydrin test showed the reaction to be incomplete. The resin was recoupled using 0.88 g (2.5 mmol) of N-Fmoc-isonipecotic acid, 0.56 g of BOP-Cl, and 0.87 mL of DIPEA for 12 h (ninhydrin negative). The resin was washed, deblocked with 20% piperidine in DMA, washed, and dried in vacuo. Cleavage with
 20 TFA and extraction as per the general protocol gave 75 mg of a solid after lyophilization, which was purified by reverse phase HPLC (15-20μ, 300 Å, Vydac C-18, 1X 50 cm, gradient: 17 - 32% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 45 min) to give 23 mg of impure product. Rechromatography (15-20μ, 300 Å, Vydac C-18, 1X 50 cm, gradient: 35 - 55% methanol (0.1% TFA) in water (0.1% TFA) in 80 min at 10 mL/min, rt = 30 min) gave 11 mg
 25 of the pure title compound. MS (electrospray, M+H) 801.6.

Example 5



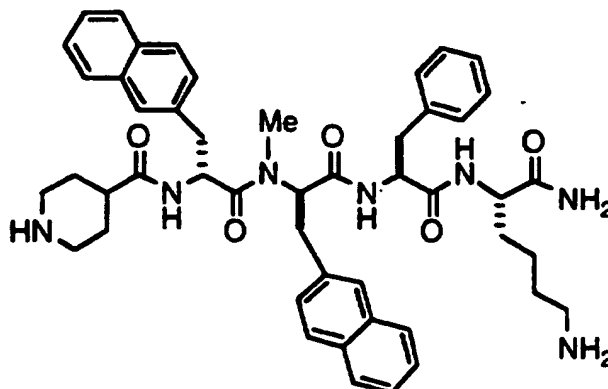
(inip)-(N-Me-DβNal)-DβNal-Phe-Lys-amide,
 TFA salt

Step A: (N-Me-D β Nal)-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin)

A 0.8 g (approx. 0.3 mmol) sample of BOC-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) (Example 1, Method B, Step C) was deblocked, washed, and reacted with a preactivated DMA solution of 0.33 g (1.0 mmol) of BOC-(N-Me-D β Nal) (Example 4, step B), 0.44 g of BOP, 0.14 g of HOBT, and 0.11 mL of NMM for 1.5 h. Washing and deprotection as per the general BOC protocol above gave the title compound displaying a light orange, positive ninhydrin test.

Step B: (inip)-(N-Me-D β Nal)-D β Nal-Phe-Lys-amide, TFA salt

The intermediate from step A was treated with 0.23 g (1.0 mmol) of N-BOC-isonipecotic acid (Example 1, Method B, Step E), 0.29 g of BOP-Cl, and 0.20 mL of DIPEA in DCM for 12 h. Washing, drying, and cleavage as per the general BOC protocol above gave 250 mg of a powder. A 102 mg aliquot was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 32 min) to give 23 mg of the title compound. MS (electrospray, M+H) 812.4.

Example 6**(inip)-D β Nal-(N-Me-D β Nal)-Phe-Lys-amide, TFA salt****Step A: (N-Me-D β Nal)-Phe-(2-Cl-CBZ)Lys-(MBHA-resin)**

A 0.80 g (approx. 0.30 mmol) sample of BOC-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) (Example 1, Method B, Step B) was deblocked, washed, and reacted with a preactivated DMA solution of 0.33 g (1.0 mmol) of BOC-(N-Me-D β Nal) (Example 4, step B), 0.44 g of BOP, 0.14 g of HOBT, and 0.11 mL of NMM for 1.5 h. Washing and deprotection as per the general BOC protocol above gave the title compound displaying a light orange positive ninhydrin test.

Step B: D β Nal-(N-Me-D β Nal)-Phe-(2-Cl-CBZ)Lys-(MBHA-resin)

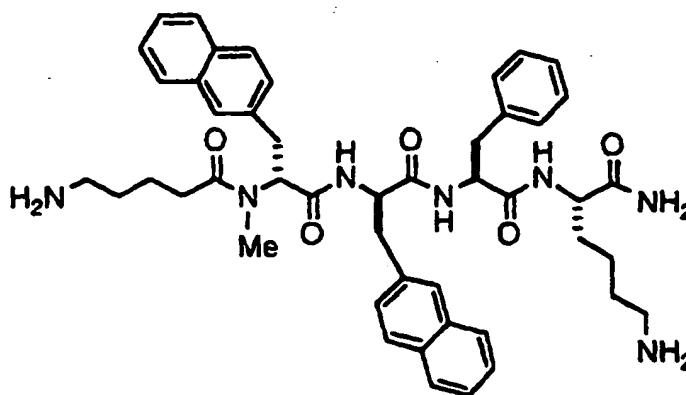
The intermediate from step A was treated with 0.32 g (1.0 mmol) of N-BOC-D- β -naphthylalanine, 0.29 g of BOP-Cl, and 0.20 mL of DIPEA in DCM for 12 h. Washing and deprotection as per the general BOC protocol above gave the title compound displaying a

5 positive ninhydrin test.

Step C: (inip)-D β Nal-(N-Me-D β Nal)-Phe-Lys-amide, TFA salt

The intermediate from step B was reacted with a preactivated DMA solution of 0.23 g (1.0 mmol) of N-BOC-isonipecotic acid, 0.44 g of BOP, 0.14 g of HOBT, and 0.11 mL of NMM for 1 h.

10 Deblocking, washing, drying, and HF cleavage, as per the general BOC protocol above gave 200 mg of a powder. A 67 mg aliquot was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 13 - 28% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 30 min) to give 25 mg of the title compound. MS (electrospray, M+H) 812.4.

Example 7

**(5-Aminovaleryl)-(N-Me-D β Nal)-D β Nal-Phe-Lys-amide,
TFA salt**

15 Step A: BOC-(N-Me-D β Nal)-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin)

A 0.33 mmol sample of BOC-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) (Example 1, Method B, Step C) was treated with TFA deblock, neutralized, washed, and coupled with 3 eq of BOC-(N-Me-D β Nal) (from Example 4, Step B), 3 eq of BOP, 3 eq of HOBT and 4.5 eq of NMM in DMA/DCM for 1 h. The resin was washed with DMA (5X) to give the title compound

20 (ninhydrin negative).

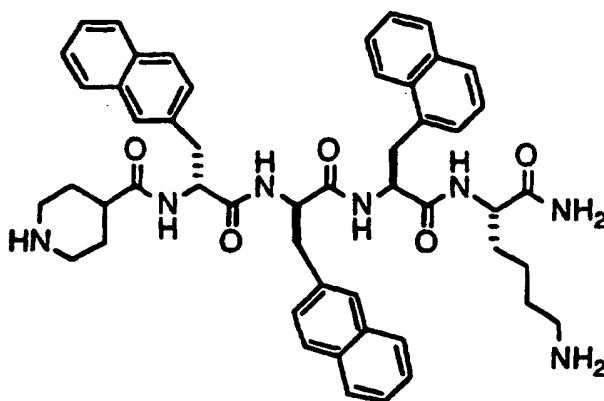
Step B: (5-Aminovaleryl)-(N-Me-D β Nal)-D β Nal-Phe-Lys-amide, TFA salt

The above sample of BOC-(N-Me-D β Nal)-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin) was deblocked with TFA, neutralized, washed, and coupled with 4 eq of N-BOC-5-aminovaleric

acid, 4 eq of BOP-Cl, and 6 eq of DIPEA in DCM overnight. After confirming complete coupling with the ninhydrin test, the peptide was deblocked with TFA, washed, and dried in vacuo to give (5-aminovaleryl)-(N-Me-D β Nal)-D β Nal-Phe-(2-Cl-CBZ)Lys-(MBHA-resin). The peptide was cleaved from the resin and lyophilized, using the methods described in the general procedure, to provide 193 mg of a crude solid. A 53 mg aliquot was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 43 min) to give 7.4 mg of pure (5-aminovaleryl)-(N-Me-D β Nal)-D β Nal-Phe-Lys-amide, TFA salt as a colorless powder after lyophilization. MS (electrospray, M+H) 800.2.

10

Example 8



(inip)-D β Nal-D β Nal- α Nal-Lys-amide,
TFA salt

Step A: BOC- α Nal-(2-Cl-CBZ)Lys-(MBHA-resin)

A 1.0 g sample of BOC-(2-Cl-CBZ)Lys-(MBHA-resin) from Example 1, Method B, Step A, was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 945 mg (3 mmol) of BOC-L- α -naphthylalanine, 1.32 g of BOP, and 0.45 mL of NMM for 1 h, giving BOC- α Nal-(2-Cl-CBZ)Lys-(MBHA-resin) (ninhydrin negative).

15

Step B: BOC-D β Nal- α Nal-(2-Cl-CBZ)Lys-(MBHA-resin)

The intermediate from Step A was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 945 mg (3 mmol) of BOC-D- β -naphthylalanine, 1.32 g of BOP, and 0.45 mL of NMM for 1 h, giving BOC-D β Nal- α Nal-(2-Cl-CBZ)Lys-(MBHA-resin) (ninhydrin negative).

20

Step C: BOC-D β Nal-D β Nal- α Nal-(2-Cl-CBZ)Lys-(MBHA-resin)

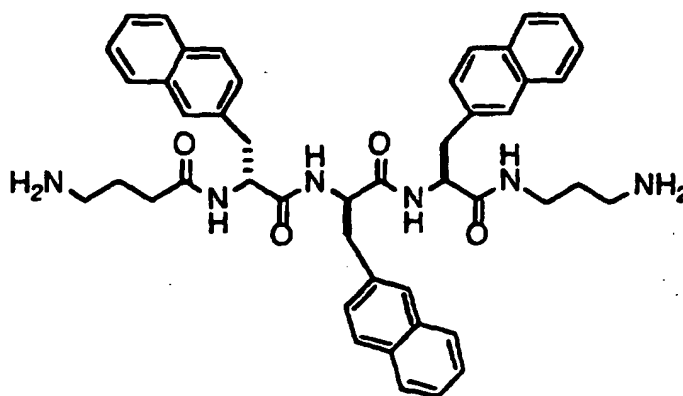
The resin from Step B was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 945 mg (3 mmol) of BOC-D- β -naphthylalanine, 1.32 g of BOP, and 0.45 mL of NMM for 1 h, giving BOC-D β Nal-D β Nal- α Nal-(2-Cl-CBZ)Lys-(MBHA-resin) (ninhydrin negative).

Step D: (inip)-D β Nal-D β Nal- α Nal-(2-Cl-CBZ)Lys-(MBHA-resin)

The resin from Step C was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 690 mg (3 mmol) of N-BOC-isonipecotic acid, 1.32 g of BOP, and 0.45 mL of NMM for 1 h, giving BOC-(inip)-D β Nal-D β Nal- α Nal-(2-Cl-CBZ)Lys-(MBHA-resin) (ninhydrin negative). The resin was deblocked, washed with methanol, and dried in vacuo to give 1.1 g of the title compound.

Step E: (inip)-D β Nal-D β Nal- α Nal-Lys-amide, TFA salt

The above resin (1.1 g) was cleaved with HF according to the general procedure to afford 87 mg of a solid that was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 25%-40% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 32 min) to give 11 mg of pure (inip)-D β Nal-D β Nal- α Nal-Lys-amide, TFA salt. MS (electrospray, M+H) 848.4.

Example 9

**(4-Aminobutanoyl)-D β Nal-D β Nal- β Nal-[N-(3-aminopropyl)] amide,
TFA salt**

Step A: 1,3-Propanediamine-carbonylbenzyloxy-resin (PDA-COO-resin)

Hydroxymethyl resin (10 g, 1 meq/g, 100-200 mesh, Bachem RMIS35) was washed with 5% DIPEA/DCM, toluene (6X), suspended in 60 ml of toluene and gently agitated with nitrogen bubbling. A solution of phosgene in toluene (70 mL, 1.93 M, Fluka) was added and, after 20

min, the resin was washed with toluene (6X), resuspended in 60 mL of dry THF, and 1,3-Propanediamine (5.0 g, Fluka) was added. The mixture was agitated for 1 h, washed with DMA (5X), and shown to give a positive ninhydrin test. A solution of 10% TFA in DCM was added and the resin was washed with DCM (4X), methanol (2X), and dried in vacuo.

5

Step B: BOC- β Nal-(PDA-COO-resin)

A 3.0 g sample of PDA-COO-resin was neutralized with 5% DIPEA/DCM, washed, and coupled with 2.84 g (9.0 mmol) of BOC- β Nal, 3.96 g of BOP, and 1.5 mL of NMM for 1 h (ninhydrin negative). After washing with DCM, the resin was washed with methanol and dried.

10

Step C: BOC-D β Nal- β Nal-(PDA-COO-resin)

A 1.0 g sample of the resin from Step B was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 945 mg (3 mmol) of BOC-D β Nal, 1.32 g of BOP, and 0.45 mL of NMM for 1 h, giving BOC-D β Nal- β Nal-(PDA-COO-resin) (ninhydrin negative).

15

Step D: BOC-D β Nal-D β Nal- β Nal-(PDA-COO-resin)

The above sample was washed, deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 945 mg (3 mmol) of BOC-D β Nal, 1.32 g of BOP, and 0.45 mL of NMM for 1 h giving BOC-D β Nal-D β Nal- β Nal-(PDA-COO-resin) (ninhydrin negative).

20

Step E: (4-Aminobutanoyl)-D β Nal-D β Nal- β Nal-(PDA-COO-resin)

The above sample was washed, deblocked, washed, neutralized with 5% DIPEA/DCM, washed and coupled with 609 mg (3 mmol) of N-BOC-4-aminobutyric acid, 1.32 g of BOP, and 0.45 mL of NMM for 1 h (ninhydrin negative). The resin was washed, deblocked, washed with methanol, and dried in vacuo to give 1.3 g of the title compound.

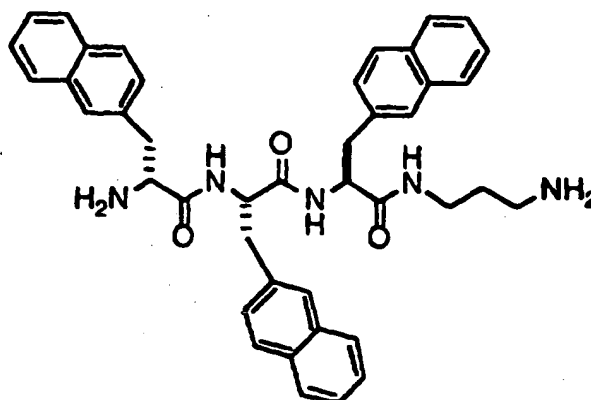
25

Step F: (4-Aminobutanoyl)-D β Nal-D β Nal- β Nal-[N-(3-aminopropyl)] amide, TFA salt

The above resin (1.3 g) was cleaved with HF to afford 225 mg of a solid after lyophilization. A 60 mg sample was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 25 - 40% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 30 min) to give 29 mg of the title compound. MS (electrospray, M+H) 751.4.

30

Example 10



DβNal-βNal-βNal-[N-(3-aminopropyl)] amide,
TFA salt

Step A: BOC-βNal-βNal-(PDA-COO-resin)

- A 2 g sample of BOC-βNal-(PDA-COO-resin) (from Example 9, Step B) was washed, deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 1.89 g (6.0 mmol) of BOC-βNal, 2.64 g of BOP, and 0.90 mL of NMM for 1 h, giving BOC-βNal-βNal-(PDA-COO-resin) (ninhydrin negative). The resin was washed with methanol, and dried in vacuo to give the title compound.

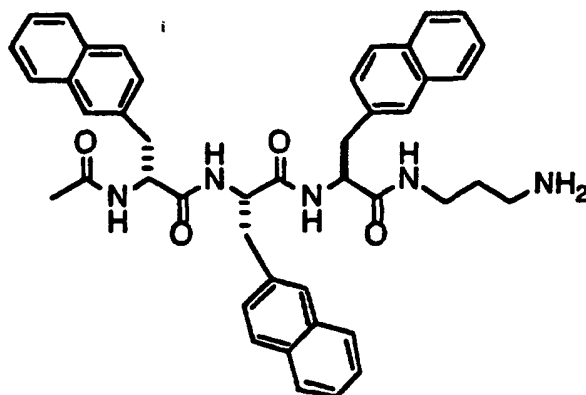
Step B: BOC-DβNal-βNal-βNal-(PDA-COO-resin)

- One gram of the above sample was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 945 mg (3 mmol) of BOC-DβNal, 1.32 g of BOP, and 0.45 mL of NMM for 1 h, giving BOC-DβNal-βNal-βNal-(PDA-COO-resin) (ninhydrin negative). The resin was washed, deblocked, washed with methanol, and dried in vacuo.

Step C: DβNal-βNal-βNal-[N-(3-aminopropyl)] amide, TFA salt

The above resin (1.15 g) was cleaved with HF to afford 99 mg of a solid after lyophilization. The sample was purified as per Example 2 (gradient 27%–42% in 60 min (rt = 22 min)) to give 58 mg of DβNal-βNal-βNal-[N-(3-aminopropyl)] amide, TFA salt. MS (electrospray, M+H) 666.4.

Example 11



Acetyl-DβNal-βNal-βNal-[N-(3-aminopropyl)] amide,
TFA salt

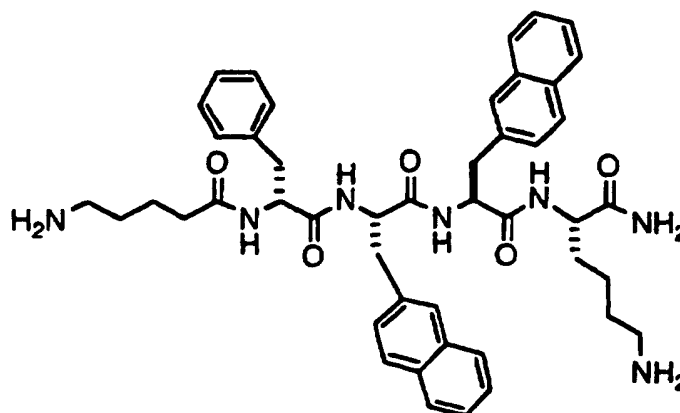
Step A: Acetyl-DβNal-βNal-βNal-(PDA-COO-resin)

A 1.0 g sample of the product of Example 10, Step B was deblocked, neutralized with 5% DIPEA/DCM, and coupled with acetic anhydride (2 mL) in 5% DIPEA/DCM (10 mL) giving Acetyl-DβNal-βNal-βNal-(PDA-COO-resin) (ninhydrin negative). The resin was washed with methanol and dried in vacuo to give 1.23 g of resin.

Step B: Acetyl-DβNal-βNal-βNal-[N-(3-aminopropyl)] amide, TFA salt

The above resin (1.23 g) was cleaved with HF to afford 155 mg of a powder after lyophilization. A 68 mg sample was purified as per Example 2 (gradient 30%-45% in 60 min (rt = 40 min)) to give 31 mg of the title compound. MS (electrospray, M+H) 708.4.

Example 12



(5-Aminovaleryl)-DPhe-βNal-βNal-Lys-amide,
TFA salt

Step A: β Nal-(N- ϵ -BOC)Lys-(Am-resin)

A 0.5 g (approx. 0.25 mmol) sample of (N- ϵ -BOC)Lys-(Am-resin) (Example 1, Method A, Step A) was reacted with a preactivated solution of 0.44 g (1.0 mmol) of Fmoc-L- β Nal, 0.44 g of BOP, and 0.17 mL of NMM for 1 h. Washing and deprotection as per the general protocol
 5 above gave the title compound.

Step B: β Nal- β Nal-(N- ϵ -BOC)Lys-(Am-resin)

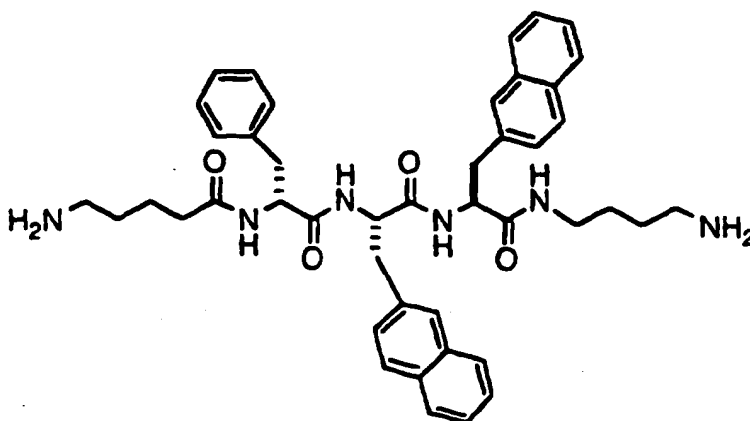
The product of step A was reacted with a preactivated solution of 0.44 g (1.0 mmol) of Fmoc-L- β Nal, 0.44 g of BOP, and 0.17 mL of NMM for 1 h. Washing and deprotection as per the
 10 general protocol above gave the title compound.

Step C: DPhe- β Nal- β Nal-(N- ϵ -BOC)Lys-(Am-resin)

The product of step B was reacted with a preactivated solution of 0.39 g (1.0 mmol) of Fmoc-D-Phenylalanine, 0.44 g of BOP, and 0.17 mL of NMM for 1 h. Washing and deprotection as
 15 per the general protocol above gave the title compound.

Step D: (5-Aminovaleryl)-DPhe- β Nal- β Nal-Lys-amide, TFA salt

The product of step C was reacted with a preactivated solution of 0.22 g (1.0 mmol) of N-BOC-5-aminovaleric acid, 0.44 g of BOP, and 0.17 mL of NMM for 1 h. Washing, drying, and
 20 cleavage as per the general Fmoc protocol above gave 100 mg of a powder. A 51 mg aliquot was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 25 - 40% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, t_r = 25 min) to give 23 mg of the title compound. MS (electrospray, $M+H$) 786.5.

Example 13**(5-Aminovaleryl)-DPhe- β Nal- β Nal-[N-(4-aminobutyl)] amide,****TFA salt**

Step A: 1,4-Butanediamine-carboxybenzyloxy-resin (BDA-COO-resin)

Hydroxymethyl resin (10 g, 1 meq/g, 100-200 mesh, Bachem RMIS35) was washed with 5% DIPEA/DCM, toluene (6X), suspended in 60 ml of toluene and gently agitated with nitrogen bubbling. A solution of phosgene in toluene (70 mL, 1.93 M, Fluka) was added and, after 20 min, the resin was washed with toluene (6X), resuspended in 60 mL of dry THF, and 1,4-Diaminobutane (5.0 g, Fluka) was added. The mixture was agitated for 1 h, washed with DMA (5X), and shown to give a positive ninhydrin test. A solution of 10% TFA in DCM was added and the resin was washed with DCM (4X), methanol (2X), and dried in vacuo.

10 Step B: BOC-βNal-(BDA-COO-resin)

A 1 g sample of BDA-COO-resin from Step A was neutralized with 5% DIPEA/DCM, washed, and coupled with 945 mg (3 mmol) of BOC-βNal, 1.32 g of BOP, and 0.45 mL of NMM for 1 h, giving BOC-βNal-(BDA-COO-resin) (ninhydrin negative).

15 Step C: BOC-βNal-βNal-(BDA-COO-resin)

The above sample was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 945 mg (3 mmol) of BOC-βNal, 1.32 g of BOP, and 0.45 mL of NMM for 1 h giving BOC-βNal-βNal-(BDA-COO-resin) (ninhydrin negative).

20 Step D: BOC-DPhe-βNal-βNal-(BDA-COO-resin)

The above sample was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 795 mg (3 mmol) of BOC-D-Phenylalanine, 1.32 g of BOP, and 0.45 mL of NMM for 1 h, giving BOC-DPhe-βNal-βNal-(BDA-COO-resin) (ninhydrin negative).

25 Step E: BOC-(5-Aminovaleryl)-DPhe-βNal-βNal-(BDA-COO-resin)

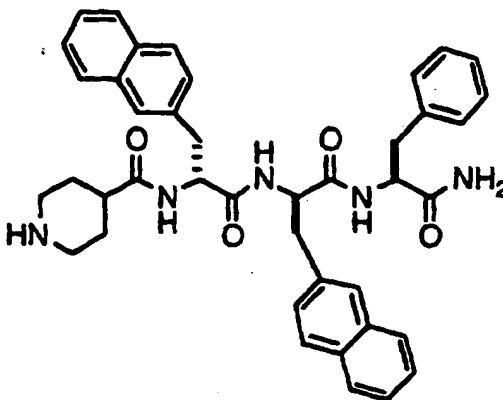
The resin from Step D was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 621 mg (3 mmol) of N-BOC-5-aminovaleric acid, 1.32 g of BOP, and 0.45 mL of NMM for 1 h giving BOC-(5-aminovaleryl)-DPhe-βNal-βNal-(BDA-COO-resin) (ninhydrin negative). The resin was washed, deblocked, washed with methanol, and dried in vacuo.

30

Step F: (5-aminovaleryl)-DPhe-βNal-βNal-[N-(4-aminobutyl)] amide, TFA salt

The above resin (1.1 g) was deblocked with HF to afford 72 mg of solid after lyophilization. The sample was purified as per Example 2 with a 25%-40% gradient ($t_r = 22$ min) to give 16 mg of the title compound. MS (electrospray, $M+H$) 729.5

Example 14



(inip)-DβNal-DβNal-Phe-amide,
TFA salt

Step A: Phe-(MBHA-resin)

An 8.0 g sample of MBHA resin (substitution @ 0.57 mmol/g, 4.56 mmol) was neutralized, washed, and reacted with a preactivated DMA solution of 5.30 g (13.7 mmol) of BOC-L-Phenylalanine, 6.05 g of BOP, 1.85 g of HOBT, and 1.50 mL of NMM for 1.5 h. Washing and deprotection as per the general BOC protocol above gave the title compound.

Step B: DβNal-Phe-(MBHA-resin)

The intermediate from step A was reacted with a preactivated DMA solution of 2.87 g (9.12 mmol) of N-BOC-D-β-naphthylalanine, 4.03 g of BOP, 1.23 g of HOBT, and 1.00 mL of NMM for 1.5 h. Washing and deprotection gave the title compound.

Step C: DβNal-DβNal-Phe-(MBHA-resin)

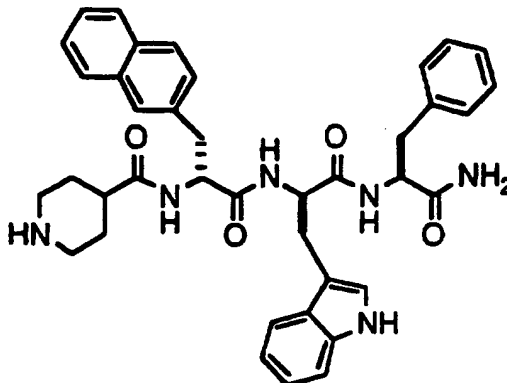
The intermediate from step B was reacted with a preactivated DMA solution of 2.87 g (9.12 mmol) of N-BOC-D-β-naphthylalanine, 4.03 g of BOP, 1.23 g of HOBT, and 1.00 mL of NMM for 1.5 h. Washing and deprotection as per the general BOC protocol above gave the title compound.

Step D: (inip)-DβNal-DβNal-Phe-amide, TFA salt

One half of the intermediate resin from step C (2.28 mmol) was reacted with a preactivated DMA solution of 1.57 g (6.84 mmol) of N-BOC-isonipecotic acid, 3.02 g of BOP, and 1.00 mL of NMM for 3 h. Deblocking, washing, drying, and HF cleavage as per the general BOC protocol above gave 560 mg of a powder which was purified by reverse phase HPLC (15-20μ, 300 Å, Vydac C-18, 1X 50 cm, gradient: 30 - 44% acetonitrile (0.1% TFA) in water (0.1% TFA) in 150

min at 9 mL/min, rt = 60 min) to give 436 mg of the title compound. MS (electrospray, M+H) 670.4.

Example 15



(inip)-DβNal-DTrp-Phe-amide,
TFA salt

Step A: DTrp-Phe-(MBHA-resin)

- 5 A 2.0 g (approx. 1.14 mmol) sample of Phe-(MBHA-resin) (from Example 14, Step A) was reacted with a preactivated DMA solution of 1.04 g (3.42 mmol) of N-BOC-D-Tryptophan, 1.51 g of BOP, 0.46 g of HOBT, and 0.38 mL of NMM for 1.5 h. Washing and deprotection as per the general BOC protocol above gave the title compound.

10 Step B: DβNal-(D-Trp)-Phe-(MBHA-resin)

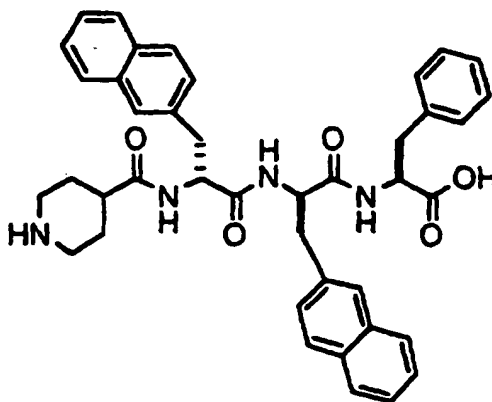
The intermediate from step A was reacted with a preactivated DMA solution of 1.08 g (3.42 mmol) of N-BOC-D-β-naphthylalanine, 1.51 g of BOP, 0.46 g of HOBT, and 0.38 mL of NMM for 1.5 h. Washing and deprotection as per the general BOC protocol above gave the title compound.

15

Step C: (inip)-DβNal-(D-Trp)-Phe-amide, TFA salt

- The intermediate from step B was reacted with a preactivated DMA solution of 0.78 g (3.42 mmol) of N-BOC-isonipecotic acid, 1.51 g of BOP, 0.46 g of HOBT, and 0.38 mL of NMM for 2 h. Deblocking, washing, drying, and HF cleavage as per the general BOC protocol above gave 240 mg of a powder. A 50 mg aliquot was purified by reverse phase HPLC (15-20μ, 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 30 min) to give 23 mg of the title compound. MS (electrospray, M+H) 659.2.
- 20

Example 16



(inip)-DβNal-DβNal-Phe,

TFA salt

Step A: BOC-DβNal-Phe-(O-resin)

Commercially available N-BOC-Phe-(O-resin) (8.0 g, 0.55 mmol/g, 4.41 mmol, 1.0 eq) was swelled in DCM, deblocked, washed, and coupled with 2.78 g (8.82 mmol, 2.0 eq) of N-BOC-D-β-naphthylalanine using 7.8 g of BOP, 1.2 g of HOBT, and 1.45 ml of NMM for 3 h, to give N-BOC-DβNal-Phe-(O-resin) (ninhydrin negative) after washing.

Step B: BOC-DβNal-DβNal-Phe-(O-resin)

The above sample of BOC-DβNal-Phe-(O-resin) was deblocked, washed, and coupled with 2.78 g (8.82 mmol, 2.0 eq) of BOC-D-β-naphthylalanine using 7.8 g of BOP, 1.2 g of HOBT, and 1.45 ml of NMM for 3 h to give BOC-DβNal-DβNal-Phe-(O-resin) (ninhydrin negative).

Step C: BOC-(inip)-DβNal-DβNal-Phe-(O-resin)

The above sample of BOC-DβNal-DβNal-Phe-(O-resin) was deblocked, washed, and coupled with 3.0 g (13.2 mmol, 3.0 eq) of N-BOC-isonipecotic acid (from Example 1, Method B, Step E) using 7.8 g of BOP, 1.2 g of HOBT, and 1.45 ml of NMM for 2 h to give BOC-(inip)-DβNal-DβNal-Phe-(O-resin) (ninhydrin negative), which was washed with methanol and dried in vacuo. Yield: 10.1 g

Step D: (inip)-DβNal-DβNal-Phe, TFA salt

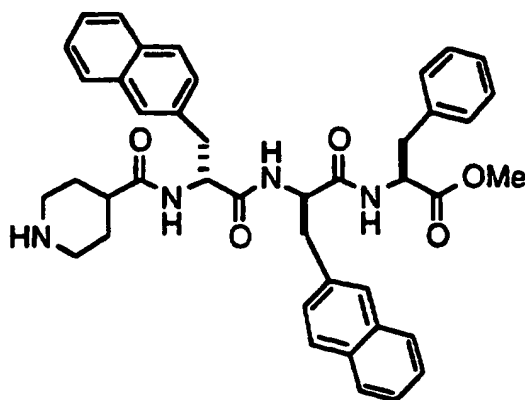
A 3.0 g sample of the above BOC-(inip)-DβNal-DβNal-Phe-(O-resin) was swelled in DCM, deblocked, washed with methanol, dried, and the product cleaved from the resin by the general HF procedure to give 790 mg of (inip)-DβNal-DβNal-Phe of approximately 90% purity. A 37 mg aliquot was purified by HPLC (Vydac C-18, 27 to 42% acetonitrile in water over 60

min, 0.1% TFA, 9 mL/min, 214 nm, rt= 41-51 min) to give 4.3 mg of the title compound. MS (electrospray, M+H) 671.2.

5

Example 17

10



15

(inip)-DβNal-DβNal-Phe methyl ester,
TFA salt

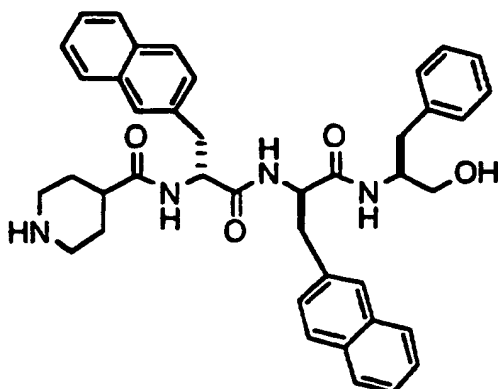
20

To a solution of 50 mg of (inip)-DβNal-DβNal-Phe (from Example 16, Step D) in 20 ml of methanol was added 10 drops of 1 M HCl in diethyl ether. After stirring overnight, the reaction mixture was concentrated and the product purified by HPLC to give 22 mg of the title compound (Vydac C-18, 1 X 50 cm, 9 mL/min, 27 to 42% acetonitrile in water over 60 min, 0.1% TFA, 214 nm, rt= 39-52 min). MS (electrospray, M+H) 685.4.

25

Example 18

30



35

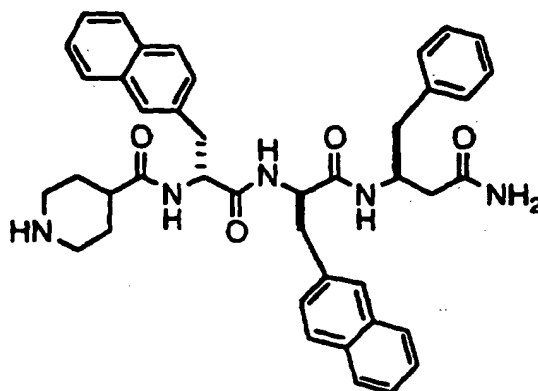
(inip)-DβNal-DβNal-(L-Phenylalanol),
TFA salt

Step A: BOC-(inip)-D β Nal-D β Nal-(L-Phenylalanol)

To 50 ml of dry THF was added 1.5 g (ca. 0.8 mmol, 1.0 eq) of BOC-(inip)-D β Nal-D β Nal-Phe-(O-resin) (from Example 16, Step C) and 4.0 ml (80 mmol, 10 eq) of 2.0 M lithium borohydride solution in THF. The reaction mixture was gently stirred for 1.5 h, 10 ml of HOAc added dropwise, and stirring continued for another 0.5 h. The resin was filtered off, washed with methanol, and the combined filtrates partially evaporated. The product was partitioned between ethyl acetate and water and the organic phase was washed with brine, dried, evaporated, and the product crystallized from ethyl acetate. IR (cm⁻¹): 3409, 3289, 3957, 2930, 1695, 1635, 1536, 1164, 739, 699. MS (FAB, M+H) 757.4.

Step B: (inip)-D β Nal-D β Nal-(L-Phenylalanol), TFA salt

A solution of 50 mg of BOC-(inip)-D β Nal-D β Nal-L-Phenylalanol (step A) in 2 ml of DCM was treated with 2 ml of TFA for 1 h, concentrated, and the product purified by HPLC (Vydac C-18, 1 x 50 cm, 31 to 45% acetonitrile in water over 60 min, 0.1% TFA, 9 mL/min, 214 nm, rt=38 - 45 min). MS (electrospray, M+H) 657.4.

Example 19

(inip)-D β Nal-D β Nal-(3(S)-3-benzyl- β -Ala) amide,
TFA salt

Step A: 3(S)-3-(t-Butoxycarbonylamino)-4-phenylpropionic acid (N-BOC-3(S)-3-benzyl- β -Ala)

This compound was most conveniently prepared by the method of Ondetti and Engel, *J. Med. Chem.* 18(7), (1975), 761-763. Using this procedure, 2.0 g of commercially available N-BOC-L-Phenylalanine was converted to 760 mg of the title compound in 36% overall yield. ¹H NMR (300 MHz, CDCl₃) δ 11.3 (1H, s, exch), 7.2 (5H, m), 5.1 (1H, d), 2.85 (2H, m), 2.5 (2H, m), 1.4 (9H,

s). IR (cm^{-1}) 3316, 2977, 2930, 1709, 1662, 1496, 1370, 1164, 1050, 1025, 746, 699. MS (FAB, M+H) 280.0.

Step B: BOC-(3(S)-3-benzyl- β -Ala)-(MBHA-resin)

- 5 A 2.54 g sample of MBHA-resin (0.64 mmol/g, 2.06 mmol) was swelled in 1:1 DCM /DMA and coupled with 0.60 g (2.15 mmol, 1.1 eq) of N-BOC-(3(S)-3-benzyl- β -Ala) (from Step A) using 1.44 g of BOP, 241 mg of HOBt, and 196 μ l of NMM for 72 h. The resin was washed and capped by acetylation with a mixture of acetic anhydride, TEA, and pyridine in DCM for 15 min, giving the title compound after washing (ninhydrin negative).

10

Step C: BOC-D β Nal-(3(S)-3-benzyl- β -Ala)-(MBHA-resin)

The above sample of BOC-(3(S)-3-benzyl- β -Ala)-(MBHA-resin) was deblocked, washed, and coupled with N-BOC-D- β -naphthylalanine using 2.85 g of BOP, 290 mg of HOBt, and 708 μ l of NMM for 2 h, according to the general procedure, to give BOC-D β Nal-(3(S)-3-benzyl- β -Ala)-

15 (MBHA-resin) (ninhydrin negative).

Step D: BOC-D β Nal-D β Nal-(3(S)-3-benzyl- β -Ala)-(MBHA-resin)

The above sample of BOC-D β Nal-(3(S)-3-benzyl- β -Ala)-(MBHA-resin) was deblocked, washed, and coupled with N-BOC-D- β -naphthylalanine, using 2.85 g of BOP, 290 mg of HOBt, and 708

20 μ l of NMM for 2 h to give the title compound (ninhydrin negative).

Step E: BOC-(inip)-D β Nal-D β Nal-(3(S)-3-benzyl- β -Ala)-(MBHA-resin)

The product of Step D was deblocked, washed, and coupled with N-BOC-isonipecotic acid, using 2.85 g of BOP, 581 mg of HOBt, and 708 μ l of NMM for 18 h to give BOC-(inip)-D β Nal-

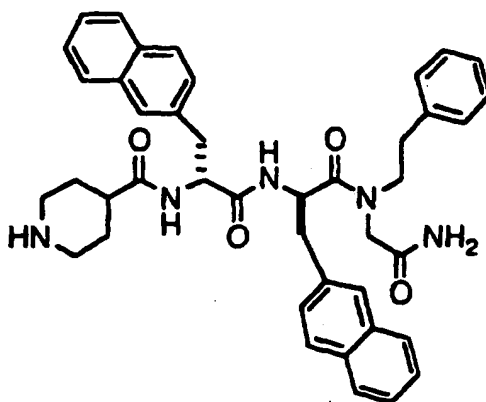
25 D β Nal-(3(S)-3-benzyl- β -Ala)-(MBHA-resin) (ninhydrin negative).

Step F: (inip)-D β Nal-D β Nal-(3(S)-3-benzyl- β -Ala)-amide, TFA salt

The above sample of BOC-(inip)-D β Nal-D β Nal-(3(S)-3-benzyl- β -Ala)-(MBHA-resin) was deblocked, washed with methanol, and dried. The product was cleaved from the resin with HF

30 according to the general procedure to give 280 mg of a solid. A 56 mg portion was purified by HPLC (Vydac C-18, 1x50 cm, 27 to 42% acetonitrile in water over 60 min, 9 mL/min, 0.1% TFA, t_r = 28 - 38 min) to give 23 mg of the title compound. MS (electrospray, M+H) 684.2.

Example 20

(inip)-D β Nal-D β Nal-(N-2-phenylethyl-Gly) amide, TFA salt

Step A: 2-Bromoacetyl-(MBHA-resin)

A 2.0 g sample of MBHA-resin (0.64 mmol/g, 1.28 mmol) was swelled in DCM and coupled with 0.36 g (2.56 mmol, 2.0 eq) of bromoacetic acid using 3.84 ml (3.84 mmol, 3.0 eq) of 1M DIPC in DCM for 2 h. The resin was washed with DCM (5x) to give 2-bromoacetyl-(MBHA-resin) (ninhydrin negative).

Step B: BOC-D β Nal-(N-2-phenylethyl-Gly)-(MBHA-resin)

The above sample of 2-Bromoacetyl-(MBHA-resin) was taken up in DCM and 4.0 ml of phenethylamine added. After 5 h, the resin was washed (ninhydrin positive) and coupled with 806 mg (2.56 mmol, 2.0 eq) of N-BOC-D- β -naphthylalanine using 1.70 g of BOP, 346 mg of HOBT, and 421 μ l of NMM for 18 h, to give the title compound. (ninhydrin negative).

Step C: BOC-D β Nal-D β Nal-(N-2-phenylethyl-Gly)-(MBHA-resin)

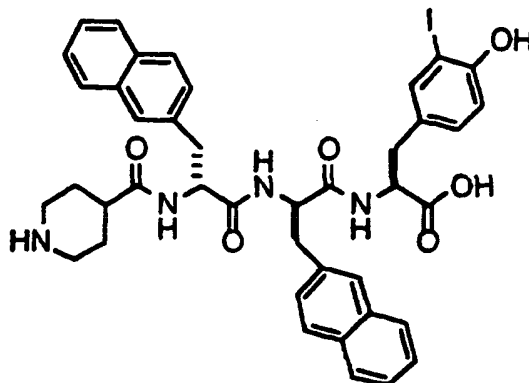
The product of step B was deblocked, washed, and coupled to 806 mg (2.56 mmol, 2.0 eq) of N-BOC-D- β -naphthylalanine using 1.70 g of BOP, 346 mg of HOBT, and 421 μ l of NMM for 4 h to give BOC-D β Nal-D β Nal-(N-2-phenylethyl-Gly)-(MBHA-resin) (ninhydrin negative).

Step D: BOC-(inip)-D β Nal-D β Nal-(N-2-phenylethyl-Gly)-(MBHA-resin)

The above sample of BOC-D β Nal-D β Nal-(N-2-phenylethyl-Gly)-(MBHA-resin) was deblocked, washed, and coupled to 586 mg (2.56 mmol, 2.0 eq) of N-BOC-isonipecotic acid using 1.70 g of BOP, 346 mg of HOBT, and 421 μ l of NMM for 4 h to give BOC-(inip)-D β Nal-D β Nal-(N-2-phenylethyl-Gly)-(MBHA-resin) (ninhydrin negative).

Step E: (inip)-D β Nal-D β Nal-(N-2-phenylethyl-Gly) amide, TFA salt

The resin from Step D was deblocked, washed with methanol, dried, and cleaved with HF according to the general procedure to give 802 mg of a solid. A 70 mg portion was purified by HPLC (Vydac C-18, 1 x 50 cm, 25 to 40% acetonitrile in water over 60 min, 0.1% TFA, 9 mL/min, rt= 33-50 min) to give 42 mg of the title compound. MS (electrospray, M+H) 684.2.

Example 21

(inip)-D β Nal-D β Nal-(3-I-Tyr),
TFA salt

Step A: [3-I-Tyr(3-BrBzl)]-(O-resin)

N-BOC-(O-3-Bromobenzyl)-3-Iodo-L-Tyrosine [Peninsula Labs, BOC-(3-I-Tyr(3-BrBzl))] was coupled to Hydroxymethyl-resin (Bachem, 1%DVB, 100-200 mesh, 1.0 mmol/g) with DIPC (3 eq) and DMAP (0.25 eq) in DMA for 3 h. The resin was washed, deblocked, and washed again, according to the general procedure to give the title compound (ninhydrin positive).

Step B: D β Nal-[3-I-Tyr(3-BrBzl)]-(O-resin)

BOC-D β Nal (SyntheTech, 3 eq) was activated with HBTU (Richelieu Biotechnologies, 4 eq) and DIPEA in DMA and coupled to the resin for 1 h (ninhydrin negative). The resin was washed, deblocked, and washed again, to give the title compound (ninhydrin positive).

Step C: D β Nal-D β Nal-[3-I-Tyr(3-BrBzl)]-(O-resin)

BOC-D β Nal (3 eq) was activated with HBTU (4 eq) and DIPEA in DMA and coupled to the resin for 1 h (ninhydrin negative). The resin was washed, deblocked, and washed again, to give the title compound (ninhydrin positive).

Step D: (inip)-D β Nal-D β Nal-[3-I-Tyr(3-BrBzl)]-(O-resin)

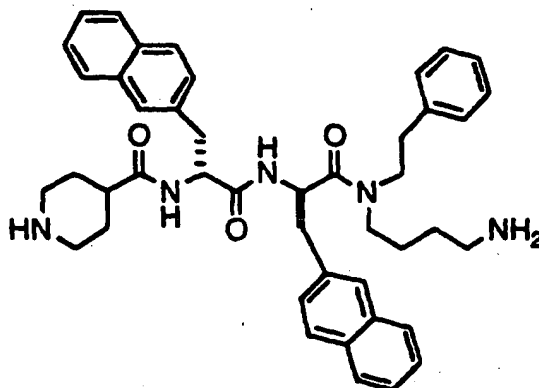
N-BOC-isonipecotic acid (3 eq) was activated with HBTU (4 eq) and DIPEA in DMA and coupled to the above resin for 1 h (ninhydrin negative). The resin was washed, deblocked, washed with DCM, MeOH, and dried in vacuo to give the title compound (ninhydrin positive).

5

Step E: (inip)-D β Nal-D β Nal-(3-I-Tyr), TFA salt

The intermediate from step D was cleaved with anhydrous HF according to the general procedure to give 350 mg of a solid, which was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 2.5 X 27 cm, gradient: 32 - 46% acetonitrile (0.1% TFA) in water (0.1% TFA) in 80 min at 18 mL/min, rt = 20 min) to give 101 mg of (inip)-D β Nal-D β Nal-(3-I-Tyr), TFA salt as a colorless powder after lyophilization. MS (electrospray, M+H) 813.0.

10

Example 22

(inip)-D β Nal-D β Nal-[N-(2-phenylethyl), N-(4-aminobutyl)] amide,
TFA salt

Step A: D β Nal-[4-N-(2-phenylethyl)]-(BDA-COO-resin)

A 1.0 g sample of the BDA-COO-resin from (Example 13, Step A) was swelled with DCM, and a solution of 1% HOAc in DMF added, followed by 69 mg (1.2 eq) of phenylacetaldehyde and 60 mg of sodium cyanoborohydride. After 14 h, the resin was washed repeatedly with DMA and DCM (ninhydrin test showed a red color replacing the deep blue of the starting resin), and the resin neutralized. To a DCM slurry of the resin was added 0.45 g (1.44 mmol) of BOC-D- β -naphthylalanine, 0.43 g of BOP-Cl, and 0.30 mL of DIPEA. After 14 h, the resin was washed, deblocked, and washed again to give the title compound (ninhydrin positive).

15

20

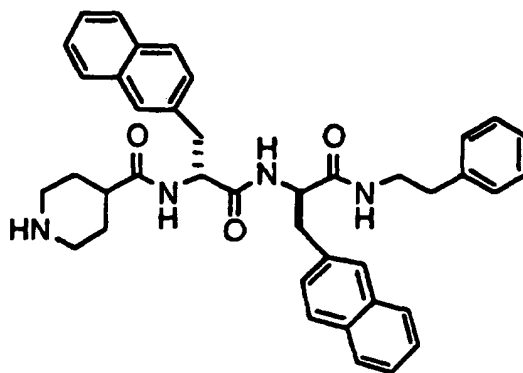
Step B: D β Nal-D β Nal-[4-N-(2-phenylethyl)]-(BDA-COO-resin)

The intermediate from step A was reacted with a preactivated DMA solution of 0.45 g (1.44 mmol) of BOC-D- β -naphthylalanine, 0.64 g of BOP, 0.19 g of HOBT, and 0.21 mL of NMM for 1.5 h. The resin was washed, deblocked, and washed again, giving the title compound (ninhydrin negative).

Step C: (inip)-D β Nal-D β Nal-[N-(2-phenylethyl), N-(4-aminobutyl)] amide, TFA salt

The intermediate from step B was reacted with a preactivated DMA solution of 0.33 g (1.44 mmol) of N-BOC-isonipecotic acid, 0.64 g of BOP, 0.19 g of HOBT, and 0.21 mL of NMM for 2 h.

The resin was washed, deblocked, washed with methanol, and dried in vacuo. HF cleavage as per the general BOC protocol above gave 100 mg of a powder, that was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 35 min) to give 10 mg of the title compound. MS (electrospray, M+H) 698.4.

Example 23

(inip)-D β Nal-D β Nal-(N-2-phenylethyl) amide,
TFA salt

Step A: FMOC-D β Nal-(Wang resin)

Wang resin (6.0 g, 0.63 mmol/g, 3.78 mmol) was swelled in DCM and coupled with 2.48 g (5.67 mmol) of N-FMOC-D- β -naphthylalanine using DIPC (11.3 mL of a 1 M solution in DCM, 11.3 mmol) and 100 mg of DMAP in DCM for 6 h, to give the title compound, after washing with methanol and drying in vacuo (yield 8.14 g).

Step B: D β Nal-D β Nal-(Wang resin)

Employing the standard FMOC chemistry cycle above, a 4.0 g sample of FMOC-D β Nal-(Wang-resin) (0.63 mmol/g, 2.52 mmol) was swelled in DCM, deblocked, washed, and coupled with

2.20 g (5.04 mmol) of N-FMOC-D β Nal using 2.23 g of BOP, and 0.83 ml of NMM for 1.5 h. The sample was then deblocked and washed to give D β Nal-D β Nal-(Wang-resin) (ninhydrin positive).

5 **Step C: BOC-(inip)-D β Nal-D β Nal-(Wang resin)**

The above sample of D β Nal-D β Nal-(Wang-resin) was coupled with 2.31 g (10.1 mmol) of N-BOC-isonipecotic acid (from Example 1, Method B, Step E) using 4.45 g of BOP and 1.38 ml of NMM for 2 h to give BOC-(inip)-D β Nal-D β Nal-(Wang resin) (ninhydrin negative), which was washed with methanol and dried in vacuo.

10

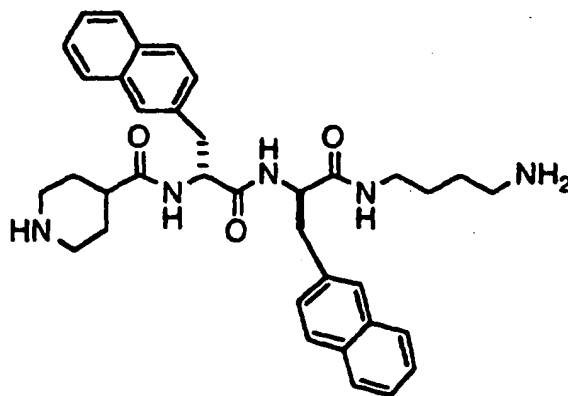
Step D: (inip)-D β Nal-D β Nal-(N-2-phenylethyl) amide, TFA salt

A 0.30 g sample of the above BOC-(inip)-D β Nal-D β Nal-(Wang resin) was suspended in 5 mL of phenethylamine (Aldrich) and 5 mL of DMA. The stirred mixture was placed in a 50 °C oil bath under nitrogen for 18 h, filtered, and the resin washed with DCM. The filtrate was concentrated and partitioned between 1 N sodium hydrogen sulfate and ethyl acetate. The organic phase was washed successively with 1 N sodium bicarbonate, water, brine, and dried over magnesium sulfate. Concentration gave an oil that was treated with 6 mL of DCM/TFA (1:1) for 1 h at ambient temperature and concentrated to give 30 mg of a gum. Purification by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 33 min) gave 7.6 mg of the title compound. MS (electrospray, M+H) 627.6.

15

20

Example 24



(inip)-D β Nal-D β Nal-[N-(4-aminobutyl)] amide,
TFA salt

Step A: BOC-D β Nal-(BDA-COO-resin)

A 5.0 g (0.24 mmol/g, 1.2 mmol) sample of the BDA-COO-resin from Example 13, Step A was neutralized, washed with DCM, and coupled with 0.76 g (2.4 mmol) of BOC-D β Nal, 1.06 g of BOP, 0.32 g of HOBT, and 0.40 mL of NMM for 2 h according to the general BOC chemistry protocols given above. The resin was washed, giving BOC-D β Nal-(BDA-COO-resin) (ninhydrin negative).

Step B: BOC-D β Nal-D β Nal-(BDA-COO-resin)

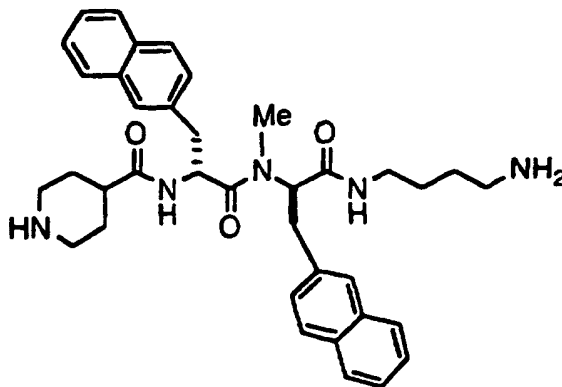
The above sample was deblocked, washed, neutralized, washed, and coupled with 0.76 g (2.4 mmol) of BOC-D β Nal, 1.06 g of BOP, 0.32 g of HOBT, and 0.40 mL of NMM for 2 h, giving BOC-D β Nal-D β Nal-(BDA-COO-resin) (ninhydrin negative).

Step C: (inip)-D β Nal-D β Nal-(BDA-COO-resin)

The resin from Step B was washed, deblocked, washed, neutralized, washed, and coupled with 0.55 g (2.4 mmol) of N-BOC-isonipecotic acid (Example 1, Method B, step E), 1.06 g of BOP, 0.32 g of HOBT, and 0.40 mL of NMM for 2 h, giving BOC-(inip)-D β Nal-D β Nal-(BDA-COO-resin) (ninhydrin negative). The resin was washed with DCM, deblocked, washed with DCM, methanol, and dried in vacuo.

Step D: (inip)-D β Nal-D β Nal-[N-(4-aminobutyl)] amide, TFA salt

The above resin (6 g) was cleaved with HF according to the general procedure and afforded 320 mg of a solid after lyophilization. The sample was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 12%-26% acetonitrile (0.1% TFA) in water (0.1% TFA) in 80 min at 9 mL/min, rt = 18 min) to give 138 mg of the title compound. MS (electrospray, M+H) 594.2.

Example 25**(inip)-D β Nal-(N-Me-D β Nal)-[N-(4-aminobutyl)] amide, TFA salt**

Step A: BOC-(N-Me-D β Nal)-(BDA-COO-resin)

A 1.0 g sample of BDA-COO-resin from Example 13, Step A was washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 981 mg (3 mmol) of BOC-(N-Me-D β Nal), 1.32 g of BOP, and 0.45 mL of NMM for 1 h giving BOC-(N-Me-D β Nal)-(BDA-COO-resin) (ninhydrin negative).

Step B: BOC-D β Nal-(N-Me-D β Nal)-(BDA-COO-resin)

The above sample was washed, deblocked, washed, neutralized with 5% DIPEA/DCM, and washed. BOC-D β Nal (3.29 g, 10 mmol) was activated with DIPC (5.0 mL, 1.0 M in DCM) in 15 mL DCM for 4 min, then added to the resin. After 6 h, the resin was washed, giving BOC-D β Nal-(N-Me-D β Nal)-(BDA-COO-resin) (ninhydrin negative).

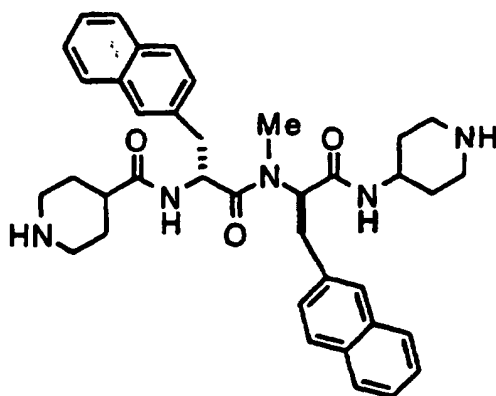
Step C: (inip)-D β Nal-(N-Me-D β Nal)-(BDA-COO-resin)

The above sample was deblocked, washed, neutralized with 5% DIPEA/DCM, washed, and coupled with 648 mg (3 mmol) of N-BOC-isonipecotic acid, 1.32 g of BOP, and 0.45 mL of NMM for 1 h, giving BOC-(inip)-D β Nal-(N-Me-D β Nal)-(BDA-COO-resin) (ninhydrin negative). The resin was washed with DCM, deblocked, washed with DCM, methanol, and dried in vacuo to give 1.2 g of the title compound.

Step D: (inip)-D β Nal-(N-Me-D β Nal)-[N-(4-aminobutyl)] amide, TFA salt

The above resin (1.2 g) was cleaved with HF according to the general procedure to afford 100 mg of a solid after lyophilization. A 57 mg sample was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 20 - 35% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 28 min) to give 27 mg of the pure title compound. MS (electrospray, M+H) 607.7.

Example 26



(inip)-D β Nal-(N-Me-D β Nal)-N-(4-piperidinyI) amide,
TFA salt

Step A: N-CBZ-isonipecotic acid

Benzyl chloroformate (16.4 mL, 115 mmol) in toluene (50 mL) was added dropwise to a stirred solution of 12.9 g (100 mmol) of isonipecotic acid (Aldrich) and 21.0 g (250 mmol) of sodium bicarbonate in 200 mL of water. After 14 h, the mixture was extracted with ether (3 x 50 ml) and the ether layers were discarded. The aqueous layer was acidified with conc. HCl to pH 2, causing the product to precipitate. The product was partitioned into ethyl acetate (3 x 50 mL) and the combined organic layers were washed with brine, dried over magnesium sulfate, and concentrated in vacuo to yield 22.6 g (86%) of N-CBZ-isonipecotic acid as a viscous oil.

Step B: N-CBZ-4-(BOC-amino)-piperidine

A solution of N-CBZ-isonipecotic acid (10.3 g, 38.9 mmol) in tert-butyl alcohol (100 mL) and DCM (100 mL) was treated with diphenylphosphoryl azide (11.8 g, 42.8 mmol), TEA (5.97 mL, 42.8 mmol), and the resulting mixture was heated at reflux for 3 days. The solution was concentrated in vacuo and the residue was partitioned between ether and water. The organic layer was washed successively with 10% aq citric acid, sat. sodium bicarbonate, brine, dried over magnesium sulfate, and concentrated to an oil. This residue was purified by silica gel flash chromatography (gradient elution, 7:3 to 1:1 hexane-ether) to afford 3.2 g (25%) of the title compound as a colorless crystalline solid: TLC R_f 0.21 (1:1 hexane/ethyl ether).

Step C: 4-(BOC-amino)-piperidine

N-CBZ-4-(BOC-amino)-piperidine (3.0 g, 9.0 mmol) was dissolved in ethanol (100 mL) and transferred into a Parr shaker bottle. After adding 10% palladium on carbon (0.5 g), the mixture was shaken under an atmosphere of hydrogen at 50 psi for 0.75 h on a Parr apparatus. The catalyst was removed by filtration through a pad of Celite. The filter cake was washed

with ethanol and the combined filtrate and washings were concentrated in vacuo to yield 1.8 g (100%) of crude 4-(BOC-amino)-piperidine as a pale yellow oil. This product was used immediately in the next step without further purification.

5 Step D: [4-(BOC-amino)-piperidine]-(COO-resin)

Hydroxymethyl resin (4.0 g of 0.45 mmol/g, 1.8 mmol) was rinsed several times with toluene. A solution of 20% phosgene in toluene (50 mL) was added to the hydroxymethyl resin (2 x 30 min) to generate the chloroformate intermediate. After rinsing the resin several times with toluene and dioxane, a solution of 4-(BOC-amino)-piperidine (Step C, 1.8 g, 9.0 mmol) in
10 dioxane was added, and the resulting mixture was agitated for 3 h. The resin was rinsed with dioxane, DCM, and dried in vacuo, to provide 4.4 g of the title compound.

Step E: BOC-(N-Me-D β Nal)-[4-(4-amino-piperidine)]-(COO-resin)

An aliquot (0.82 g, ~0.33 mmol) of the resin from Step D was treated with TFA deblock,
15 neutralized, washed, and coupled with 3 eq of BOC-(N-Me-D β Nal) (from Example 4, Step B), 3 eq of BOP, 3 eq of HOBT and 4.5 eq of NMM in DMA/DCM for 1 h, after which a negative ninhydrin test was observed.

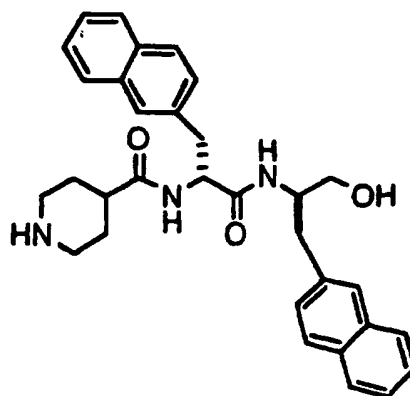
Step F: FMOC-D β Nal-(N-Me-D β Nal)-[4-(4-amino-piperidine)]-(COO-resin)

20 The above sample of BOC-(N-Me-D β Nal)-[4-(4-amino-piperidine)]-(COO-resin) was deblocked with TFA, neutralized, washed, and coupled with 4 eq of FMOC-D- β -naphthylalanine, 4 eq of BOP-Cl, and 6 eq of DIPEA in DCM overnight, after which a negative ninhydrin test was observed.

25 Step G: (inip)-D β Nal-(N-Me-D β Nal)-N-(4-piperidinyI) amide, TFA salt

The above sample of FMOC-D β Nal-(N-Me-D β Nal)-[4-(4-amino-piperidine)]-(COO-resin) was deblocked with 20% piperidine/DMA, washed, and coupled with 3 eq of N-BOC-isonipetric acid (from Example 1, Method B, Step E), 3 eq of BOP, 3 eq of HOBT, and 4.5 eq of NMM in DMA/DCM for 1 h, after which a negative ninhydrin test was observed. The peptide was
30 deblocked with TFA, washed, and dried in vacuo to give (inip)-D β Nal-(N-Me-D β Nal)-[4-(4-amino-piperidine)]-(COO-resin). This peptide was cleaved from the resin with HF and lyophilized as per the general procedure to provide 88 mg of a crude solid. This solid was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 43 min) to give 39 mg
35 of the title compound as a colorless powder after lyophilization. MS (electrospray, M+H) 619.4.

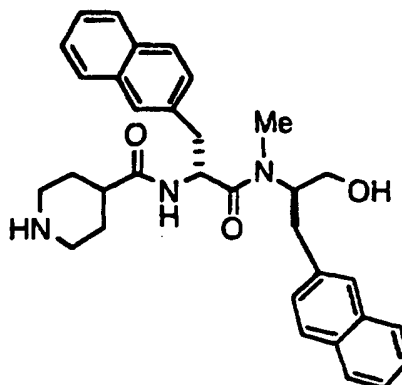
Example 27



(inip)-DβNal-(D-β-naphthylalanol),
TFA salt

A 0.30 g sample of BOC-(inip)-DβNal-DβNal-(Wang resin) (from Example 23, Step C) was suspended in 5 mL of THF under nitrogen and 0.90 mL of a 2.0 M solution of lithium borohydride in THF (Aldrich) added. After 1.5 h, 2 mL of HOAc was added carefully, the suspension filtered, and the resin washed with MeOH. The combined filtrates were concentrated three times from MeOH to give a solid, which was treated with 9 mL of TFA/DCM (2:1) containing a few drops of triethylsilane for 1 h. The solution was concentrated to give 270 mg of a solid, which was purified by reverse phase HPLC (15-20μ, 300 Å, Vydac C-18, 2.5 X 27 cm, gradient: 25 - 39% acetonitrile (0.1% TFA) in water (0.1% TFA) in 80 min at 18 mL/min, rt = 50 min) to give 39 mg of the title compound. MS (electrospray, M+H) 510.0.

Example 28



(inip)-DβNal-(N-Methyl-D-β-naphthylalanol),
TFA salt

Step A: (N-Me-D β Nal)-(O-resin)

A 1.48 g (4.50 mmol) sample of BOC-N-Methyl-D- β -naphthylalanine (from Example 4, Step B) was coupled to 1.5 g (1.0 mmol/g, 1.5 mmol) of hydroxymethyl-resin with DIPC (4.50 mL of a 1.0 M solution in DCM, 4.50 mmol) and 55 mg (0.45 mmol) of DMAP in DMA/DCM (1:1) for 2 h. The resin was washed, deblocked, and washed according to the general BOC procedure, to give the title compound (beads give an orange ninhydrin test).

Step B: D β Nal-(N-Me-D β Nal)-(O-resin)

The above (N-Me-D β Nal)-(O-resin) was coupled with 1.42 g (4.50 mmol) of BOC-D- β -naphthylalanine, 1.01 g of BOP-Cl, and 1.46 mL of DIPEA in DCM for 12 h. The resin was washed and deblocked (ninhydrin positive) to give the title compound.

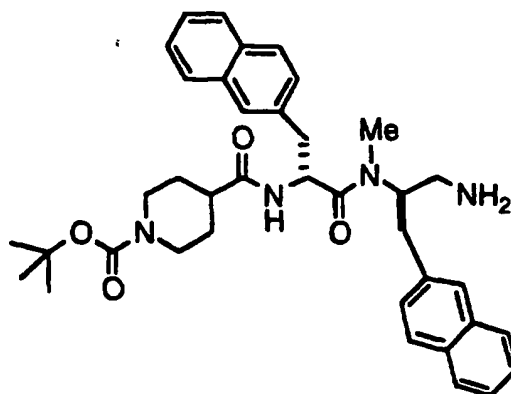
Step C: BOC-(inip)-D β Nal-(N-Me-D β Nal)-(O-resin)

To a slurry of the above D β Nal-(N-Me-D β Nal)-(O-resin) in DCM/DMA (1:1) was added 1.03 g (4.50 mmol) of N-BOC-isonipecotic acid, 1.99 g of BOP, and 0.61 g of HOBT, followed by 1.0 mL of NMM. After 1 h, the resin was washed, washed again with methanol, and dried in vacuo to give 2.33 g of the title compound.

Step D: (inip)-D β Nal-(N-Methyl-D- β -naphthylalanol), TFA salt

A 1.0 g (0.64 mmol) sample of the above BOC-(inip)-D β Nal-(N-Me-D β Nal)-(O-resin) was suspended in 10 mL of THF under nitrogen and 3.2 mL of a 2.0 M solution of lithium borohydride in THF (Aldrich) added. After 1.5 h, 2 mL of HOAc was added carefully, the suspension filtered, and the resin washed with MeOH. The combined filtrates were concentrated 3X from MeOH to give a solid, which was treated with 6 mL of TFA/DCM (1:1), containing a few drops of triethylsilane, for 1 h. The solution was concentrated to give 700 mg of a solid containing salts. A 386 mg aliquot was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1 X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 40 min) to give 21 mg of the title compound. MS (electrospray, M+H) 525.0

Example 29



BOC-(inip)-DβNal-[N-methyl, N-[(2R)-2-(1-amino-3-(2-naphthyl)propyl)] amide, TFA salt

Step A: BOC-(inip)-DβNal

To a stirred solution of 10.0 g (44.0 mmol, 1.0 eq) of N-BOC-isonipecotic acid (from Example 1, Method B, Step E) in 200 ml of DCM was added 100 mg of DMAP and 4.53 g (22 mmol, 0.5 eq) of DCC. After 1 h, the dicyclohexylurea was filtered off and the filtrate added to a solution of 4.7 g (22.0 mmol, 0.5 eq) of D-β-naphthylalanine and 35 ml of NMM in 200 ml of DCM. The reaction was stirred overnight, concentrated, and the residue partitioned between ethyl acetate and 0.5 N citric acid. The organic phase was washed with water, brine, evaporated, and the product recrystallized from ethyl acetate, to give 3.95 g of BOC-(inip)-DβNal. Another 2.14 g was obtained by chromatography of the mother liquors on silica (ethyl acetate/HOAc, 98:2), combined yield: 65%. ¹H NMR (300 MHz, d₆-acetone) δ 7.8 (3H, m), 7.65 (1H, s), 7.4 (3H, m), 7.2 (1H, d), 4.8 (1H, m), 3.88 (2H, m), 3.35 (1H, m), 3.15 (1H, m), 2.62 (2H, m), 2.35 (1H, m), 1.6-1.4 (4H, m), 1.35 (9H, s). MS (FAB, M+H) 427.2.

Step B: BOC-(N-Me-D-β-naphthyl)alanol

To a cold solution of 3.25 g (9.9 mmol, 1.0 eq) of BOC-(N-Me-DβNal) (from Example 4, Step B) and 1.10 g (10.9 mmol, 1.1 eq) of TEA in 50 ml of dry THF, was added dropwise over 30 min, a solution of 1.19 g (10.9 mmol, 1.1 eq) of ethyl chloroformate in 10 ml of dry THF. The reaction turned bright red and TEA hydrochloride precipitated and was filtered off. The red filtrate was added dropwise to a cold, stirred solution of 1.50 g (39.6 mmol, 4.0 eq) of sodium borohydride in 50 ml of methanol/water (1:1). After stirring overnight, the reaction mixture was concentrated to 1/2 the initial volume and partitioned between ethyl acetate and 0.5 N citric acid. The organic phase was washed successively with water, 10% potassium carbonate, saturated sodium bicarbonate, brine, dried over sodium sulfate, filtered, and concentrated to yield 2.91 g of an oil (94%). ¹H NMR (300 MHz, CDCl₃) δ 7.75 (3H, m), 7.6 (1H, s), 7.4 (3H, m),

4.3 (1H), 3.7 (2H, m), 3.0-2.8 (6H, m), 1.3 (9H, 2s). IR (cm^{-1}) 3428, 3050, 2977, 1689, 1669, 1363, 1144. MS (EI, e/m) 427.2.

Step C: (2R)-1-Azido-2-(BOC-methylamino)-3-(2-naphthyl) propane

- 5 A cold solution of hydrazoic acid (4.14 mmol, 1.5 eq) in benzene was prepared by acidification of a stirred mixture of 269 mg (4.14 mmol) of sodium azide, 5 ml of water and 5 ml of benzene with 5 ml of 3.6 N sulfuric acid at 10 °C. The benzene phase was separated, dried with sodium sulfate, and filtered. This solution was added to a stirred mixture of 867 mg (3.31 mmol, 1.2 eq) of triphenylphosphine and 669 mg of diisopropylazodicarboxylate (3.31 mmol, 1.2 eq) in dry THF at -78 °C. A solution of 870 mg (2.76 mmol, 1.0 eq) of BOC-(N-Me-D- β -naphthyl)alanol (step B) in 10 ml of dry THF was added and the reaction mixture allowed to come to room temperature over 2 h. Saturated sodium bicarbonate (20 mL) was added and the reaction mixture partially concentrated to remove the THF. The mixture was partitioned between ethyl acetate and saturated sodium bicarbonate and the organic phase was washed with brine, dried over sodium sulfate, filtered, and evaporated. Flash chromatography on silica (hexane/ethyl acetate, 80:20) gave 600 mg (64%) of the title compound. ^1H NMR (300 MHz, CDCl_3) δ 7.75 (3H, m), 7.6 (1H, d), 7.42 (2H, m), 7.25 (1H, m), 4.4 (1H, m), 3.6-2.8 (4H, m), 2.7 (3H, 2s), 1.3 (9H, 2s). IR (cm^{-1}) 3057, 2977, 2094, 1689.
- 10
- 15

20 **Step D: BOC-(inip)-D β Nal-(N-methyl, N-[(2R)-2-(1-azido-3-(2-naphthyl)propyl)] amide**

- A solution of 300 mg of (2R)-1-azido-2-(BOC-methylamino)-3-(2-naphthyl) propane (0.88 mmol, 1.0 eq) (step C) in 2.0 ml of DCM was treated with 2.0 ml of TFA for 1 h then evaporated several times from DCM. The residue was dissolved in a solution of 5 ml of DCM and 250 μl of NMM then added to a previously prepared mixture of 751 mg (1.76 mmol, 2.0 eq) of N-BOC-(inip)-D β Nal (step A), 363 mg (1.76 mmol, 2.0 eq) of DCC, 238 mg (1.76 mmol, 2.0 eq) of HOBT, and 250 μl of NMM in 15 ml of DCM/DMF (2:1). The reaction was stirred overnight, triturated with 1 mL of water and the precipitated dicyclohexylurea was filtered off. The reaction mixture was taken up in ethyl acetate and washed successively with 0.5 N citric acid, water, 10% potassium carbonate, water, saturated sodium bicarbonate, brine, dried over sodium sulfate, filtered, and evaporated. Flash chromatography on silica (ethyl acetate/hexane (1:1), R_f = 0.4) gave 380 mg (67%) of the title compound. IR (cm^{-1}) 3309, 3057, 2977, 2930, 2100, 1689, 1635, 1171. MS (FAB, $M+H$) 649.4.
- 25
- 30

Step E: BOC-(inip)-D β Nal-(N-methyl, N-[(2R)-2-(1-amino-3-(2-naphthyl)propyl)] amide,

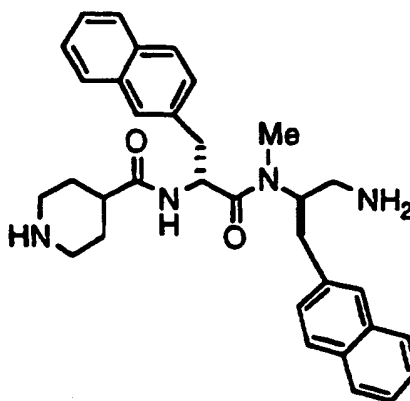
35 **TFA salt**

A solution of 280 mg (0.43 mmol) of the azide from step D in 15 ml of methanol and 1 ml HOAc was hydrogenated over 100 mg of 10% palladium on carbon at 30 psi for 8 hrs. The catalyst was filtered off and the filtrate evaporated to give 250 mg of crude product. A 20 mg aliquot

was purified by reverse phase HPLC (15-20 μ , 300 Å, Vydac C-18, 1 X 50 cm, gradient: 30 - 45% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 39 - 48 min) to give 8 mg of the title compound. IR (cm⁻¹) 3409, 3296, 3057, 2977, 1675, 1430, 1204, 1171, 1131. MS (electrospray, M+H) 624.5

5

Example 30

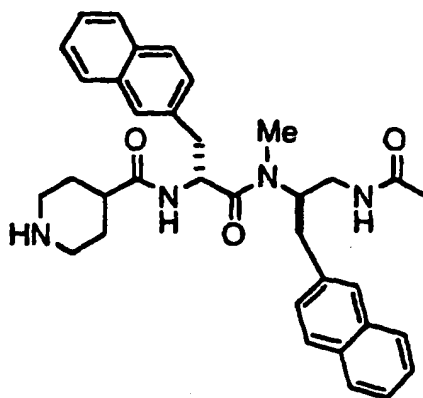


(inip)-D β Nal-[N-methyl, N-[(2R)-2-(1-amino-3-(2-naphthyl)propyl)]] amide,
TFA salt

A solution of 200 mg of BOC-(inip)-D β Nal-[N-methyl, N-[(2R)-2-(1-amino-3-(2-naphthyl)propyl)]] amide, TFA salt (from Example 29, step E) in 2 ml of DCM was treated with 2 ml of TFA, stirred for 1 h, and concentrated to give 490 mg of crude product. A 100 mg aliquot was purified by HPLC (Vydac C-18, 1 x 50 cm, 20 to 35% acetonitrile in water, 60 min, 0.1% TFA, 9 mL/min, rt= 21-31 min) giving 17 mg of the title compound. MS (electrospray, M+H) 523.2.

10

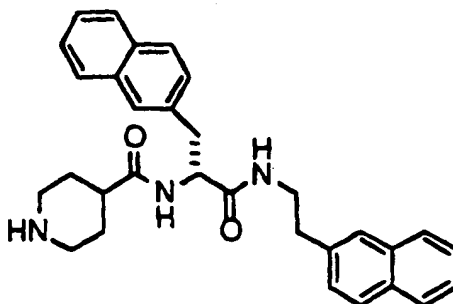
Example 31



(inip)-DβNal-[N-methyl, N-[(2R)-2-(1-acetamido-3-(2-naphthyl)propyl)]] amide,
TFA salt

To a solution of 33 mg (0.05 mmol) of BOC-(inip)-DβNal-[N-methyl, N-[(2R)-2-(1-amino-3-(2-naphthyl)propyl)]] amide, TFA salt (from Example 29, step E) in 0.5 ml of DCM was added 0.5 ml of TEA and 0.5 ml of acetic anhydride. The reaction mixture was stirred for 1 h then
5 evaporated and partitioned between ethyl acetate and water. The organic phase was dried over sodium sulfate and evaporated. The residue was taken up in 2 ml of DCM and 2 ml of TFA added. The mixture was stirred for 1 h, evaporated and the crude product (48 mg) was purified by HPLC (Vydac C-18, 1 x 50 cm, 25 to 50% acetonitrile in water over 60 min, 9 mL/min, 0.1% TFA, rt= 29 min) giving 15 mg of the title compound. MS (electrospray, M+H)
10 565.4.

Example 32



(inip)-DβNal-[N-1-[2-(2-naphthyl)ethyl]] amide,
TFA salt

Step A: 2-(2-Naphthyl)ethylamine hydrochloride

A solution of lithium aluminum hydride (LAH, 100 mmol, 3.0 eq) in 300 ml of dry ether at 0 °C was stirred and a solution of 2-naphthylacetonitrile (5.0 g, 30 mmol, 1.0 eq) in 200 ml of dry

ether added over 2 h. To the resultant bright orange slurry, was added dropwise, 25 ml of cold (0 °C) 12 N sulfuric acid, and the mixture stirred until colorless. The reaction mixture was partitioned between ether and water and the ether phase, containing mostly unreacted 2-naphthylacetonitrile, discarded. The aqueous phase was basified with sodium hydroxide and the separated free amine extracted into ether, dried over sodium sulfate, filtered, and acidified with anhydrous HCl in dioxane. The precipitated HCl salt was collected by filtration to give 890 mg of the title compound. ¹H NMR (300 MHz, D₂O) δ 7.82 (3H, t), 7.7 (1H, s), 7.45 (2H, m), 7.38 (1H, d), 3.25 (2H, t), 3.05 (2H, t). MS (FAB, M+H) 172.1.

10 **Step B: BOC-(inip)-DβNal-[N-1-(2-(2-naphthyl)ethyl)] amide**

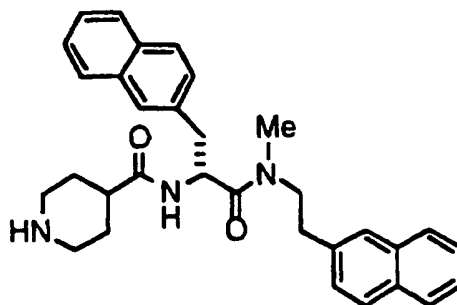
A mixture of 100 mg (0.23 mmol, 1.0 eq) of N-BOC-(inip)-DβNal (from Example 29, step A), 42 mg (0.23 mmol, 1.0 eq) of 2-(2-naphthyl)ethylamine hydrochloride (step A), 132 mg (0.69 mmol, 3.0 eq) of EDC, 31 mg (0.23 mmol, 1.0 eq) of HOBt, 87 μl (0.69 mmol, 3.0 eq) of NMM, and 5.0 ml of DMF was stirred overnight at ambient temperature. The reaction mixture was partitioned between ethyl acetate and dilute hydrochloric acid and the separated organic phase washed successively with water, saturated sodium bicarbonate, brine, dried over sodium sulfate, filtered, and evaporated. The crude product was chromatographed on silica (ethyl acetate/hexane (70:30), R_f= 0.5) to give 110 mg of the title compound. IR (cm⁻¹) 3289, 3057, 2977, 2930, 2857, 1695, 1642, 1423, 1171, 819, 739. MS (FAB, M+H) 580.3.

20

Step C: (inip)-DβNal-[N-1-(2-(2-naphthyl)ethyl)] amide, TFA salt

A solution of 110 mg of N-BOC-(inip)-DβNal-[N-1-(2-(2-naphthyl)ethyl)] amide (step B) in 4 ml of DCM was treated with 2 ml of TFA and stirred for 2 h. The concentrated crude product (121 mg) was purified by HPLC (1 x 50 cm, Vydac C-18, 25 to 40% acetonitrile in water over 60 min, 9 mL/min, 0.1% TFA, rt= 25-30 min) to give 29 mg of the title compound. MS (electrospray, M+H) 479.8.

Example 33



(inip)-DβNal-[N-methyl, N-1-(2-(2-naphthyl)ethyl)] amide, TFA salt

Step A: N-Methyl-N-2-(2-naphthyl)ethylamine, TFA salt

A solution of 2-(2-naphthyl)ethylamine (1.4 mmol, 1.0 eq) in DCM was prepared by partitioning 250 mg of 2-(2-naphthyl)ethylamine hydrochloride (from Example 32, step A) between 5 ml of DCM and 5 ml of 10% aq sodium hydroxide. The organic phase was dried
5 over sodium sulfate, filtered, and 363 mg (1.67 mmol, 1.2 eq) of di-*t*-butyldicarbonate added, followed by 1.0 ml of TEA. After stirring for 30 min, the reaction mixture was concentrated and the product crystallized from hexane at -78 °C. Yield: 250 mg.

This product was dissolved in 10 ml of dry THF, 262 μ l (4.2 mmol, 3.0 eq) of methyl iodide added, and the solution cooled to 0 °C. Sodium hydride (47 mg, 60% dispersion in mineral oil,
10 1.96 mmol) was added in portions with stirring over 10 min, and the reaction allowed to warm to ambient temperature overnight. The solvents were evaporated and the crude product taken up in ethyl acetate/hexane and washed successively with saturated sodium bicarbonate, 1 N sodium bisulfite, water, brine, dried over sodium sulfate, filtered, and evaporated. Treatment
15 with TFA/DCM (1:1) for 1 h and concentration gave 190 mg of the title compound as an oil. ¹H NMR (300 MHz, CDCl₃) δ 7.78 (3H, t), 7.62 (1H, s), 7.4 (2H, m), 7.32 (1H, d), 2.9 (4H, m), 2.4 (3H, s), 1.55 (1H, s). MS (FAB, M+H) 186.0.

Step B: BOC-(inip)-D β Nal-[N-methyl, N-1-[2-(2-naphthyl)ethyl]] amide

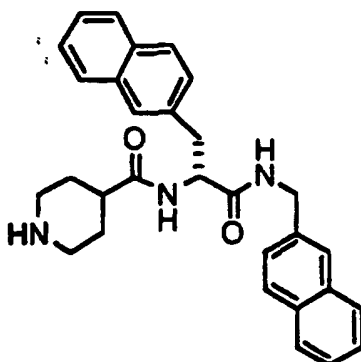
A mixture of 525 mg (1.23 mmol, 1.2 eq) of BOC-(inip)-D β Nal (from Example 29, step A), 190
20 mg (1.02 mmol, 1.0 eq) of N-methyl-N-2-(2-naphthyl)ethylamine (step A), 584 mg (3.06 mmol, 3.0 eq) of EDC, 138 mg (1.02 mmol, 1.0 eq) of HOBT, 412 mg of (4.08 mmol, 4.0 eq) of NMM, and 15 ml of DMF was stirred overnight at ambient temperature. The reaction mixture was partitioned between ethyl acetate and dilute citric acid and the separated organic phase washed
25 successively with water, saturated sodium bicarbonate, brine, dried over sodium sulfate, filtered, and evaporated. The crude product was chromatographed on silica (80:20, ethyl acetate/hexane) to give 550 mg of the title product. IR (cm⁻¹) 3302, 3050, 2977, 2930, 1689, 1629, 1423, 1164, 819, 732.

Step C: (inip)-D β Nal-[N-methyl, N-1-[2-(2-naphthyl)ethyl]] amide, TFA salt

30 A solution of 550 mg of BOC-(inip)-D β Nal-[N-methyl, N-1-[2-(2-naphthyl)ethyl]] amide (step B) in 4 ml of DCM was treated with 3 ml of TFA, stirred for 2 h, and concentrated to give 569 mg of crude product. A 90 mg aliquot was purified by HPLC (1 x 50 cm, Vydac C-18, 27 to 42% acetonitrile in water over 60 min, 0.1% TFA, 9 mL/min, 214 nm, rt=30-45 min) to give 29 mg of the title compound. MS (electrospray, M+H) 493.8.

35

Example 34



(inip)-DβNal-(N-(2-naphthyl)methyl) amide,
TFA salt

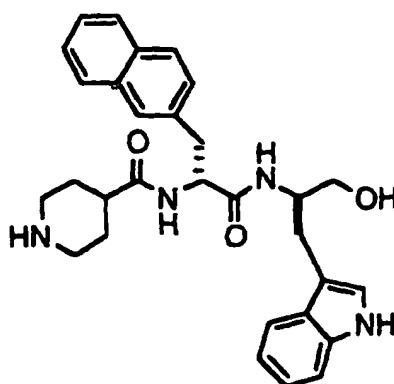
Step A: 2-Aminomethylnaphthylene hydrochloride

A stirred 0 °C solution of 20.0 g of 2-naphthaldehyde (128 mmol) and 98.7 g (1.28 mol) of ammonium acetate in 200 mL of MeOH/HOAc (99:1) was treated with 5.62 g (90.0 mmol) of sodium cyanoborohydride, portionwise. The solution was stirred at ambient temperature for 24 h, concentrated in vacuo, resuspended in water, and basified with sodium hydroxide. The product was extracted into ether, washed with water, brine, dried over magnesium sulfate, and filtered. The filtrate was treated with a dry ethereal solution of HCl, and the precipitated product washed with ether and dried to give 16.5 g (65%) of 2-aminomethylnaphthylene hydrochloride.

Step B: (inip)-DβNal-(2-aminomethylnaphthyl) amide, TFA salt

A mixture of 124 mg (0.29 mmol) of BOC-(inip)-DβNal (from Example 29, step A), 84.5 mg (0.54 mmol) of 2-aminomethylnaphthylene hydrochloride (step A), 67 mg (0.348 mmol) of EDC, 47 mg (0.348 mmol) of HOBt, and 140 μL of NMM in 5 mL of DMF was stirred overnight at ambient temperature. The reaction mixture was partitioned between ethyl acetate and water and the separated organic phase washed successively with 1 N sodium hydrogen sulfate, 1 N sodium bicarbonate, brine, dried over magnesium sulfate, filtered, and evaporated. The crude product was dissolved in 4 mL of DCM/TFA (1:1), stirred for 2 h, and reconcentrated to give 160 mg of crude product. An 85 mg aliquot was purified by HPLC (1 x 50 cm, Vydac C-18, 23 to 38% acetonitrile in water over 60 min, 0.1% TFA, 9 mL/min, t_r = 45 min) to give 4.6 mg of the title compound. MS (electrospray, M+H) 466.0.

Example 35



(inip)-D β Nal-(D-Tryptophanol),
TFA salt

Step A: BOC-D β Nal-DTrp-(O-resin)

A 1.5 g sample of BOC-DTrp-(O-resin) (0.5 mmol/g, 0.75 mmol) was deblocked (TFA/DCM (1:1) containing 1 g/L of indole as a scavenger), washed, neutralized, and coupled with 0.71 g (3 eq) of BOC-D β Nal, 1.02 g (3 eq) of BOP, 0.30 g (3 eq) of HOBT, and 0.37 mL (4.5 eq) of NMM for 1.5 h, according to the general BOC procedure, giving BOC-D β Nal-DTrp-(O-resin) (ninhydrin negative).

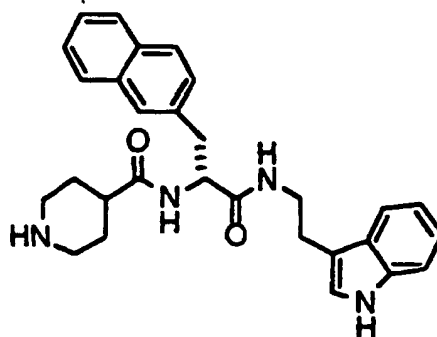
Step B: BOC-(inip)-D β Nal-DTrp-(O-resin)

10 The above sample of BOC-D β Nal-DTrp-(O-resin) was deprotected, washed, and coupled with 0.52 g (3 eq) of N-BOC-isonipecotic acid, 1.02 g (3 eq) of BOP, 0.30 g (3 eq) of HOBT, and 0.37 mL (4.5 eq) of NMM for 1 h, giving 1.76 g of the title compound after washing with methanol and drying in vacuo (ninhydrin negative).

Step C: (inip)-D β Nal-(D-Tryptophanol), TFA salt

15 The dry resin from step B (BOC-(inip)-DβNal-DTrp-(O-resin), 1.76 g) was suspended in 50 mL of THF under nitrogen and 4.80 mL of a 2.0 M solution of lithium borohydride in THF added. After 1.5 h, 4.5 mL of HOAc was added dropwise over 10 min. After 30 min, the suspension was filtered, and the resin washed with MeOH. The combined filtrates were concentrated and partitioned between ethyl acetate and water (a few drops of HOAc added). The organic phase
20 was concentrated, and treated with 20 mL of TFA/DCM (1:1) for 30 min. The TFA was removed in vacuo and the product reconstituted from DCM (3X) to give 700 mg of an oil. A 100 mg aliquot was purified by reverse phase HPLC (15-20μ, 300 Å, Vydac C-18, 1 X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 20 min) to give 23 mg of the title compound. MS (electrospray, M+H) 499.5.

Example 36



(inip)-DβNal-[N-tryptaminy] amide, TFA salt

Step A: BOC-(inip)-DβNal-[N-tryptaminy] amide

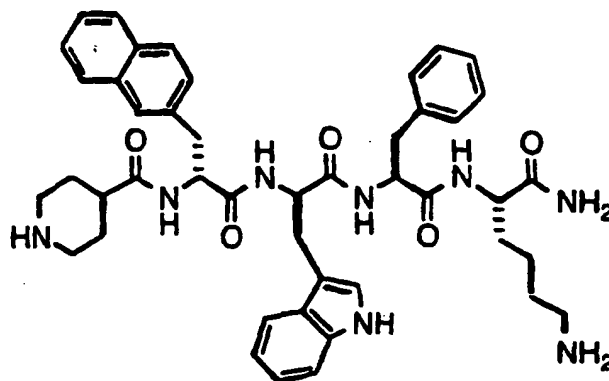
A mixture of 100 mg (0.23 mmol, 1.0 eq) of N-BOC-(inip)-DβNal (from Example 29, step A), 132 mg (0.69 mmol, 3.0 eq) of EDC, and 47 mg (0.35 mmol, 1.5 eq) of HOBT in 15 ml of DCM/DMF (2:1) was stirred for 10 min, then 40 mg (0.25 mmol, 1.1 eq) of tryptamine and 40 μ l (0.35 mmol, 1.5 eq) of NMM were added. After 14 h at ambient temperature, the reaction mixture was partitioned between ethyl acetate and dilute citric acid and the separated organic phase was washed successively with water, saturated sodium bicarbonate, brine, dried over sodium sulfate, filtered, and evaporated. The crude product (160 mg) was chromatographed on silica (ethyl acetate/hexane (70:30), R_f = 0.5) to give 90 mg of the title compound as a crystalline solid.

IR (cm^{-1}) 3342, 3289, 3057, 2977, 2924, 2857, 1675, 1636, 1556, 1436, 1224, 1164, 739. MS (FAB, $M+H$) 569.3.

Step B: (inip)-DβNal-[N-tryptaminy] amide, TFA salt

A solution of 70 mg of N-BOC-(inip)-DβNal-[N-tryptaminy] amide (step A) in 4 ml of DCM was treated with 3 ml of TFA and stirred for 1 h. The concentrated crude product was purified by HPLC (1 x 50 cm, Vydac C-18, 20 to 35% acetonitrile in water over 60 min, 9 mL/min, 0.1% TFA, r_t = 30-48 min) to give 32 mg of the title compound. MS (electrospray, $M+H$) 469.4.

Example 37



(inip)-DβNal-DTrp-Phe-Lys-amide, TFA salt

The title compound was prepared in an identical fashion to (inip)-DβNal-DβNal-Phe-Lys-amide, TFA salt (Example 1, method B) with the only change being substitution of BOC-D-Tryptophan for BOC-DβNal in the third coupling (step C). Washing, drying, and cleavage as per the general BOC protocol above gave 250 mg of a powder. A 102 mg aliquot was purified by reverse phase HPLC (15-20μ, 300 Å, Vydac C-18, 1X 50 cm, gradient: 23 - 38% acetonitrile (0.1% TFA) in water (0.1% TFA) in 60 min at 9 mL/min, rt = 32 min) to give 23 mg of the title compound. MS (electrospray, M+H) 812.4.

Example 38

Anterior Pituitary Cell Assays
("Pit" Cell Assays)

Dispersion: Adult female Sprague-Dawley (160-180 g., Charles River) rats were group-caged in a 12:12 light:dark cycle with food and water available ad libitum. Pituitaries from ten rats were removed, the posterior pituitary discarded and the anterior pituitary placed in Hanks' Balanced Salt Solution (HBSS: without calcium chloride, w/o magnesium chloride, w/o magnesium sulfate; Gibco) containing 20 mM HEPES (Gibco) and 100 U/ml penicillin streptomycin (PS: JRH Biosciences). Under sterile conditions, pituitaries were rinsed twice then minced into small fragments with a razor blade. Fragments were resuspended in 5 ml of HBSS/HEPES containing 20 mg collagenase (Serva 17449) and 200 ml of 1 mg/ml DNase (Sigma) for a 40 min. incubation in a 37°C gyrotory water bath shaker (New Brunswick Scientific Model G76; setting 10). After the incubation, fragments were triturated to yield small clumps and single cells. The cells were centrifuged 1000 x g for 5 min, resuspended, counted and plated at a final cell concentration of 100,000 cells/ml. Incubation media was DME low glucose media w/NaHCO₃ (Gibco) containing 20 mM hepes, 100 U/ml PS and 10% FBS (Hyclone A-111-L). Cells were plated at 0.5 ml per well in 48 well plates (Falcon) and incubated at 37°C in 5% CO₂ for three

days. For challenges to determine release of other pituitary hormones cells were plated at 200,000 cells per ml with 2 ml per well of a 6-well plate (Corning).

Challenge: A (inip)bbFK-NH₂ stock (or other GHRP) concentration of 1 mM was made in DMSO and diluted with warmed media approximately 30 min prior to use. The highest concentration of DMSO in media was 0.1%. Stock solutions (1mM) of rat GHRH, somatostatin (Sigma) and GHRP antagonist HwkWfK were made fresh in media and diluted appropriately. The media used in all challenges and washing steps was DME low glucose with 20 mM Hepes, 100 U/ml PS, 10% FBS. Media was warmed to 37°C and gassed by placing it into the incubator prior to challenge. On day three, the media was discarded and fresh media (approximately 1 ml) added for the first of three washes. After the last wash, the plate was placed back in the incubator for a 15 min pre-incubation. Then cells were washed 2x (with warmed and gassed media) and fresh 0.5 ml media were added for the second 15 min. pre-incubation. After the second pre-incubation cells were washed 2x as above and 0.5 ml of control and test solutions were added for a final 15 min incubation. After this incubation, the media were removed for subsequent GH ELISA.

GH ELISA: A two-site ELISA was used to determine rat GH concentration in the media. Briefly, goat anti-rat GH antibody (lot# 19164-20) was used to coat Nunc immunoplates overnight. After blocking and washing, standard (rat GH reference preparation : Parlow) and challenge media is diluted 1:20 prior to GH assay were added for a 1 hr room temperature incubation.

STATISTICS: The mean for each group was determined and analyzed by one-way analysis of variance with a post-hoc Student-Newman-Keuls. Significance is defined as $P < 0.05$. The EC₅₀ was calculated using a 4-parameter curve-fit program (Kaleidagraph). Three to four independent EC₅₀s were used to derive the mean and SEM.

RIA of pituitary hormones: LH, FSH, TSH, and Prolactin were determined with commercially available kits from Amersham, and ACTH levels were determined by a RIA kit from ICN.

CALCIUM FLUX EXPERIMENT: Pituitary cells were plated on fibronectin (Collaborative Research) coated two-chambered slide wells (Nunc). After four days in monolayer culture cells were rinsed three times with HBSS (Gibco) in 1% BSA and 15 mM HEPES and then incubated for 30 min at 37°C with 5 μM Indo-1 AM (Molecular Probes, Eugene) in HBSS also containing 1% pluronic F127 (Molecular Probes). The cells were rinsed once and fresh media added for RT incubation. Cells were challenged within 30 min with 10 nM (inip)bbFK-NH₂, vehicle or 2.5 μM ionomycin (Sigma). Ca⁺⁺ flux was imaged with a Meridian ACAS 570 using stage

scanning at 21 second intervals. Ca^{++} bound Indo-1 was measured at 405 ± 22 nm and Ca^{++} free Indo-1 was measured at 530 ± 15 nm. The ratio of bound vs free Indo-1 was calculated and corrected with a standard curve created under identical instrument settings (Grynkiewicz G, M Peonie, RT Tsien. A new generation of Ca^{++} indicators with greatly improved fluorescence properties. *Journal of Biological Chemistry* 260: 3440-3450, [1985]).

Example 39

In vitro and *In vivo* Biological Data

Biological data for selected prior art compounds is provided in Table II

Table II

Literature code	Structure	C-term	"Pit" Cell $\text{EC}_{50}(\text{nM})$	SE	n	Rat IV $\text{ED}_{50}(\mu\text{g})$
-	Y w w F	amide	1000		3	
"GHRP 6"	H w A W f K	amide	6.2	1.5	5	1.0
"GHRP 2"	a b A W f K	amide	1.0	0.2	3	0.35
-	(Ava) b A W f K	amide	0.2	0.03	3	1.5
"GHRP 1"	A H b A W f K	amide	1.4		2	
L-692,429	benzo-fused lactam		26.2	5.3	5	100
L-692,585	benzo-fused lactam		10.6	4.0	4	10

The following selected *in vitro* and *in vivo* biological data for compounds represented by formula IV is provided in Table III

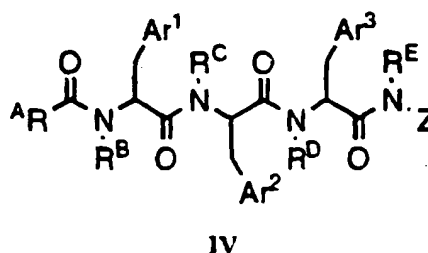


Table III

	Structure	C-term	PIT Cell EC ₅₀ (nM)	SE	n	RAT IV ED ₅₀ (µg)
	Y w w F K	amide	1000		3	
5	H w w F K	amide	25.8	3.9	3	50
	H b w F K	amide	6.8	1.2	3	4.2
	H w b F K	amide	15.0	5.0	3	50
	H b b F K	amide	2.4	1.2	3	15.4
10	G w w F K	amide	2.1	0.3	3	2.5
	(Ava) w w F K	amide	13.0	2.6	4	5
	a w w F K	amide	4.1	2	3	1.47
	G b w F K	amide	0.8		2	4.8
	(Ava) b w F K	amide	4.6	1.5	6	11.8
15	H w w nmF K	amide	37.3	11.3	3	
	G b b F K	amide	14.9		2	100
	bA b b F K	amide	4.8	3.3	3	6.7
	(Ab) b b F K	amide	0.4	0.1	3	1
	(Ava) b b F K	amide	2.8	0.4	4	2.6
20	(ahx) b b F K	amide	1000		2	250
	(ahp) b b F K	amide	34.3	8.2	3	100
	(nba) b b F K	amide	2.8		2	100
	(inip) b b F K	amide	0.18	0.04	3	0.2
	(pyc) b b F K	amide	>300		4	200
25	(pac) b b F K	amide	100		2	
	(Ava) b b F G	amide	7.2	1.9	4	50
	(Ava) b b F N	amide	7.7	4.5	3	200
	(Ava) b b F n	amide	10		1	12.5
	(Ava) b b F k	amide	17		1	33.3
30	(Ava) b b F P	amide	51		1	50
	b b F K	amide	1000		2	
	(Ava) b b F K Y	amide	13.6	2.6	3	20
	(Ava) b nmb F K	amide	3.8	1.4	3	100
35	(Ava) nmb b F K	amide	1.8	0.6	3	0.4
	(Ava) b b B K	amide	0.7		1	50
	(Ava) b b hF K	amide	5.0	1.7	3	50

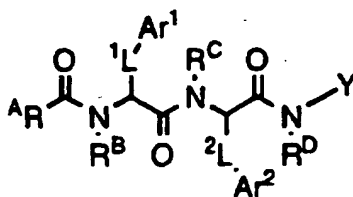
Table III (cont.)

	Structure	C-term	PIT Cell EC ₅₀ (nM)	SE	n	RAT IV ED ₅₀ (µg)
5	(Ava) a b F K	amide	>300		2	
	(Ava) b b chA K	amide	44.3	9.5	3	
	(Ava) b b (Pg) K	amide	>100		1	
	(Ava) b b (pG) K	amide	>100		1	
10	(inip) b b npA K	amide	0.1		2	1
	(Ava) b b A K	amide	>150		2	
	(Ava) b a F K	amide	>300		3	
	a b b F K	amide	3.5	1.5	3	10
15	A b b F K	amide	2.1	0.8	4	10
	(mab) b b F K	amide	1.4		1	0.7
	(inip) nmb nmb F K	amide	1.4		1	
	(inip) nmb b F K	amide	0.27	0.06	4	0.4
20	(inip) b nmb F K	amide	0.2		2	1
	(inip) b b F K Y	amide	0.43	0.01	4	1
	(inip) b b F bA	amide	0.2	0.05	4	2
	nmb b F K	amide	>100		1	1000
25	(Ab) nmb b F K	amide	0.5		2	1
	(inip) nmb w F K	amide	0.2		2	0.2
	(inip) b b F K GGSGGSY	amide	0.4		2	
	(inip) nmb b F dam		6.0		1	
30	(inip) nmb b F mor		0.8		1	10
	nmb b Y K	amide	1000		4	500
	(Ava) b b npA K	amide	1000		1	
	(cho) b b F K	amide	1000		1	
35	(Ab) b b B ram		2.4	1.1	3	10
	(inip) b b f dam		12		1	
	(inip) b b F dam		12		1	10
	(inip) b b f mam		7.0		1	
35	(inip) b b F mam		2.0		1	2
	(inip) b b F tbn		10		1	10
	(inip) b b Y(I) K	amide				
	(inip) b b Y K	amide	0.2		1	

Table III (cont.)

Structure	C-term	PIT Cell EC ₅₀ (nM)	SE	n	RAT IV ED ₅₀ (μg)
(inip) b b F K Y(I)	amide	0.69	0.54	3	
(inip) b w F K	amide	1.1	0.5	3	0.070

The following selected *in vitro* and *in vivo* biological data for compounds represented by formula III is provided in Table IV



III

Table IV

Structure	C-term	PIT Cell EC ₅₀ (nM)	SE	n	RAT IV ED ₅₀ (μg)
H w w F	amide	1000		4	
(Ava) f f F	amide	1000		1	
(Ava) f w F	amide	>300		3	1000
(Ava) b b F	amide	17.1	5.7	3	100
(Ava) b b fem		>200		4	
(inip) b b bam		18.5		2	3.3
(Ava) b F K	amide	>300		4	
(inip) b b fem		2.04	0.9	3	
(inip) b b F	amide	0.28	0.07	5	1.7
(inip) nmb b F	amide	0.5	0.2	4	5
(inip) nl b F	amide	101		1	
(inip) b nl F	amide	72		1	
(inip) b b nL	amide	4.3		2	3.3
(inip) b b P	amide	9		1	5
(inip) b b amF	amide	0.5		1	5
(inip) b b nmF	amide	0.6		2	5
(inip) nmb b F	acid	1.4		1	5

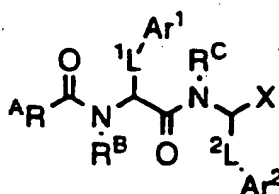
Table IV (cont.)

	Structure	C-term	PIT Cell EC50(nM)	SE	n	RAT IV ED50 (μ g)
5	(inip) nmb b F	Me ester	1.4		2	10
	(inip) nmb B F		>100		1	
	(inip) b b feg	amide	0.25	0.19	3	1
	(inip) b b feb	amide	0.3		2	0.67
	(inip) b b fbd		0.9		2	0.67
10	(inip) b b hcF	amide	0.2	0.09	3	2.5
	(inip) hf b F	amide	27		1	
	(inip) b hf F	amide	14		1	10
	(inip) nmb B F	acid	7		1	100
	(inip) b b npA	amide	0.2		1	5
15	(inip) b b B	amide	0.1		2	50
	(Ab) b b F	amide	3.1		1	10
	(inip) b b Pol		0.2	0.02	3	5
	(inip) b chf F	amide	2.7		1	5
	(inip) chf b F	amide	2.2		1	10
20	(inip) b nmb F	amide	1		2	5
	(inip) b b bA	amide	10		1	10
	(inip) b b F	Me ester	2.8		1	2
	(inip) b b F	acid	0.2		2	2
	(inip) w w F	amide	0.9		2	5
25	(inip) b w F	amide	0.1		2	0.67
	(inip) b b Abx	amide	13		1	100
	(inip) b Abx F	amide	>100		1	
	(inip) b b tam		11		1	20
	(inip) b b ram		16		1	5
30	(inip) b b pam		43		1	
	(inip) b b cxa		14		1	10
	(inip) b b ppz		10		1	10
	(inip) b b ab	amide	10		1	10
	(inip) b b apc		39		1	
35	(inip) b b api		2.7		2	25
	(inip) b b O	amide	5.9		2	1

Table IV (cont.)

Structure	C-term	PIT Cell EC ₅₀ (nM)	SE	n	RAT IV ED ₅₀ (μg)
(inip) b b o	amide	10		1	
(inip) b b Y	acid	5		1	10
(inip) b nmb bam		0.3		2	1.67
(inip) nmb B bam		1000		1	
(inip) nmb b bam		n/d			
(inip) b nmb api		0.5		2	2
(inip) b b Y(I)	acid	0.1		2	

The following selected *in vitro* and *in vivo* biological data for compounds represented by formula II is provided in Table V



II

Table V

Structure	C-term	PIT Cell EC ₅₀ (nM)	SE	n	RAT IV ED ₅₀ (μg)
(inip) b b	amide	17.6	5.6	3	
(inip) b Bmn		36		1	20
(inip) b b	acid	82		1	
(inip) b bol		2		2	10
(inip) b b	Me ester	5		1	100
(inip) b mbm		7		1	20
(inip) b Bol		100		4	
(inip) B Bol		1000		1	
(inip) B bol		1000		1	
(inip) b wol		10.6	6.2	3	0.8
(inip) w wol		4		1	20
(inip) w bol		1000		1	

Table V (cont.)

Structure	C-term	PIT Cell EC ₅₀ (nM)	SE	n	RAT IV ED ₅₀ (μg)
(inip) b npe		11		2	500
(inip) b men		12		2	500
(inip) b man		9		1	500
(inip) b tam		1000		1	
(Ava) b bol		190		1	
(inip) b nbol		3		1	20
(amb) b bol		1000		1	
(inip) b miz		12		1	
(inip) b mbm Ac		90		2	
Boc (inip) b mbm		13		1	
(inip) b tra		100		2	100

The following selected *in vitro* and *in vivo* biological data for the "retroinverso" compounds represented by formula V is provided in Table VI

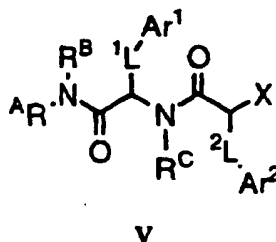


Table VI

Structure	C-term	PIT Cell EC ₅₀ (nM)	SE	n	RAT IV ED ₅₀ (μg)
(dhc) B B bam		>200		3	
(Ava) f B B K	amide	36		1	
(Ava) f B B bam		22		1	100
(Ab) b B B ram		2		2	
b B B ram		15.2		1	100
Ac b B B ram		4.8		2	100
(inip) B B api		96		2	
(inip) B B bam		1000		1	

Example 40**GHRP Induced Growth Hormone Secretion: Intravenous Administration**

Immature weanling female Sprague Dawley rats were purchased from Charles River
5 Labs (Portage, Oregon) and group housed with water and food available ad libitum. When the
rats were 24-30 days old (weighing 50-90 g) they were anesthetized with pentobarbitone (4 mg
in 0.5 ml, approximately 60 mg/kg) given by intraperitoneal injection. The rats were then
placed briefly on a heated pad, to distend their tail veins, and given an intravenous tail vein
10 injection of the peptides 20 minutes after receiving the anesthetic. The intravenous injection
was of 0.1 ml using a 1 ml syringe. The injections contained graded doses of peptides or the
vehicle (vehicle for all peptides given intravenously was a buffer of 20 mM sodium acetate, 45
g/l mannitol, pH 5.0). Ten minutes after the intravenous injection blood was taken by cardiac
puncture, using a 3 ml syringe, and the rats were then sacrificed.
The blood was then clotted on ice, centrifuged, serum decanted and frozen for subsequent
15 analysis using the rat GH ELISA described elsewhere in the application. For the rat GH ELISA
the serum was diluted 1:50 or 1:250, depending on the expected serum GH concentrations
achieved, and assayed in duplicate.

20

Example 41**GHRP Induced Dose Dependent Weight Gain in Rats**

Methods: Forty normal Sprague Dawley female rats (Supplier, Charles River, 90 days of age,
average weight 200 g) were group housed in a room controlled for temperature and lighting
25 and fed a standard pelleted rat diet and tap water ad libitum. The rats were weighed on the
day of surgery (see below) and randomized into 5 groups of 8/group using a grouping
program.

The GHRP (inip) b b F K-NH₂, was dissolved in a sodium acetate (20 mM) buffer (pH
5.0) containing mannitol (45 g/l) at 8 g/l, 1.6 g/l and 0.33 g/l. Rat GHRH (1-43) was dissolved
30 in the same buffer at 25 g/l. Osmotic minipumps (Alza, Palo Alto, model 2002, pump rate 0.52
μl/hr for 14 days, fill volume 230 μl) were filled with these solutions (1/rat for (inip) b b F K-
NH₂ and 2/rat for rat GHRH); a fifth set of pumps were filled with the sodium acetate buffer.
All the pumps were primed by being incubated in isotonic saline overnight in a refrigerator.

The next day these osmotic pumps were inserted into rats. To do this the rats were
35 anesthetized with ketamine/xylazine (62.5 and 12.5 mg/kg/rat, respectively, by i.p. injection).
The dorsal neck was then shaved, swabbed with betadine solution and cleaned with alcohol.
An incision was then made on the dorsal neck and the a subcutaneous pocket created caudally

by blunt dissection. The pumps were then inserted into the pocket with the end of the pump delivering the solution positioned away from the incision. The incision was then closed with wound clips, the rat placed on a heated pad and when ambulatory was returned to its home cage.

5 The rats were then weighed every day and on day 14 they were sacrificed using inhalation of carbon dioxide. They were then bled from the heart and organs taken. The pituitary, spleen, heart, kidney, liver, and thymus were taken and weighed while the tibias were removed and placed in 10% formalin for subsequent histological evaluation. To do this the tibias were sectioned longitudinally and the width of the epiphyseal plate was measured
10 using a microscope fitted with an ocular micrometer.

Serum chemistries were measured by standard automated procedures. Serum insulin-like growth factor-1 (IGF-1) was measured by radioimmunoassay, using an antibody raised in rabbits, after acid ethanol extraction to remove the IGF-1 binding protein.

Statistical significance was gauged by analysis of variance, which if significant ($p < 0.05$)
15 was followed by a Duncan's New Multiple Range Test to test for differences between the individual treatment groups. Data are presented as mean \pm standard error of the mean with 8 rats per group.

The body weight gains plotted against time for the 5 treatment groups are shown in Figure 19. Both (inip) b b F K-NH₂ and rat GHRH induced significant body weight and organ
20 weight gain compared to the vehicle treated rats.

Example 42

Comparison of SC injections and SC infusions

25 Methods: Forty normal Sprague Dawley female rats (supplier Charles River, 150 days of age, average weight 280 g) were group housed in a room controlled for temperature and lighting and fed a standard pelleted rat diet and tap water ad libitum. The rats were weighed on the day of surgery (see below) and randomized into 5 groups of 8/group using a grouping
30 program.

GHRP (inip) b b F K-NH₂ was dissolved in a sodium acetate (20 mM) buffer (pH 5.0) containing mannitol (45 g/l) at 8 g/l and 1.6 g/l to fill the minipumps and at 0.5 and 0.1 g/l for the injection solutions. Osmotic minipumps (Alza, Palo Alto, model 2002, pump rate 0.52 μ l/hr for 14 days, fill volume 230 μ l, one per rat) were filled with these solutions; a fifth set of
35 pumps were filled with the sodium acetate buffer. All the pumps were primed by being incubated in isotonic saline overnight in a refrigerator.

The next day osmotic pumps were inserted into all rats. To do this the rats were anesthetized with ketamine/xylazine (62.5 and 12.5 mg/kg/rat, respectively, by i.p. injection).

The dorsal neck was then shaved, swabbed with betadine solution and cleaned with alcohol. An incision was then made on the dorsal neck and the a subcutaneous pocket created caudally by blunt dissection. The pumps were then inserted into the pocket with the end of the pump delivering the solution positioned away from the incision. The incision was then closed with wound clips, the rat placed on a heated pad and when ambulatory was returned to its home cage.

The treatment groups were;

- 1) Excipient pump, excipient injections 2 times a day.
- 10 2) (inip) b b F K-NH₂ pump (100 µg/day), excipient injections 2 times a day.
- 3) (inip) b b F K-NH₂ pump (20 µg/day), excipient injections 2 times a day.
- 4) Excipient pump, (inip) b b F K-NH₂ injections 50 µg 2 times a day.
- 5) Excipient pump, (inip) b b F K-NH₂ injections 10 µg 2 times a day.

15 The rats were then weighed every day and injected twice daily with either excipient or the two doses of (inip) b b F K-NH₂. On day 14 they were sacrificed using inhalation of carbon dioxide. They were then bled from the heart and organs taken. The rats were skinned and eviscerated to weigh the amount of skin, muscle and bone (the carcass). The pituitary, spleen, heart, kidney, liver, thymus and the soleus muscle were also taken and weighed while the
20 tibias were removed and placed in 10% formalin for subsequent histological evaluation. The tibias were sectioned longitudinally and the width of the epiphyseal plate was measured using a microscope fitted with an ocular micrometer.

Serum chemistries were measured using standard automated techniques. Serum insulin-like growth factor-1 (IGF-1) was measured by radioimmunoassay, using an antibody
25 raised in rabbits, after acid ethanol extraction to remove the IGF-1 binding protein.

Statistical significance was gauged by analysis of variance, which if significant ($p < 0.05$) was followed by a Duncan's New Multiple Range Test to test for differences between the individual treatment groups. Data are presented as mean \pm standard error of the mean with 8 rats per group.

30 (inip) b b F K-NH₂ at 20 and 100 µg/day, delivered by both injection and infusion, induced significant body weight gain compared to vehicle treated rats. The dose-related nature of the body weight gains to injections of (inip) b b F K-NH₂ can be seen in Figure 20. In contrast there were similar weight gains in response to infusions of both 20 and 100 µg/day of (inip) b b F K-NH₂. In addition there were very different patterns of weight gain in response to
35 infusions or injections of 100 µg/day of (inip) b b F K-NH₂ as can be seen in Figure 21.

Example 43

Combination GHRP and IGF-1 Treatment of Obese Rats

Methods: Forty-eight (48) obese male Zucker Diabetic Fatty (ZDF) rats (Genetic Models Inc., Indianapolis, IN 46268) 6 weeks of age were group housed in a room controlled for temperature and lighting and fed a standard pelleted rat diet and tap water *ad libitum*. The rats were weighed on the day of surgery (see below) and randomized into 6 groups of 8/group using a grouping program. Ten lean ZDF rats served as an additional control group.

GHRP (inip) b b F K-NH₂ was dissolved in a sodium acetate (20 mM) buffer (pH 5.0) containing mannitol (45 g/l) at 0.5 g/l. This GHRP was given by sc. injection twice daily, each dose of 100 µl therefore containing 50 µg/injection or 100 µg/day.

Recombinant human IGF-1 (rhIGF-1) at 13.8 mg/ml in acetate buffer was loaded into osmotic minipumps (Alza, Palo Alto, model 2ML4, pump rate 2.29 µl/hr for 28 days, fill volume 2064 µl). Other pumps were filled with acetate buffer. The pumps were primed by being incubated in isotonic saline overnight in a refrigerator. The delivered dose of rhIGF-1 was therefore 758 µg/day. Recombinant human growth hormone (rhGH, Lot R9092AX, Genentech Inc.) was diluted in sterile water to 2.5 g/l and a 100 µl injection given twice daily (250 µg/injection, or 500 µg/day).

The next day the osmotic pumps were inserted into rats. To do this the rats were anesthetized with ketamine/xylazine (62.5 and 12.5 mg/kg/rat, respectively, by i.p. injection). The dorsal neck was then shaved, swabbed with betadine solution and cleaned with alcohol. An incision was then made on the dorsal neck and the a subcutaneous pocket created caudally by blunt dissection. The pumps were then inserted into the pocket with the end of the pump delivering the solution positioned away from the incision. All rats not receiving rhIGF-1 containing pumps were implanted with pumps delivering the acetate buffer excipient. The incision was then closed with wound clips, the rat placed on a heated pad and when ambulatory was returned to its home cage.

The rats were then weighed every day, and injected twice daily with either active drug (GH or GHRP) or vehicle excipient. On day 24 blood was withdrawn after a 4 hour fast and 1.5 U/kg of regular insulin was injected i.p. and a second blood sample taken 30 minutes later. The rats were then sacrificed using carbon dioxide, bled from the heart, and organs taken. Serum glucose was measured by standard automated procedures.

Statistical significance was gauged by analysis of variance, which if significant ($p < 0.05$) was followed by a Duncan's New Multiple Range Test to test for differences between the individual treatment groups. Data are presented as mean \pm standard error (SE) of the mean with 8 rats per group.

Body Weight Gain: The body weight gains plotted against time for all treatment groups over the whole study are shown in Figure 22 and for the first 7 days for the GHRP (inip)

b b F K-NH₂ and IGF-1 treatment groups are shown in Figure 23. The basal blood glucose values plotted against time for all the treatment groups for the entire experiment are shown in Figure 24 and the blood glucose responses to an intravenous insulin challenge at the end of the experiment are shown in Figure 25.

5

Example 44

Combination GHRP and IGF-1 Treatment of Normal Rats

Methods: Sixty normal adult female SD rats (Supplier, Charles River, 120 days of age, 10 250 to 320 g) were group housed in a room controlled for temperature and lighting and fed a standard pelleted rat diet and tap water *ad libitum*. The rats were weighed on the day of surgery and randomized into 12 groups of 5/group using a grouping program.

The GH secretagogues (GHRPs and GHRH) were dissolved in a sodium acetate (20 mM) buffer (pH 5.0) containing mannitol (45 g/l) at 0.5 g/l. The GH secretagogues were given 15 by sc. injection twice daily, each dose of 100 µl. Different doses of the molecules were given based on their potency in the IV assay (for example L-692,585 was given at 3-fold higher doses as it was less the least potent of the secretagogues). Recombinant human IGF-1 (rhIGF-1) at 2.5 mg/ml in acetate buffer was loaded into osmotic minipumps (Alza, Palo Alto, model 2ML1, pump rate 10.16 µl/hr for 7 days, fill volume 2086 µl). Other pumps were filled with acetate 20 buffer. The pumps were primed by being incubated in isotonic saline overnight in a refrigerator. The delivered dose of rhIGF-1 was therefore 610 µg/day.

The next day the osmotic pumps were inserted into rats. To do this the rats were anesthetized with ketamine/xylazine (62.5 and 12.5 mg/kg/rat, respectively, by i.p. injection). The dorsal neck was then shaved, swabbed with betadine solution and cleaned with alcohol. 25 An incision was then made on the dorsal neck and the a subcutaneous pocket created caudally by blunt dissection. The pumps were then inserted into the pocket with the end of the pump delivering the solution positioned away from the incision. All rats not receiving rhIGF-1 containing pumps were implanted with pumps delivering the acetate buffer excipient. The incision was then closed with wound clips, the rat placed on a heated pad and then when it 30 was ambulatory returned to its home cage. The treatment groups were;

- | | | |
|----------------------------------------------|----------------|----------------|
| 1) Excipient | 2 injections/d | Excipient pump |
| 2) Excipient | 2 injections/d | IGF-1 Pump |
| 3) GHRH (300 µg/dose) | 2 injections/d | Excipient pump |
| 35 4) GHRH (300 µg/dose) | 2 injections/d | IGF-1 pump |
| 5) GHRP-6 (50 µg/dose) | 2 injections/d | Excipient pump |
| 6) GHRP-6 (50 µg/dose) | 2 injections/d | IGF-1 pump |
| 7) (inip) b b F-NH ₂ (50 µg/dose) | 2 injections/d | Excipient pump |

- | | | |
|----------------------------------------------|----------------|----------------|
| 8) (inip) b b F-NH ₂ (50 µg/dose) | 2 injections/d | IGF-1 pump |
| 9) (inip) b nmb bam (50 µg/dose) | 2 injections/d | Excipient pump |
| 10) (inip) b nmb bam (50 µg/dose) | 2 injections/d | IGF-1 pump |
| 11) L-692,585 (150 µg/dose) | 2 injections/d | Excipient pump |
| 5 12) L-692,585 (150 µg/dose) | 2 injections/d | IGF-1 pump |

10 The rats were then weighed every day, and injected twice daily with either active drug (GHRH or GHRP) or vehicle excipient). The rats were sacrificed using carbon dioxide, bled from the heart and organs taken. Serum chemistries were measured by standard automated procedures.

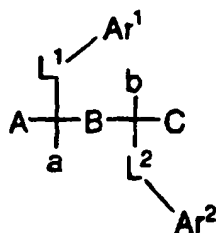
Statistical significance was gauged by analysis of variance, which if significant ($p < 0.05$) was followed by a Duncan's New Multiple Range Test to test for differences between the individual treatment groups. Data are presented as mean \pm standard error of the mean with 8 rats per group.

15 Body Weight Gain: The body weight gains plotted against time for the groups treated only with the GH secretagogues are shown in Figure 26. The responses to the combination of the GH secretagogues and IGF-1 tended to be greater than to IGF-1 alone (Figure 27).

20 All references described herein are expressly incorporated by reference.

WHAT IS CLAIMED IS:

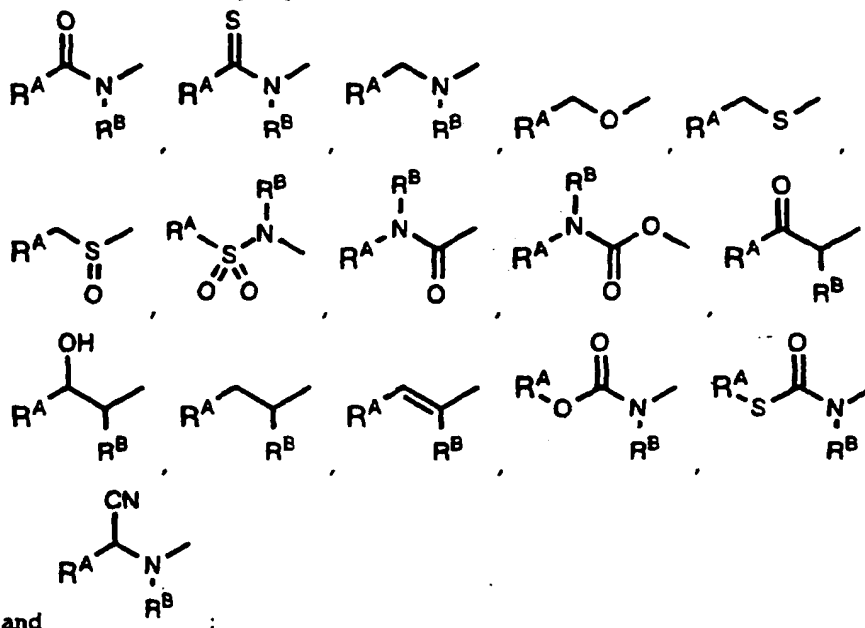
1. A compound represented by structural Formula (I)



I

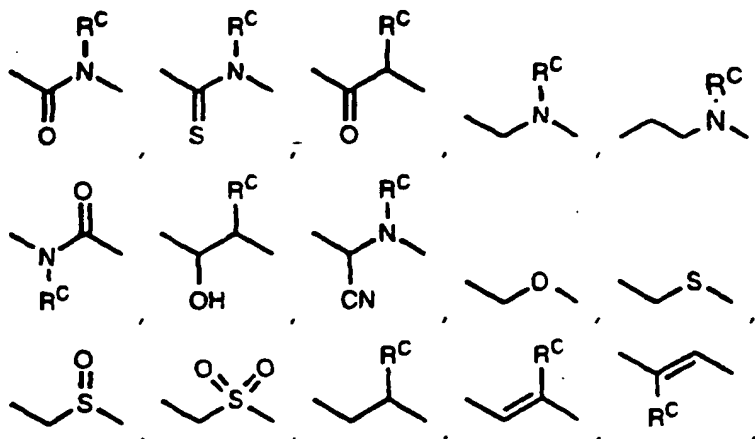
where

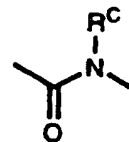
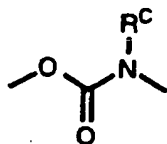
A is selected from the group



and

B is selected from the group





, and C₁-C₆alkyl substituted with

5

B may optionally be selected from the group

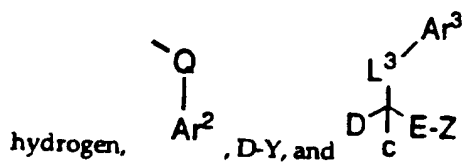
a covalent bond, and

C₁-C₃alkyl,

when L² is -N(R^C)-Q;

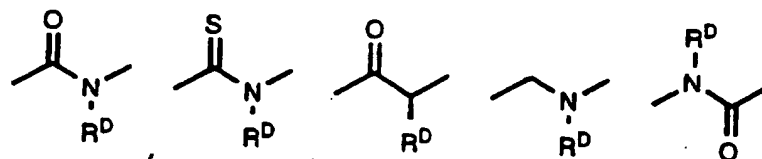
C is selected from the group

10

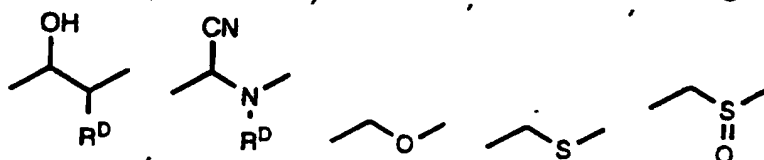


D is selected from the group

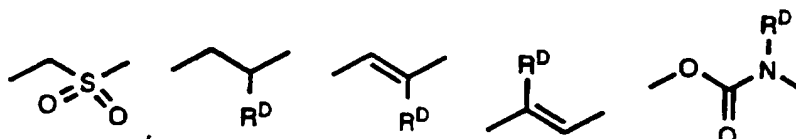
15



20



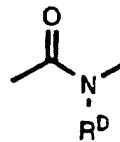
25



and

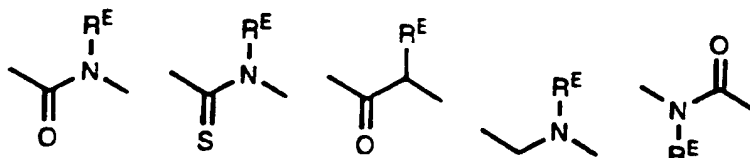
30

C₁-C₆alkyl substituted with



E is selected from the group

35



pair together with the carbon to which they are attached may
independently form a 5 or 6 member carbocyclic ring;

a, b and c are independently selected from

hydrogen, and

5 C₁-C₆alkyl;

n and o are independently 1, 2 or 3;

L¹ is selected from

-CH₂-O-,

-CH₂-CH₂-O-

10 -CH₂-,

-CH₂-CH₂-, and

-CH₂-CH₂-CH₂-;

L² and L³ are independently selected from

a covalent bond,

15 -O-,

-O-CH₂-,

-N(R^C)-Q and

L¹;

Q is selected from the group

20 -L²-,

-S(=O)₂-L²-,

-C(=O)-,

-C(=O)-O-,

-CH(X)-, and

25 -CH(X)-CH₂-;

R^A is selected from the group

C₀-C₃alkyl-heterocycle where the heterocycle comprises a mono-, bi-, or

tricycle containing 5-12 ring atoms, one or two of which are

heteroatoms selected from O, S, and N, provided at least one

30 heteroatom is N, where any N atom is optionally substituted with

R¹,

C₀-C₆alkyl substituted with one or two substituents selected from

the group

NR²R³,

35 imidazoliny,

pyridiny,

dihydropyridiny, and

piperidiny;

R^B , R^C and R^D are selected from the group

R^A ,
 L^1-Ar^1 ,
 L^2-Ar^2 ,
5 hydrogen,
 C_1-C_6alkyl , and
halo(F, Cl, Br, I) C_1-C_6alkyl ;

R^A and R^B together with the N to which they are bonded may form a 5- or
6-member heterocycle, optionally containing one additional hetero
10 atom selected from O, S, and N where any N is optionally
substituted with R^1 , any carbon is optionally substituted with R^6
and where the heterocycle is optionally fused to a phenyl ring,
optionally substituted with R^4 ;

R^1 is selected from

15 hydrogen,
 C_1-C_6alkyl ,
 $C(=O)-C_1-C_6alkyl$,
 $C(=O)-NR^2R^3$,
 $C(=NR^2)-NR^2R^3$,
20 $C(=O)O-C_1-C_6alkyl$,
halo(F, Cl, Br, I) C_1-C_6alkyl , and
 C_2-C_6alkyl substituted with 1-3 hydroxyl groups;

R^2 and R^3 are independently selected from

R^1 , and
25 piperidinyl;

R^2 and R^3 together with the N to which they are bonded may form a 5- or
6-member heterocycle, optionally containing one additional hetero
30 atom selected from O, S, and N where any N is optionally
substituted with R^1 , any carbon is optionally substituted with R^6
and where the heterocycle is optionally fused to a phenyl ring,
optionally substituted with R^4 ;

R^4 and R^5 are independently selected from the group

hydrogen,
halo(F, Cl, Br, and I),
35 cyano,
amino,
amido,

nitro,

hydroxy,

C₁-C₆alkyl optionally substituted with 1-3 R⁶,

C₂-C₆alkynyl optionally substituted with 1-3 R⁶,

5 C₁-C₆alkyloxy optionally substituted with 1-3 R⁶,

C₁-C₆acylamino optionally substituted with 1-3 R⁶,

C₁-C₆alkylcarbonyl optionally substituted with 1-3 R⁶,

C₁-C₆alkyloxycarbonyl optionally substituted with 1-3 R⁶,

10 N-(C₁-C₆alkyl),N-(C₁-C₆acyl)amino optionally substituted with 1-3 R⁶,

N-(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶,

N,N-di(C₀-C₆alkyl)amino optionally substituted with 1-3 R⁶,

N,N-di(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶,

15 C₁-C₄perfluoroalkyl, and

C₁-C₃perfluoroalkoxy;

R⁶ is selected from the group

COOR²,

O(C=O)R²,

20 CONR²R³,

cyano,

NR²R³,

NR²COR³,

azido,

25 nitro, and

hydroxy;

R⁷ is selected from the group

R⁶,

C₆-C₁₀aryl optionally substituted with

30 halo(F, Cl, Br, and I),

cyano,

amino,

amido,

nitro,

35 hydroxy,

C₁-C₄perfluoroalkyl, and

C₁-C₃perfluoroalkoxy;

X is selected from the group

hydrogen,

C₁-C₆alkyl optionally substituted with 1-3 R⁶, and

C₁-C₆acyl optionally substituted with a group selected from

L²-Ar²,

R^A, and

R⁶;

Y is selected from the group

-(C=O)-R^A,

C₁-C₆alkyl substituted with 1-2 R⁷,

C₂-C₆alkynyl optionally substituted with 1-2 R⁷,

C₂-C₆alkenyl optionally substituted with 1-2 R⁷, and

C₁-C₆alkoxy optionally substituted with 1-2 R⁷,

Y and R^D together with the N to which they are bonded may form a 5- or 6-member heterocycle, optionally containing one additional hetero atom selected from O, S, and N where any N is optionally substituted with R¹, any carbon is optionally substituted with R⁷ and where the heterocycle is optionally fused to a phenyl ring;

Z is selected from the group

C₁-C₆alkyl substituted with 1-2 R⁷,

C₂-C₆alkynyl optionally substituted with 1-2 R⁷,

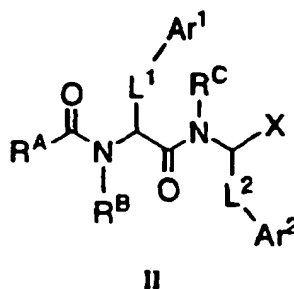
C₂-C₆alkenyl optionally substituted with 1-2 R⁷,

C₁-C₆alkoxy optionally substituted with 1-2 R⁷, and

piperidiny; and

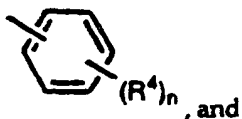
pharmaceutically acceptable salts thereof.

2. The compound of Claim 1 represented by Formula II

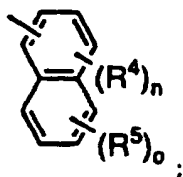


where

Ar¹ and Ar² are each independently selected from indoyl,



5



n and o are independently 1, 2 or 3;

L^1 is selected from

10

$-\text{CH}_2-\text{O}-$,
 $-\text{CH}_2-\text{CH}_2-\text{O}-$
 $-\text{CH}_2-$,
 $-\text{CH}_2-\text{CH}_2-$, and
 $-\text{CH}_2-\text{CH}_2-\text{CH}_2-$;

15

L^2 is selected from

a covalent bond,
 $-\text{O}-$,
 $-\text{O}-\text{CH}_2-$, and
 L^1 ;

20

R^A is selected from the group

$\text{C}_0\text{-C}_3\text{alkyl-heterocycle}$,
 $\text{O-C}_0\text{-C}_3\text{alkyl-heterocycle}$, and
 $\text{NR}^2\text{-C}_2\text{-C}_6\text{alkyl-heterocycle}$,

25

where the heterocycle comprises a mono-, bi-, or tricycle
 containing 5-12 ring atoms, one or two of which are
 heteroatoms selected from O, S, and N, provided at least one
 heteroatom is N, where any N atom is optionally
 substituted with R^1 ,

30

$\text{C}_0\text{-C}_6\text{alkyl}$ substituted with one or two substituents,
 $\text{O-C}_2\text{-C}_6\text{alkyl}$ substituted with one or two substituents, and
 $\text{NR}^2\text{-C}_2\text{-C}_6\text{alkyl}$ substituted with one or two substituents

35

where the substituents are selected from the group
 NR^2R^3 ,
 imidazoliny!,
 pyridiny!,
 dihydropyridiny!, and
 piperidiny!;

R^B and R^C are selected from the group

hydrogen,

C_1 - C_6 alkyl optionally substituted with a group selected from

NR^2R^3 , and

5 phenyl- C_1 - C_3 - NR^2R^3 , and

halo(F, Cl, Br, I)- C_1 - C_6 alkyl;

R^1 is selected from

hydrogen,

C_1 - C_6 alkyl,

10 $C(=O)$ - C_1 - C_6 alkyl,

$C(=O)$ - NR^2R^3 ,

$C(=NR^2)$ - NR^2R^3 ,

$C(=O)O$ - C_1 - C_6 alkyl,

halo(F, Cl, Br, I)- C_1 - C_6 alkyl, and

15 C_1 - C_6 alkyl substituted with 1-3 hydroxyl groups;

R^2 and R^3 are independently selected from

C_1 - C_6 alkyl- NH_2 ,

C_1 - C_6 alkyl-heterocycle,

C_1 - C_6 alkyl- NH - C_1 - C_6 alkyl,

20 C_1 - C_6 alkyl- N -(di- C_1 - C_6 alkyl),

R^1 , and

piperidinyl;

R^2 and R^3 together with the N to which they are bonded may form a 5- or 6-member

heterocycle, optionally containing one additional hetero atom selected from O, S, and N where

25 any N is optionally substituted with R^1 , any carbon is optionally substituted with R^6 , and

where the heterocycle is optionally fused to a phenyl ring, optionally substituted with R^4 ;

R^4 and R^5 are independently selected from the group

hydrogen,

halo(F, Cl, Br, and I),

30 cyano,

amino,

amido,

nitro,

hydroxy,

35 C_1 - C_6 alkyl optionally substituted with 1-3 R^6 ,

C_2 - C_6 alkynyl optionally substituted with 1-3 R^6 ,

C_1 - C_6 alkyloxy optionally substituted with 1-3 R^6 .

C₁-C₆acylamino optionally substituted with 1-3 R⁶,
 C₁-C₆alkylcarbonyl optionally substituted with 1-3 R⁶,
 C₁-C₆alkyloxycarbonyl optionally substituted with 1-3 R⁶,
 N-(C₁-C₆alkyl),N-(C₁-C₆acyl)amino optionally substituted with 1-
 3 R⁶,
 N-(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶,
 N,N-di(C₀-C₆alkyl)amino optionally substituted with 1-3 R⁶,
 N,N-di(C₁-C₆alkyl)carboxamido optionally substituted with 1-3
 R⁶,
 C₁-C₄perfluoroalkyl, and
 C₁-C₃perfluoroalkoxy;

R⁶ is selected from the group

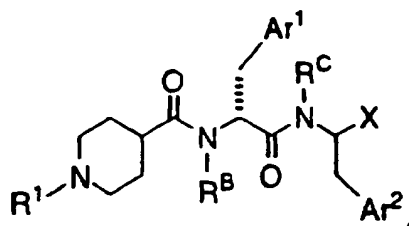
COOR²,
 CONR²R³,
 cyano,
 NR²R³,
 NR²COR³,
 azido,
 nitro, and
 hydroxy;

X is selected from the group

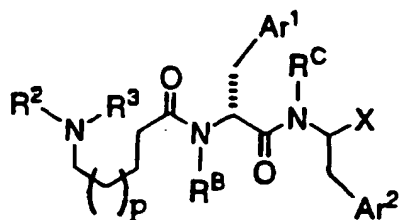
hydrogen,
 oxo (=O),
 COOR²,
 CONR²R³,
 C₀-C₆alkyl-O-C₁-C₆alkyl optionally substituted with 1-2 R⁶, and
 C₁-C₆alkyl optionally substituted with 1-2 R⁶; and

pharmaceutically acceptable salts thereof.

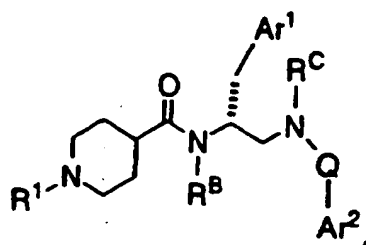
3. The compound of Claim 1 represented by Formula IIa-IIg



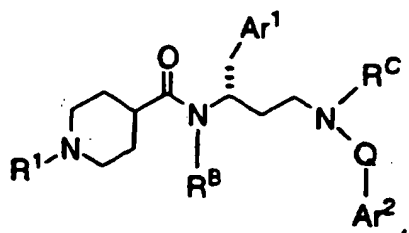
IIa



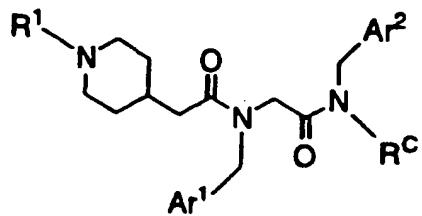
IIb



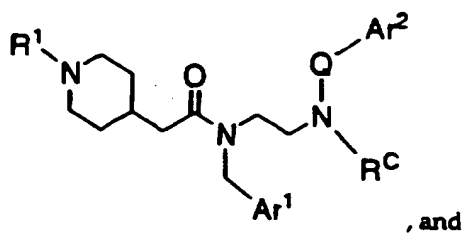
IIc



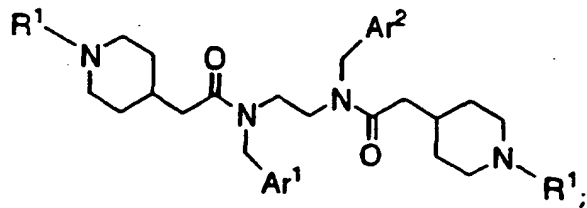
IIId



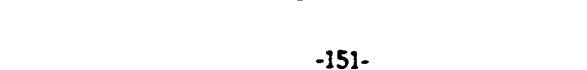
IIe



IIIf



IIg

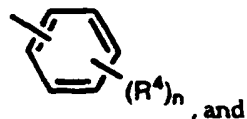


where

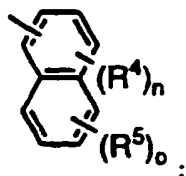
Ar¹ and Ar² are each independently selected from

indoyl,

5



10



n and o are independently 1, 2 or 3;

p is 0, 1, or 2;

L² is selected from

15

a covalent bond,

-O-,

-O-CH₂-,

-CH₂-O-,

-CH₂-CH₂-O-

-CH₂-,

20

-CH₂-CH₂-, and

-CH₂-CH₂-CH₂-;

R^B and R^C are selected from the group

hydrogen,

C₁-C₆alkyl optionally substituted with a group selected from

25

NR²R³, and

phenyl-C₁-C₃-NR²R³, and

halo(F, Cl, Br, I)C₁-C₆alkyl;

R¹ is selected from

30

hydrogen,

C₁-C₆alkyl,

C(=O)-C₁-C₆alkyl,

C(=O)-NR²R³,

C(=NR²)-NR²R³,

C(=O)O-C₁-C₆alkyl, and

35

halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ are independently selected from

hydrogen,

C₁-C₆alkyl,

piperidinyl, and
halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ together with the nitrogen to which they are attached may form
a optionally mono- or di-substituted ring selected from

5 piperidinyl,
pyrrolydinyl,
pyrryl,
imidazolyl,
piperazinyl, and
10 morpholinyl

where the substituents are selected from C₁-C₃alkyl;

R⁴ and R⁵ are independently selected from the group

hydrogen,
halo(F, Cl, Br, and I),
15 cyano,
amino,
amido,
nitro,
hydroxy,
20 C₁-C₆alkyl optionally substituted with 1-3 R⁶,
C₂-C₆alkynyl optionally substituted with 1-3 R⁶,
C₁-C₆alkyloxy optionally substituted with 1-3 R⁶,
C₁-C₆acylamino optionally substituted with 1-3 R⁶,
C₁-C₆alkylcarbonyl optionally substituted with 1-3 R⁶,
25 C₁-C₆alkyloxycarbonyl optionally substituted with 1-3 R⁶,
N-(C₁-C₆alkyl),N-(C₁-C₆acyl)amino optionally substituted with 1-
3 R⁶,
N-(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶,
N,N-di(C₀-C₆alkyl)amino optionally substituted with 1-3 R⁶,
30 N,N-di(C₁-C₆alkyl)carboxamido optionally substituted with 1-3
R⁶,
C₁-C₄perfluoroalkyl, and
C₁-C₃perfluoroalkoxy;

R⁶ is selected from the group

35 COOR²,
CONR²R³,
cyano,
NR²R³,

NR^2COR^3 ,

azido,

nitro, and

hydroxy;

5 Q is selected from the group

$-\text{L}^2$,

$-\text{S}(=\text{O})_2-\text{L}^2$,

$-\text{C}(=\text{O})$,

$-\text{C}(=\text{O})-\text{O}$,

10 $-\text{CH}(\text{X})$, and

$-\text{CH}(\text{X})-\text{CH}_2$;

X is selected from the group

hydrogen,

oxo ($=\text{O}$),

15 COOR^2 ,

CONR^2R^3 ,

$\text{C}_0\text{-C}_6\text{alkyl-O-C}_1\text{-C}_6\text{alkyl}$ optionally substituted with 1-2 R^6 , and

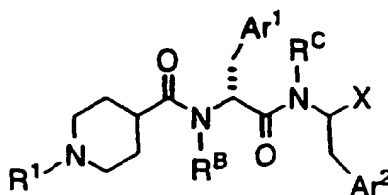
$\text{C}_1\text{-C}_6\text{alkyl}$ optionally substituted with 1-2 R^6 ; and

pharmaceutically acceptable salts thereof.

20

4. The compound of Claim 3 represented by structural Formula (IIa)

25



IIa

where

30

Ar^1 and Ar^2 are each independently selected from

indoyl, and



35

R^B and R^C are selected from the group

hydrogen, and

methyl;

R¹ is selected from

hydrogen,
C₁-C₆alkyl,
C(=O)-C₁-C₆alkyl,
5 C(=O)-NR²R³,
C(=NR²)-NR²R³,
C(=O)O-C₁-C₆alkyl, and
halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ are independently selected from

10 hydrogen,
C₁-C₆alkyl,
piperidinyl, and
halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ together with the nitrogen to which they are attached may form

15 piperidinyl,
pyrrolidinyl,
piperazinyl, and
morpholinyl;

R⁶ is selected from the group

20 COOR²,
CONR²R³,
cyano,
NR²R³,
NR²COR³,
25 azido,
nitro, and
hydroxy;

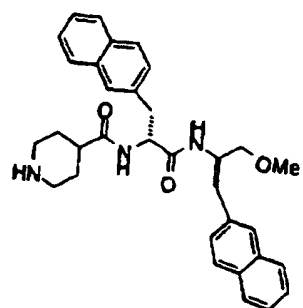
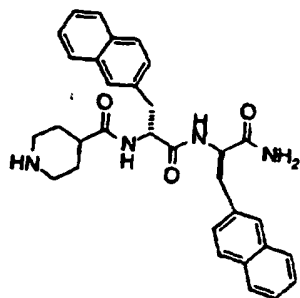
X is selected from the group

hydrogen,
30 oxo (=O),
COOR²,
CONR²R³,
C₀-C₆alkyl-O-C₁-C₆alkyl optionally substituted with 1-2 R⁶, and
C₁-C₆alkyl optionally substituted with 1-2 R⁶; and

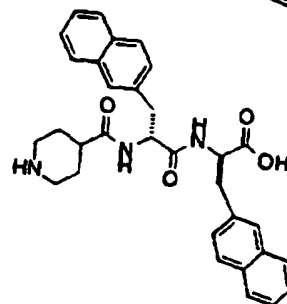
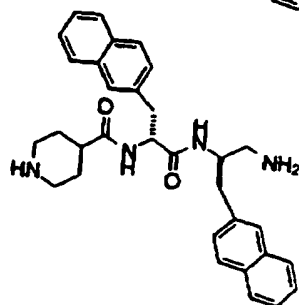
35 pharmaceutically acceptable salts thereof.

5. The compound of Claim 2 selected from the group

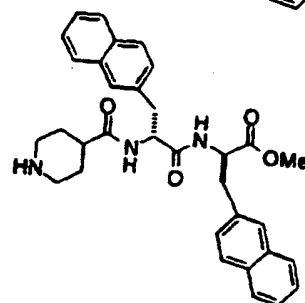
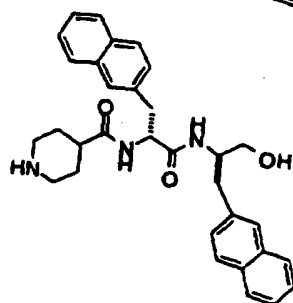
5



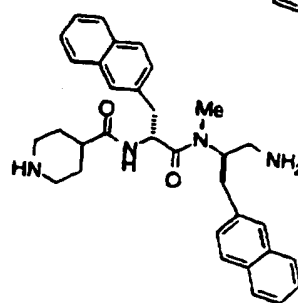
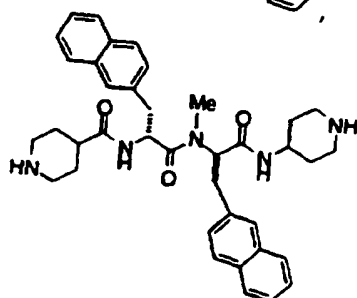
10



15

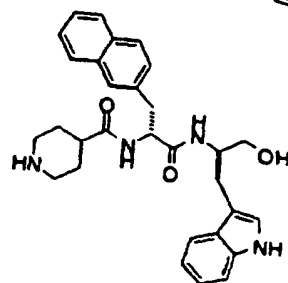
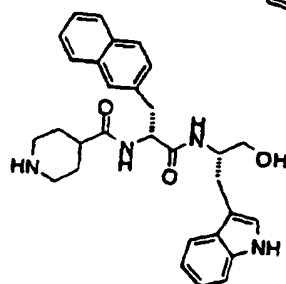


20



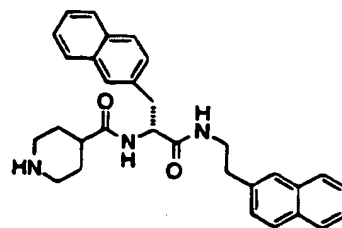
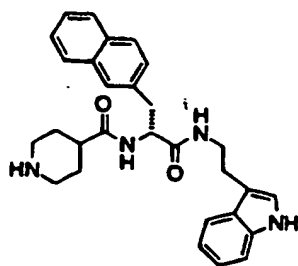
25

30

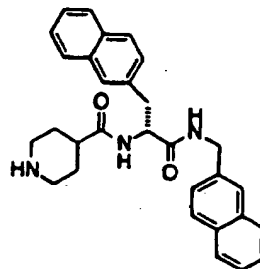
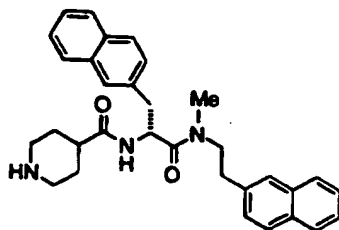


35

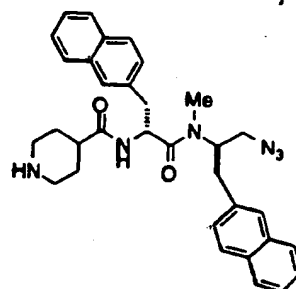
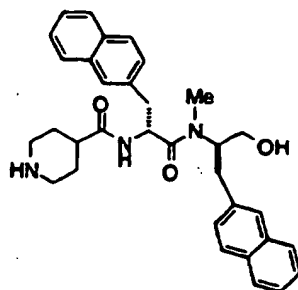
5



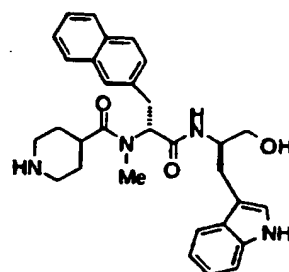
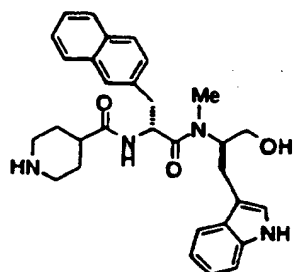
10



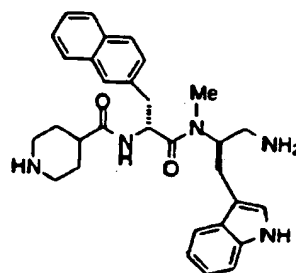
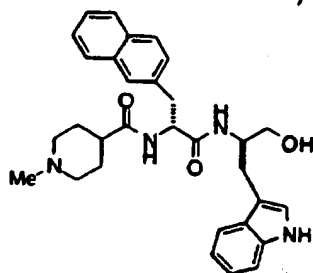
15



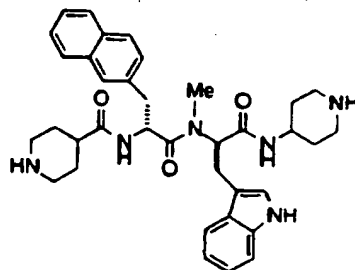
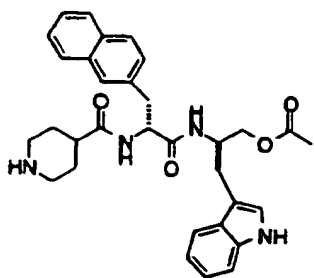
20



25

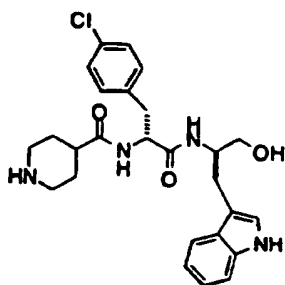


30

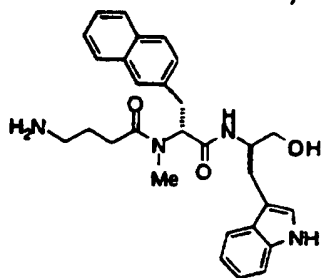


35

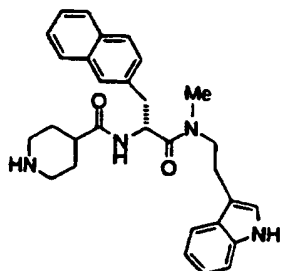
5



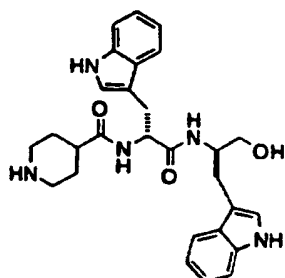
10



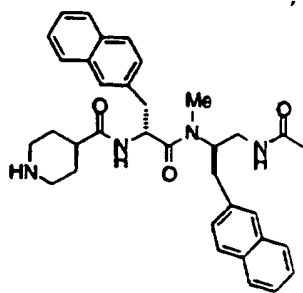
15



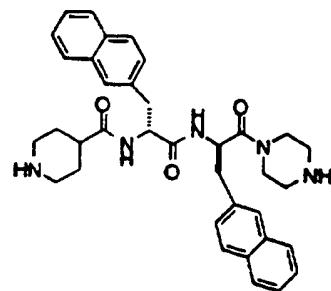
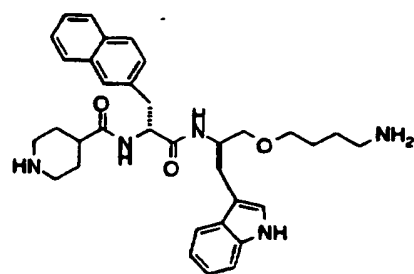
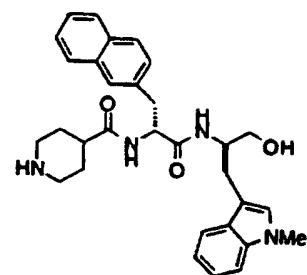
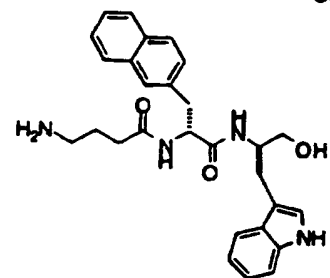
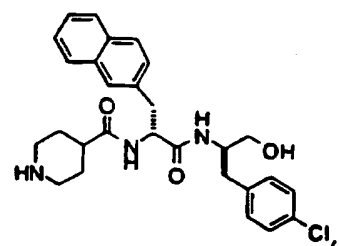
20



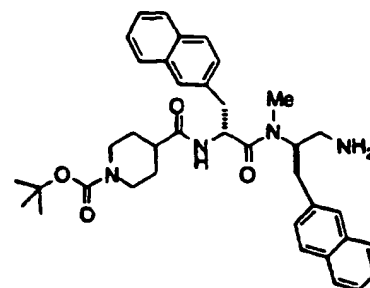
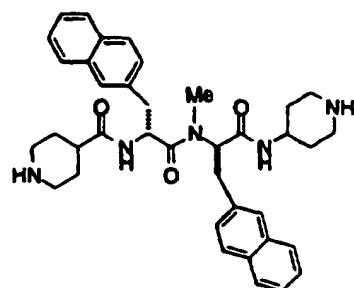
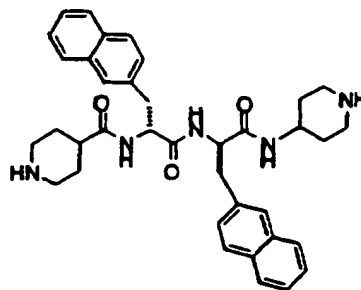
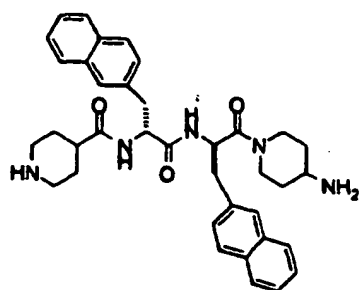
25



30



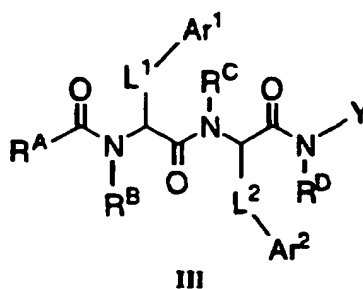
35



15 and

pharmaceutically acceptable salts thereof.

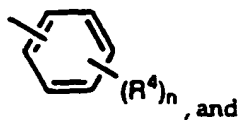
6. The compound of Claim 1 represented by structural Formula (III)



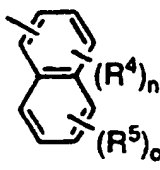
where

Ar¹ and Ar² are each independently selected from

indoyl,



and



n and o are independently 1, 2 or 3;

L^1 and L^2 are independently selected from

-CH₂-O-,
-CH₂-CH₂-O-
-CH₂-,
-CH₂-CH₂-, and
-CH₂-CH₂-CH₂-;

5

R^A is selected from the group

C₀-C₃alkyl-heterocycle where the heterocycle comprises a mono-,
bi-, or tricycle containing 5-12 ring atoms, one or two of
which are heteroatoms selected from O, S, and N, provided
at least one heteroatom is N, where any N atom is optionally
substituted with R¹,

10

C₀-C₆alkyl substituted with one or two substituents selected from
the group
NR²R³, and
imidazoliny!,
pyridiny!,
dihydropyridiny!, and
pipediny!;

15

R^B, R^C and R^D are selected from the group

hydrogen,
C₁-C₆alkyl optionally substituted with a group selected from
NR²R³, and
phenyl-C₁-C₃-NR²R³, and
halo(F, Cl, Br, I)C₁-C₆alkyl;

20

R¹ is selected from

hydrogen,
C₁-C₆alkyl,
C(=O)-C₁-C₆alkyl,
C(=O)-NR²R³,
C(=NR²)-NR²R³,
C(=O)O-C₁-C₆alkyl, and
halo(F, Cl, Br, I)C₁-C₆alkyl;

25

30

R² and R³ are independently selected from

hydrogen,
C₁-C₆alkyl,
piperidiny!, and
halo(F, Cl, Br, I)C₁-C₆alkyl;

35

R² and R³ together with the N to which they are bonded may form a 5- or

6-member heterocycle, optionally containing one additional hetero atom selected from O, S, and N where any N is optionally substituted with R^1 , any carbon is optionally substituted with R^6 and where the heterocycle is optionally fused to a phenyl ring, optionally substituted with R^4 ;

R^4 and R^5 are independently selected from the group

hydrogen,

halo(F, Cl, Br, and I),

cyano,

amino,

amido,

nitro,

hydroxy,

C_1 - C_6 alkyl optionally substituted with 1-3 R^6 ,

C_2 - C_6 alkynyl optionally substituted with 1-3 R^6 ,

C_1 - C_6 alkyloxy optionally substituted with 1-3 R^6 ,

C_1 - C_6 acylamino optionally substituted with 1-3 R^6 ,

C_1 - C_6 alkylcarbonyl optionally substituted with 1-3 R^6 ,

C_1 - C_6 alkyloxycarbonyl optionally substituted with 1-3 R^6 ,

N -(C_1 - C_6 alkyl), N -(C_1 - C_6 acyl)amino optionally substituted with 1-3 R^6 ,

N -(C_1 - C_6 alkyl)carboxamido optionally substituted with 1-3 R^6 ,

N,N -di(C_0 - C_6 alkyl)amino optionally substituted with 1-3 R^6 ,

N,N -di(C_1 - C_6 alkyl)carboxamido optionally substituted with 1-3 R^6 ,

C_1 - C_4 perfluoroalkyl, and

C_1 - C_3 perfluoroalkoxy;

R^6 is selected from the group

$COOR^2$,

$CONR^2R^3$,

cyano,

NR^2R^3 ,

NR^2COR^3 ,

azido,

nitro, and

hydroxy;

R^7 is selected from the group

R^6 , and

C₆-C₁₀aryl optionally substituted with

halo(F, Cl, Br, and I),

cyano,

amino,

amido,

nitro,

hydroxy,

C₁-C₄perfluoroalkyl, and

C₁-C₃perfluoroalkoxy;

Y is selected from the group

C₁-C₆alkyl substituted with 1-2 R⁷,

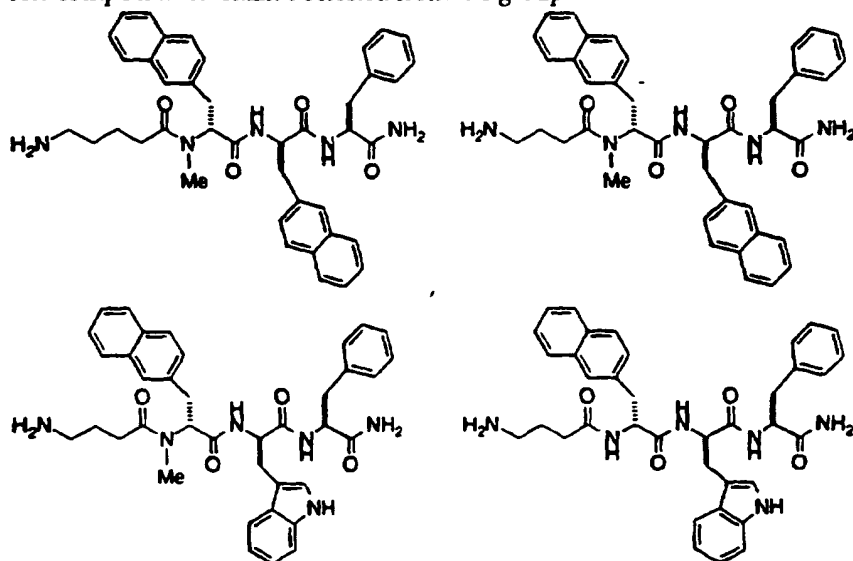
C₂-C₆alkynyl optionally substituted with 1-2 R⁷,

C₂-C₆alkenyl optionally substituted with 1-2 R⁷, and

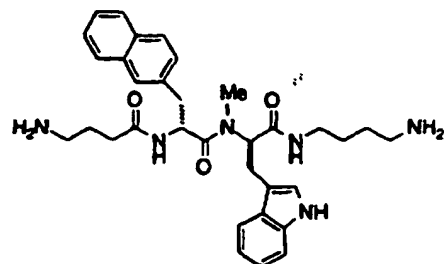
C₁-C₆alkyloxy optionally substituted with 1-2 R⁷, and

pharmaceutically acceptable salts thereof.

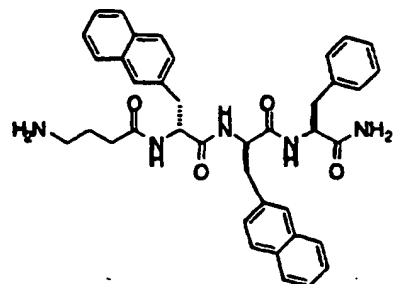
7. The compound of Claim 6 selected from the group



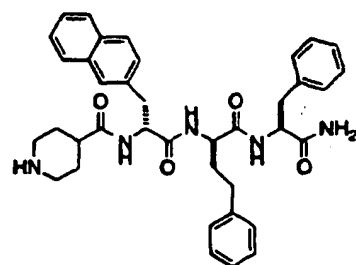
5



10



15

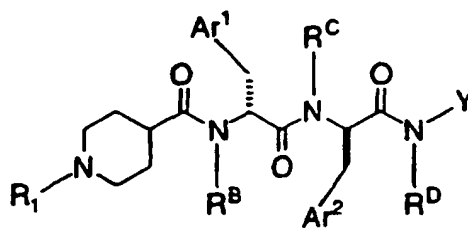


20

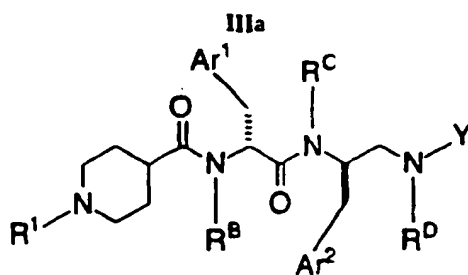
, and
pharmaceutically acceptable salts thereof.

8. The compound of Claim 1 represented by structural Formula IIIa-IIIi

25



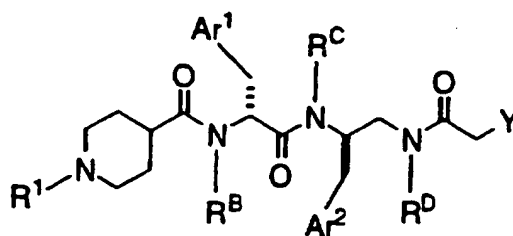
30



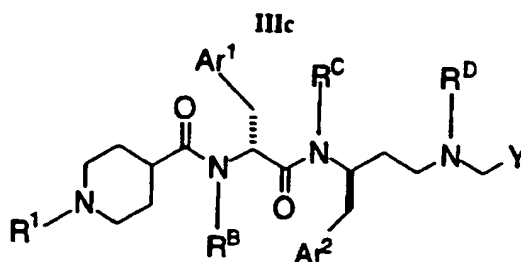
35

IIIb

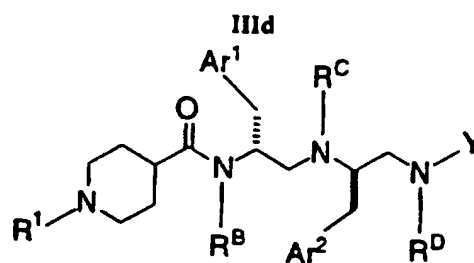
5



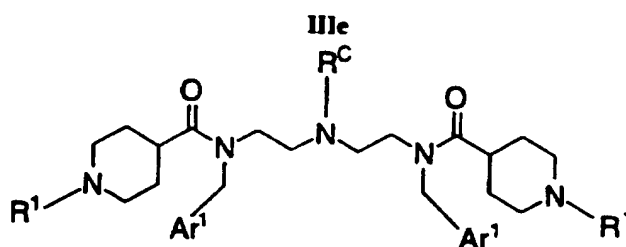
10



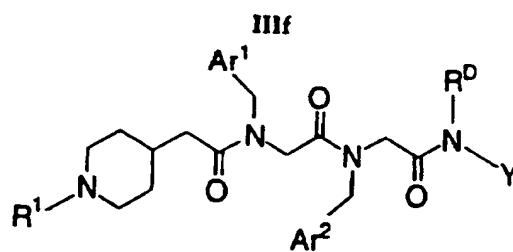
15



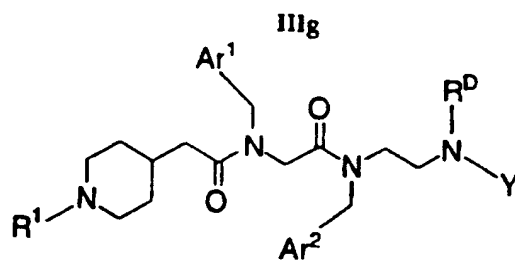
20



25



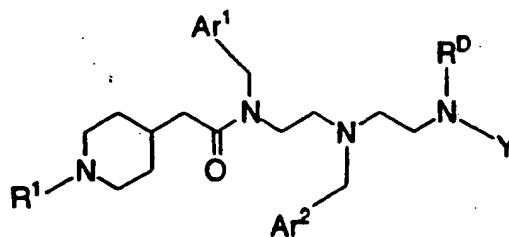
30



35

IIIh

, and

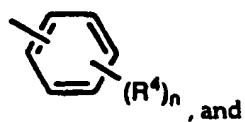


IIIi

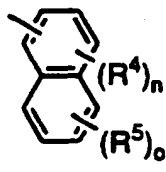
where

Ar^1 and Ar^2 are each independently selected from

indoyl,



and



n and o are independently 1, 2 or 3;

R^B , R^C and R^D are selected from the group

hydrogen,

C_1 - C_6 alkyl optionally substituted with a group selected from

NR^2R^3 , and

phenyl- C_1 - C_3 - NR^2R^3 , and

halo(F, Cl, Br, I) C_1 - C_6 alkyl;

R^1 is selected from

hydrogen,

C_1 - C_6 alkyl,

$C(=O)$ - C_1 - C_6 alkyl,

$C(=O)$ - NR^2R^3 ,

$C(=NR^2)$ - NR^2R^3 ,

$C(=O)O$ - C_1 - C_6 alkyl, and

halo(F, Cl, Br, I) C_1 - C_6 alkyl;

R^2 and R^3 are selected from

hydrogen,

C_1 - C_6 alkyl,

piperidiny, and

halo(F, Cl, Br, I) C_1 - C_6 alkyl;

R^2 and R^3 together with the N to which they are bonded may form a 5- or

6-member heterocycle, optionally containing one additional hetero
atom selected from O, S, and N where any N is optionally
substituted with R^1 , any carbon is optionally substituted with R^7
and where the heterocycle is optionally fused to a phenyl ring,
optionally substituted with R^4 ;

5

R^4 and R^5 are independently selected from the group

hydrogen,

halo(F, Cl, Br, and I),

cyano,

10

amino,

amido,

nitro,

hydroxy,

C_1 - C_4 perfluoroalkyl, and

15

C_1 - C_3 perfluoroalkoxy;

R^7 is selected from the group

$COOR^2$,

$CONR^2R^3$,

cyano,

20

NR^2R^3 ,

NR^2COR^3 ,

azido,

nitro,

hydroxy,

25

C_6 - C_{10} aryl optionally substituted with

halo(F, Cl, Br, and I),

cyano,

amino,

amido,

30

nitro,

hydroxy,

C_1 - C_4 perfluoroalkyl, and

C_1 - C_3 perfluoroalkoxy;

Y is selected from the group

35

C_1 - C_6 alkyl substituted with 1-2 R^7 ,

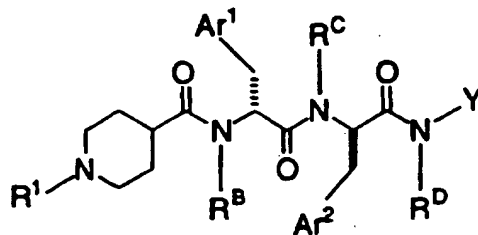
C_2 - C_6 alkynyl optionally substituted with 1-2 R^7 ,

C_2 - C_6 alkylenyl optionally substituted with 1-2 R^7 ,

C_1 - C_6 alkyloxy optionally substituted with 1-2 R^7 , and

piperidinyl; and
pharmaceutically acceptable salts thereof.

9. The compound of Claim 8 represented by structural Formula (IIIa)



IIIa

where

Ar¹ and Ar² are each independently selected from
indoyl, and



R^B, R^C and R^D are selected from the group
hydrogen,
C₁-C₆alkyl,
C₆-C₁₀aryl-C₁-C₆alkyl, and
halo(F, Cl, Br, I)C₁-C₆alkyl;

R¹ is selected from
hydrogen,
C₁-C₆alkyl,
C(=O)OC₁-C₆alkyl, and
halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ are selected from
hydrogen,
C₁-C₆alkyl,
piperidinyl, and
halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ together with the nitrogen to which they are attached may form
an optionally mono- or di-substituted ring selected from
piperidinyl,
pyrrolylidinyl,
pyrryl,

imidazolyl,
piperazinyl, and
morpholinyl

where the substituents are selected from C₁-C₃alkyl;

5 R⁷ is selected from the group

COOR²,
CONR²R³,
cyano,
NR²R³,
10 NR²COR³,
azido,
nitro,
hydroxy, and
C₆-C₁₀aryl optionally substituted with
15 halo(F, Cl, Br, and I),
cyano,
amino,
amido,
nitro,
20 hydroxy,
C₁-C₄perfluoroalkyl, and
C₁-C₃perfluoroalkoxy;

Y is selected from the group

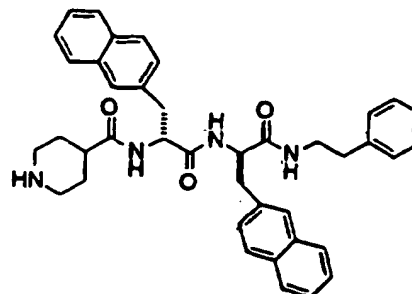
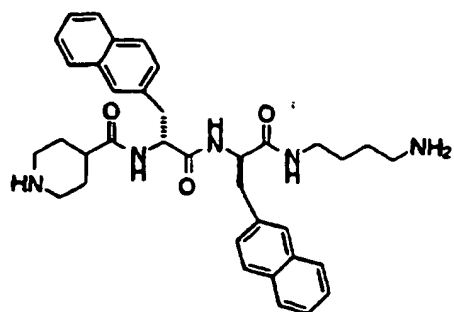
C₁-C₆alkyl substituted with 1-2 R⁷,
25 C₂-C₆alkynyl optionally substituted with 1-2 R⁷,
C₂-C₆alkenyl optionally substituted with 1-2 R⁷,
C₁-C₆alkyloxy optionally substituted with 1-2 R⁷, and
piperidinyl;

30 Y and R^D together with the N to which they are bonded may form a 5- or
6-member heterocycle, optionally containing one additional hetero
atom selected from O, S, and N where any N is optionally
substituted with R¹, any carbon is optionally substituted with R⁷
and where the heterocycle is optionally fused to a phenyl ring; and
pharmaceutically acceptable salts thereof.

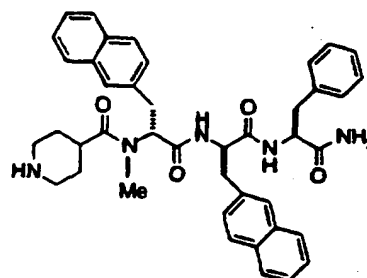
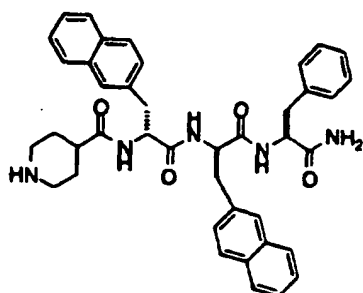
35

10. The compound of Claim 9 selected from the group

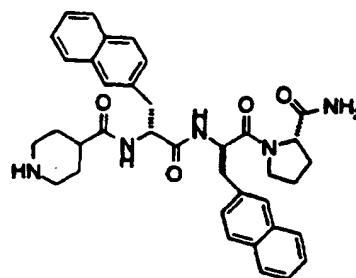
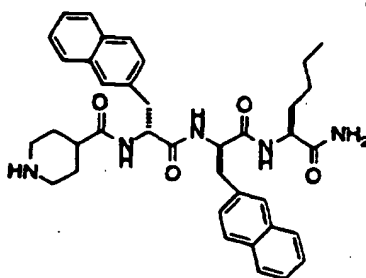
5



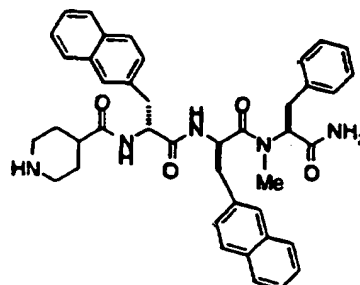
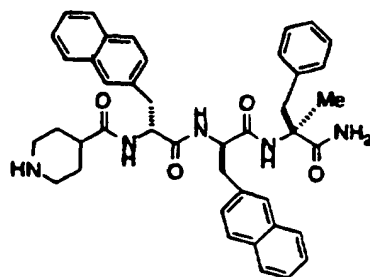
10



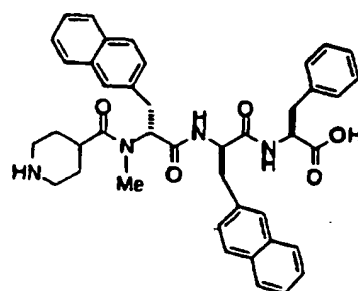
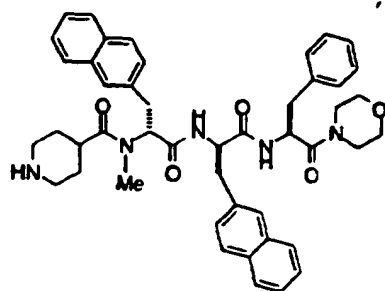
15



20



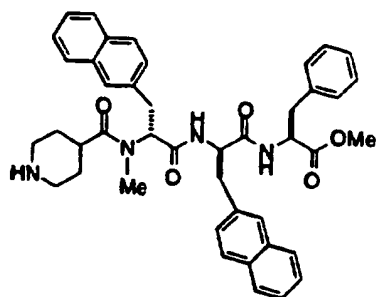
25



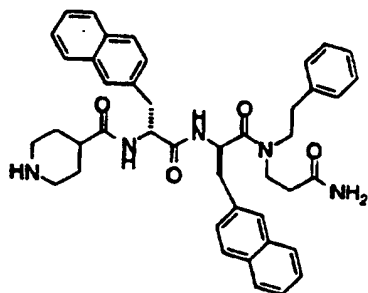
30

35

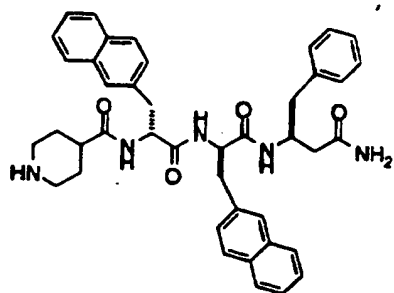
5



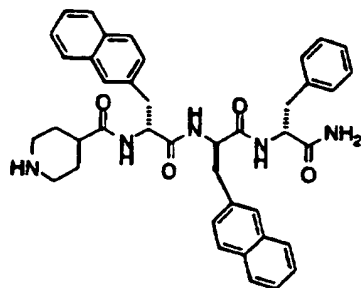
10



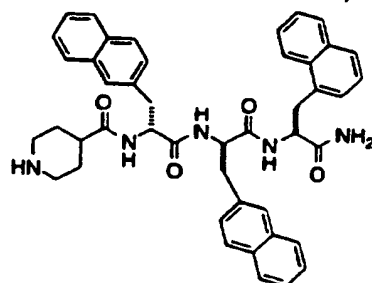
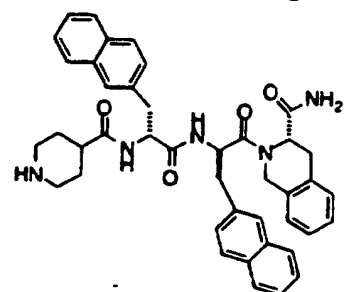
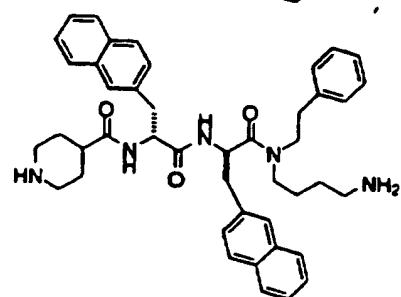
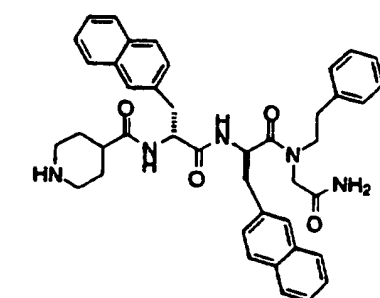
15



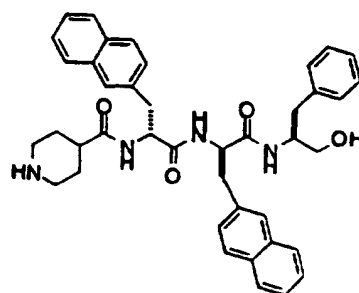
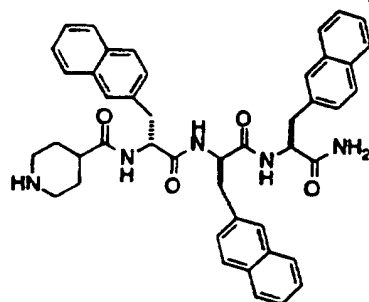
20



25

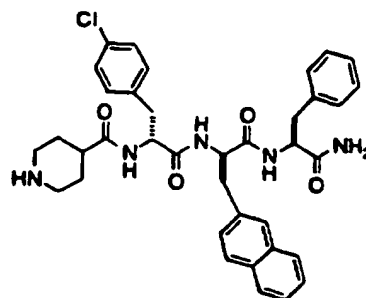
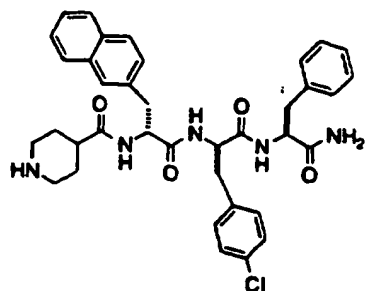


30

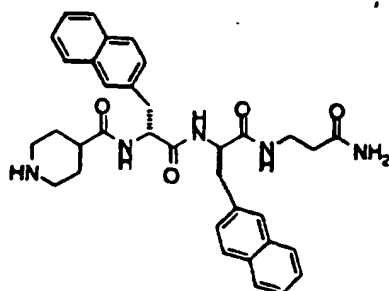
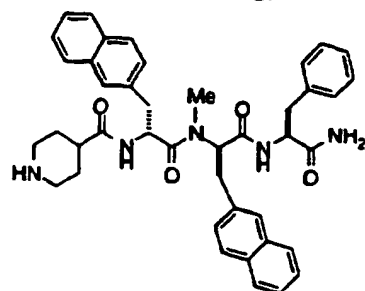


35

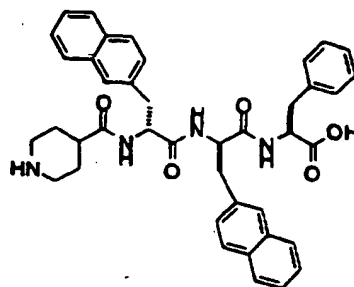
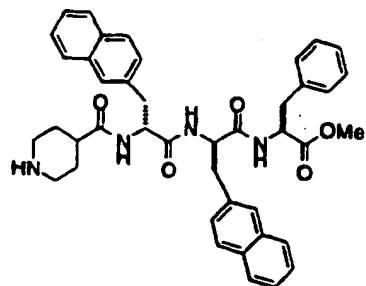
5



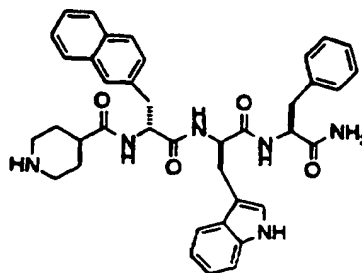
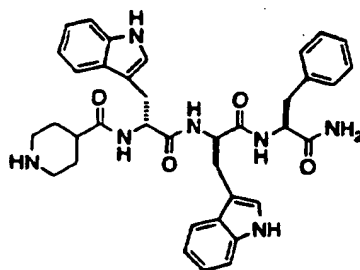
10



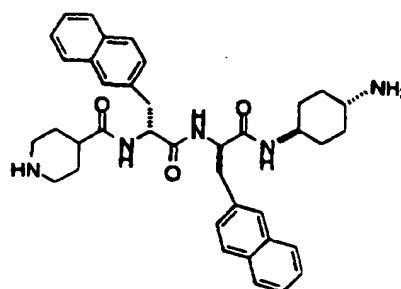
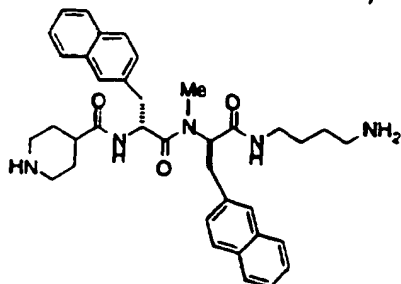
15



20



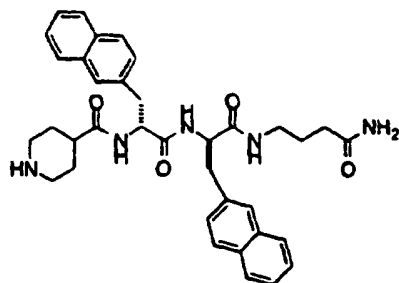
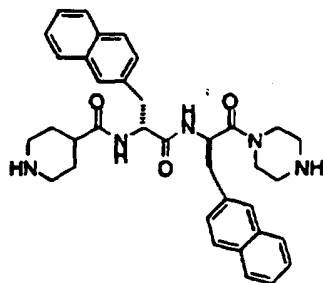
25



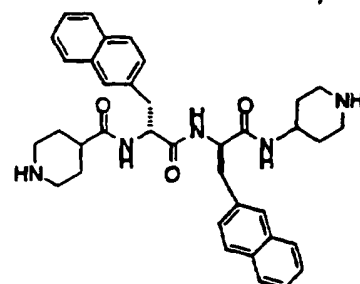
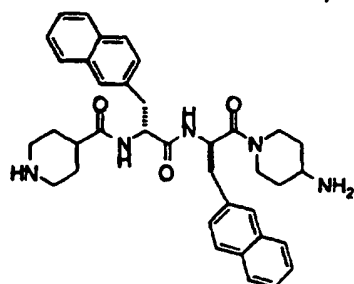
30

35

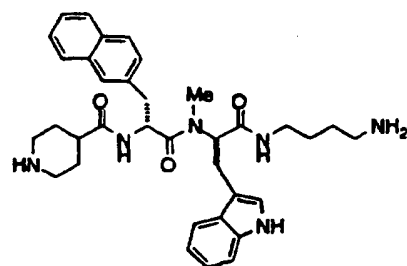
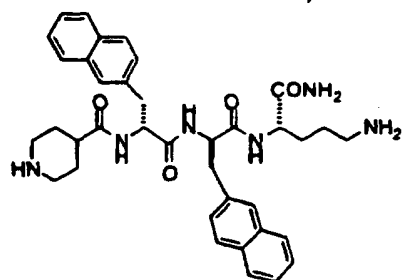
5



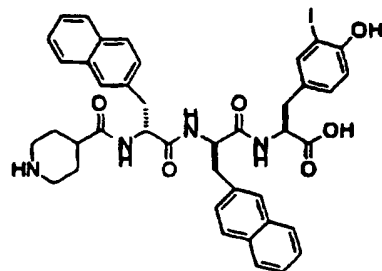
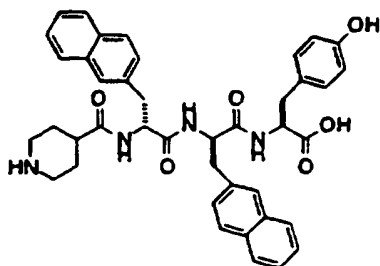
10



15



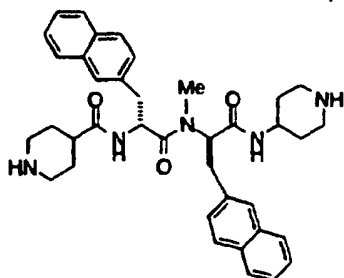
20



25

, and

30

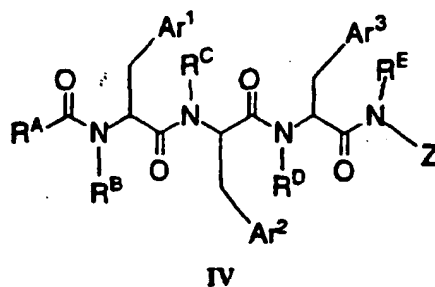


; and

pharmaceutically acceptable salts thereof.

35

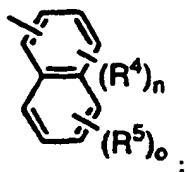
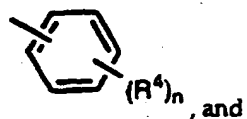
11. The compound of Claim 1 represented by structural Formula (IV)



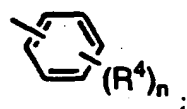
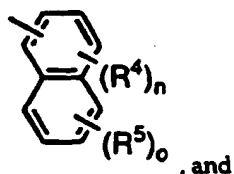
where

Ar¹ and Ar² are each independently selected from

10 indoyl,



Ar³ is selected from the group



n and o are independently 1, 2 or 3;

R^A is selected from the group

C₀-C₃alkyl-heterocycle where the heterocycle comprises a mono-,

bi-, or tricycle containing 5-12 ring atoms, one or two of

which are heteroatoms selected from O, S, and N, provided

at least one heteroatom is N, where any N atom is

optionally substituted with R¹,

C₀-C₆alkyl substituted with one or two substituents selected from

the group

NR²R³,

imidazolynyl,

pyridinyl,

dihydropyridinyl, and

piperidinyl;

R^B , R^C , R^D , and R^E are selected from the group

hydrogen,

5 C_1 - C_6 alkyl optionally substituted with a group selected from

NR^2R^3 , and

phenyl- C_1 - C_3 - NR^2R^3 , and

halo(F, Cl, Br, I) C_1 - C_6 alkyl;

R^1 is selected from

10 hydrogen,

C_1 - C_6 alkyl,

$C(=O)$ - C_1 - C_6 alkyl,

$C(=O)$ - NR^2R^3 ,

$C(=NR^2)$ - NR^2R^3 ,

15 $C(=O)O$ - C_1 - C_6 alkyl, and

halo(F, Cl, Br, I) C_1 - C_6 alkyl;

R^2 and R^3 are independently selected from

hydrogen,

C_1 - C_6 alkyl,

20 piperidinyl, and

halo(F, Cl, Br, I) C_1 - C_6 alkyl;

R^2 and R^3 together with the N to which they are bonded may form a 5- or

6-member heterocycle, optionally containing one additional hetero

atom selected from O, S, and N where any N is optionally

25 substituted with R^1 , any carbon is optionally substituted with R^6

and where the heterocycle is optionally fused to a phenyl ring,

optionally substituted with R^4 ;

R^4 and R^5 are independently selected from the group

hydrogen,

30 halo(F, Cl, Br, and I),

cyano,

amino,

amido,

nitro,

35 hydroxy,

C_1 - C_6 alkyl optionally substituted with 1-3 R^6 ,

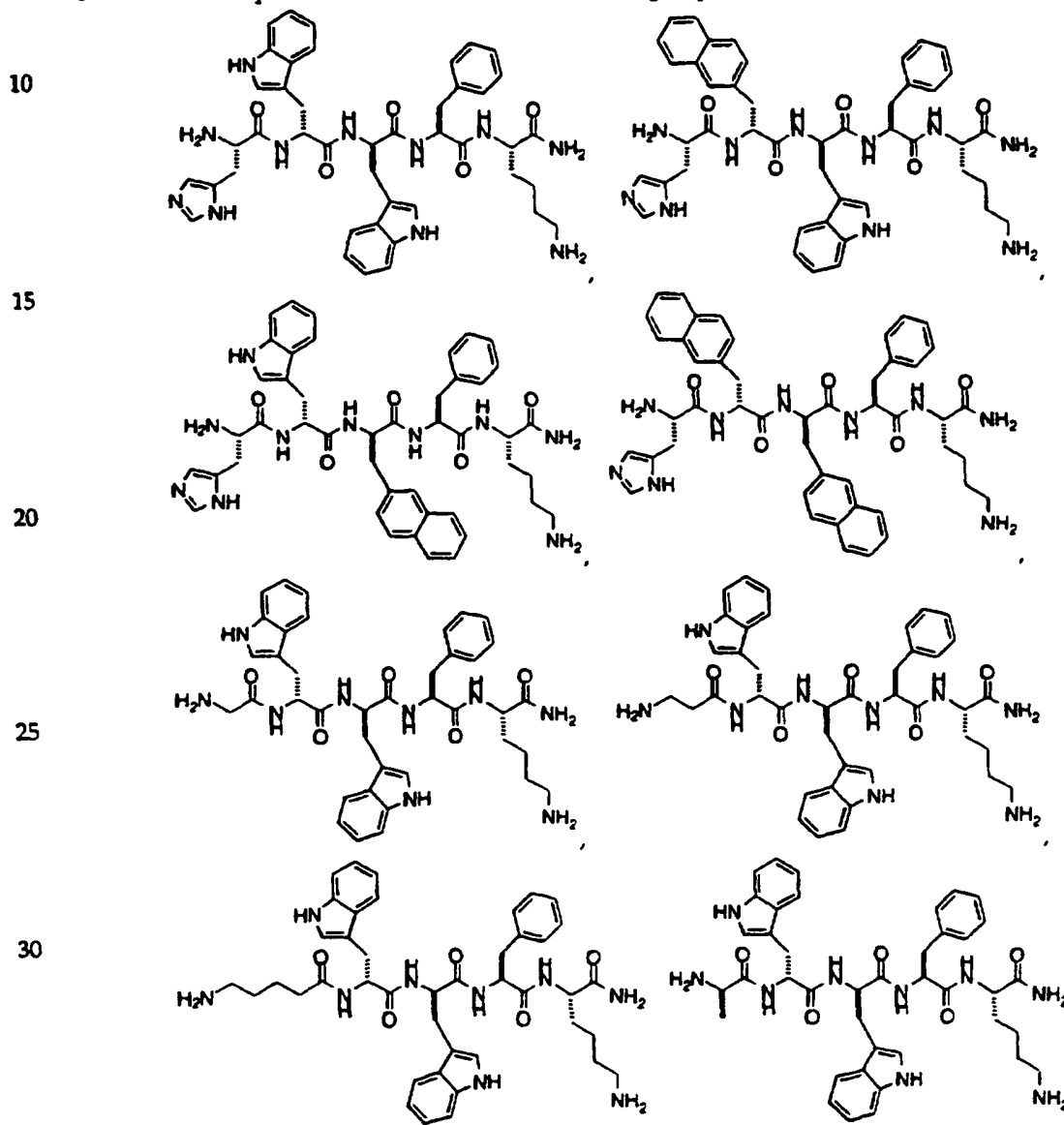
C_2 - C_6 alkynyl optionally substituted with 1-3 R^6 ,

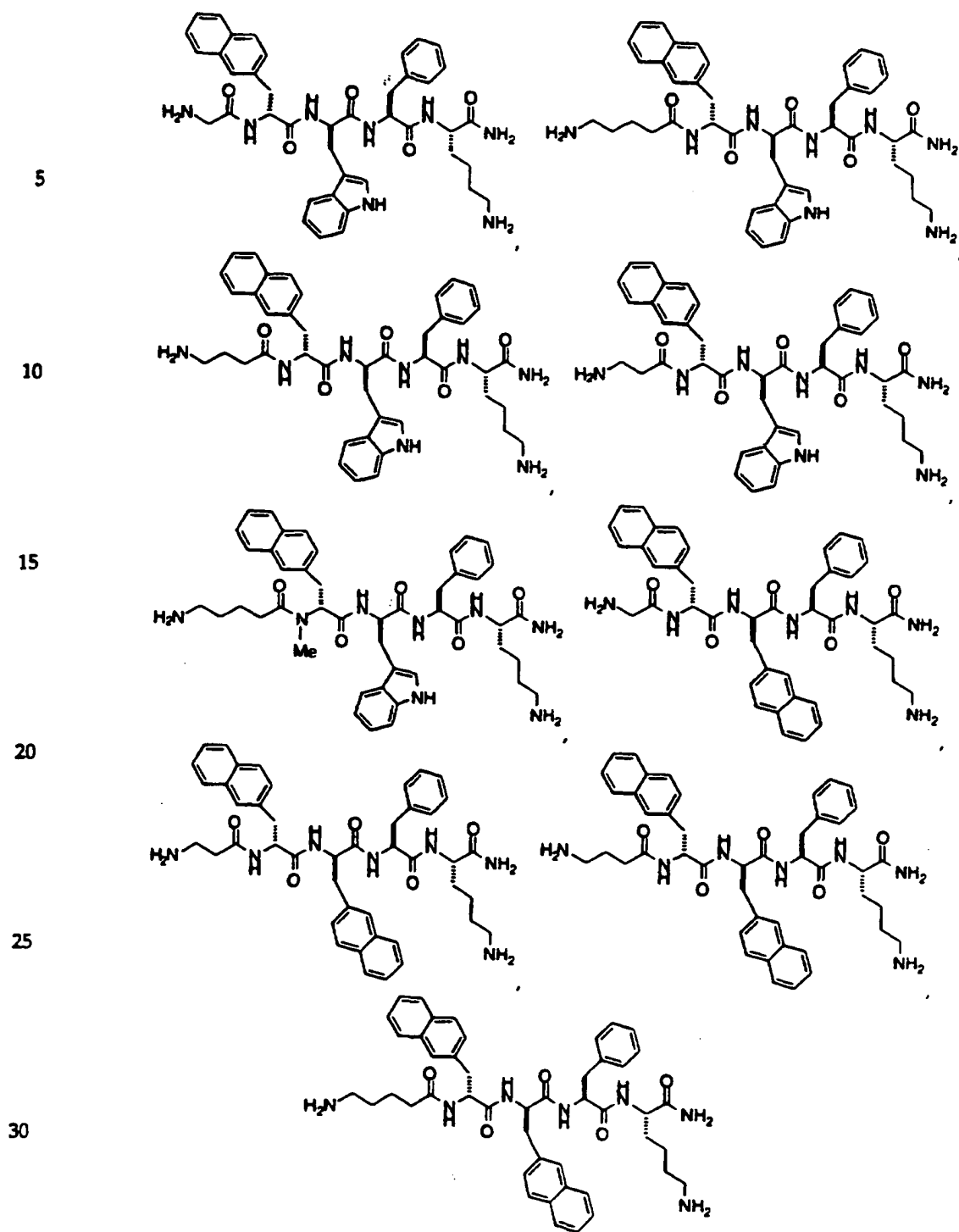
C_1 - C_6 alkyloxy optionally substituted with 1-3 R^6 ,

- C₁-C₆acylamino optionally substituted with 1-3 R⁶,
 C₁-C₆alkylcarbonyl optionally substituted with 1-3 R⁶,
 C₁-C₆alkyloxycarbonyl optionally substituted with 1-3 R⁶,
 N-(C₁-C₆alkyl),N-(C₁-C₆acyl)amino optionally substituted with 1-
 3 R⁶,
 N-(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶,
 N,N-di(C₀-C₆alkyl)amino optionally substituted with 1-3 R⁶,
 N,N-di(C₁-C₆alkyl)carboxamido optionally substituted with 1-3
 R⁶,
 C₁-C₄perfluoroalkyl, and
 C₁-C₃perfluoroalkoxy;
 R⁶ is selected from the group
 COOR²,
 CONR²R³,
 O(C=O)R²,
 cyano,
 NR²R³,
 NR²COR³,
 azido,
 nitro, and
 hydroxy;
 R⁷ is selected from the group
 R⁶, and
 C₆-C₁₀aryl optionally substituted with
 halo(F, Cl, Br, and I),
 cyano,
 amino,
 amido,
 nitro,
 hydroxy,
 C₁-C₄perfluoroalkyl, and
 C₁-C₃perfluoroalkoxy;
 Z is selected from the group
 C₁-C₆alkyl substituted with 1-2 R⁷,
 C₂-C₆alkynyl optionally substituted with 1-2 R⁷,
 C₂-C₆alkenyl optionally substituted with 1-2 R⁷, and
 C₁-C₆alkyloxy optionally substituted with 1-2 R⁷,

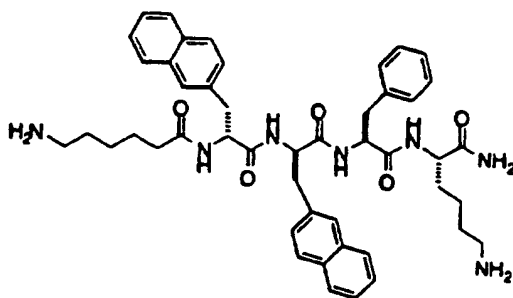
Z and R^E together with the N to which they are bonded may form a 5- or 6-member heterocycle, optionally containing one additional hetero atom selected from O, S, and N where any N is optionally substituted with R¹, any carbon is optionally substituted with R⁷ and where the heterocycle is optionally fused to a phenyl ring; and pharmaceutically acceptable salts thereof.

12. The compound of Claim 11 selected from the group

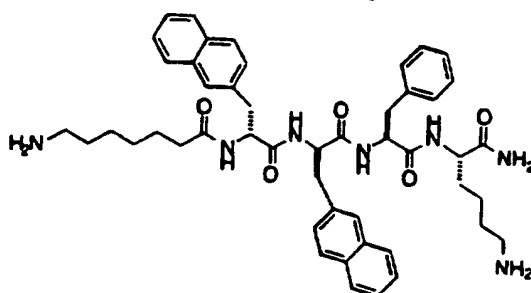




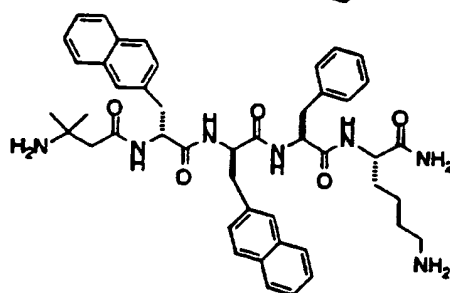
5



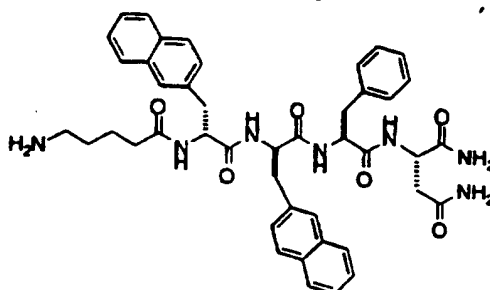
10



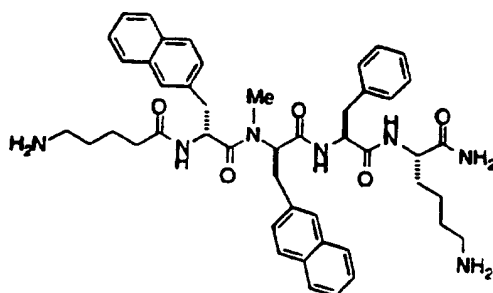
15



20



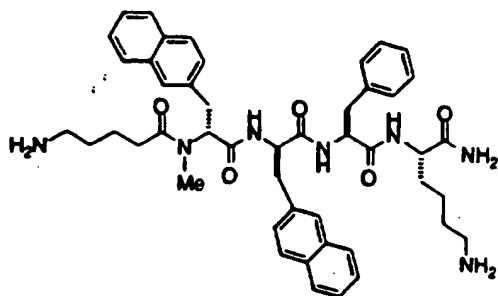
25



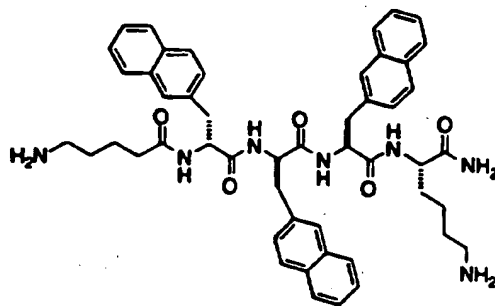
30

35

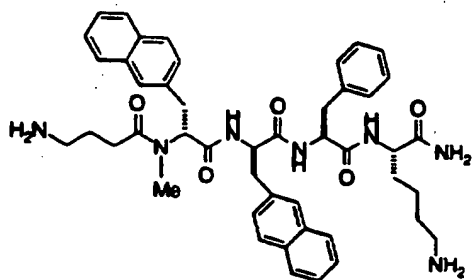
5



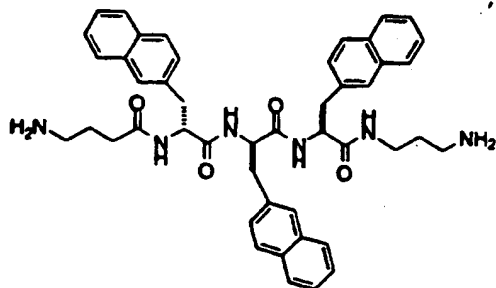
10



15

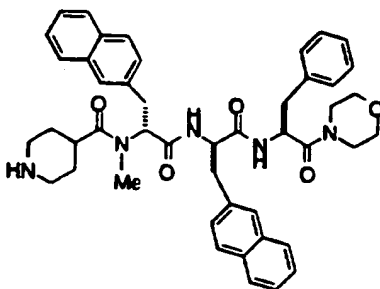


20

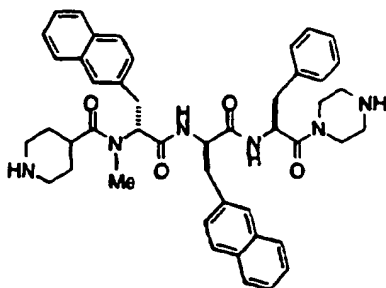


25

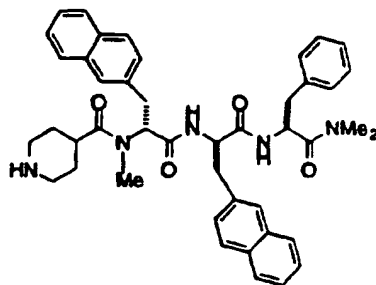
30



35



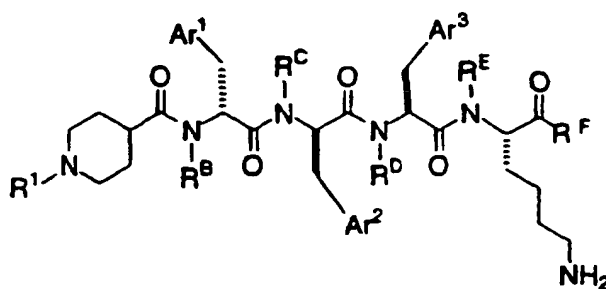
, and



; and

pharmaceutically acceptable salts thereof.

13. The compound of Claim 11 represented by structural Formula (IVa)



IVa

where

 R^B , R^C , R^D and R^E are selected from the group

hydrogen, and

 C_1 - C_6 alkyl; Ar^1 and Ar^2 are each independently selected from

indoyl, and

 Ar^3 is selected from the group



, and



R^F is selected from the group

OH,

C₁-C₄alkyloxy,NR⁵R⁶, and1 to 4 α -amino acid residues;

R^4 is selected from

hydrogen,

halo(F, Cl, Br, and I),

cyano,

amino,

amido,

nitro,

hydroxy,

C₁-C₄alkoxy,C₁-C₄perfluoroalkyl, andC₁-C₃perfluoroalkoxy;

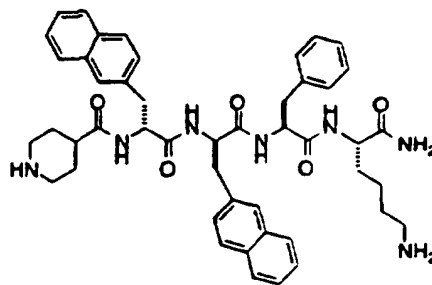
R^5 and R^6 are independently selected from

hydrogen, and

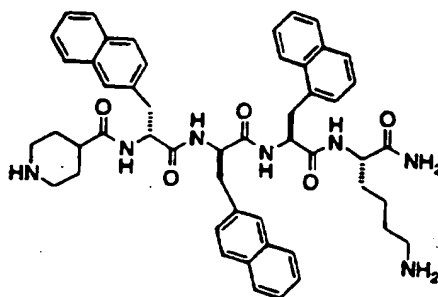
C₁-C₆alkyl; and

pharmaceutically acceptable salts thereof.

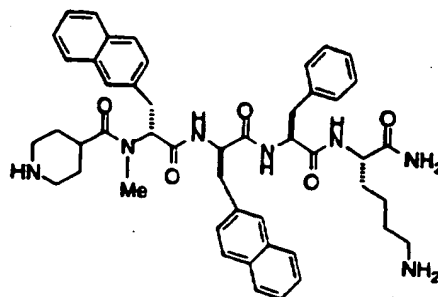
14. The compound of Claim 13 selected from the group



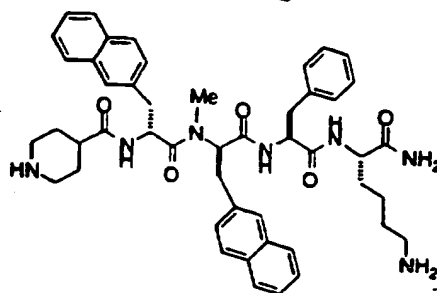
5



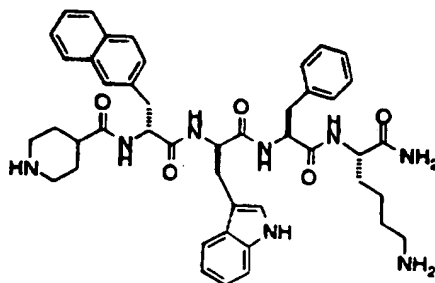
10



15

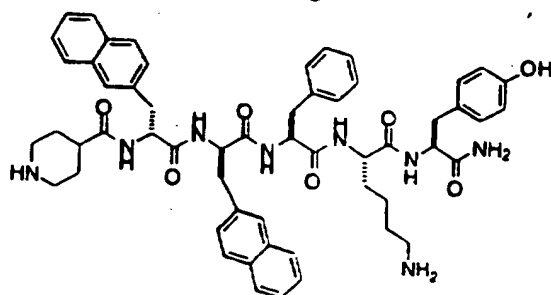


20



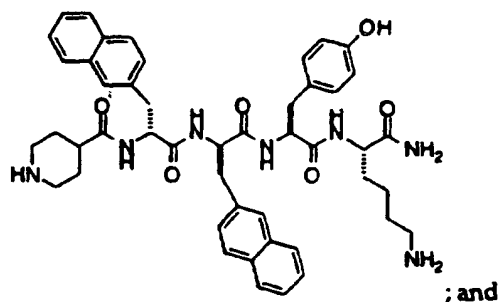
25

30



, and

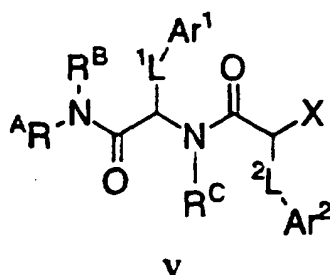
35



pharmaceutically acceptable salts thereof.

10

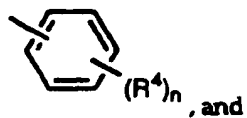
15. The compound of Claim 1 represented by Formula V



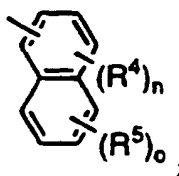
where

Ar¹ and Ar² are each independently selected from

indoyl,



, and



n and o are independently 1, 2 or 3;

L¹ is selected from

-CH₂-O-,

-CH₂-CH₂-O-

-CH₂-,

-CH₂-CH₂-, and

-CH₂-CH₂-CH₂-;

L² is selected from

a covalent bond,

-O-,

-O-CH₂-, and

L¹;

R^A is selected from the group

C₀-C₃alkyl-heterocycle where the heterocycle comprises a mono-,

5

bi-, or tricycle containing 5-12 ring atoms, one or two of

which are heteroatoms selected from O, S, and N, provided

at least one heteroatom is N, where any N atom is

optionally substituted with R¹,

C₀-C₆alkyl substituted with one or two substituents selected from

10

the group

NR²R³, and

imidazoliny,

pyridiny,

dihydropyridiny, and

15

pipediny;

R^B and R^C are selected from the group

hydrogen,

C₁-C₆alkyl optionally substituted with a group selected from

NR²R³, and

20

phenyl-C₁-C₃-NR²R³, and

halo(F, Cl, Br, I)C₁-C₆alkyl;

R^A and R^B together with the N to which they are bonded may form a 5- or

6-member heterocycle, optionally containing one additional hetero

atom selected from O, S, and N where any N is optionally

25

substituted with R¹, any carbon is optionally substituted with R⁶

and where the heterocycle is optionally fused to a phenyl ring,

optionally substituted with R⁴;

R¹ is selected from

hydrogen,

30

C₁-C₆alkyl,

C(=O)-C₁-C₆alkyl,

C(=O)-NR²R³,

C(=NR²)-NR²R³,

C(=O)O-C₁-C₆alkyl, and

35

halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ are independently selected from

R¹,

hydrogen,

C₁-C₆alkyl,
 piperidiny, and
 halo(F, Cl, Br, I)C₁-C₆alkyl;

R² and R³ together with the N to which they are bonded may form a 5- or
 6-member heterocycle, optionally containing one additional hetero
 atom selected from O, S, and N where any N is optionally
 substituted with R¹, any carbon is optionally substituted with R⁶
 and where the heterocycle is optionally fused to a phenyl ring,
 optionally substituted with R⁴;

R⁴ and R⁵ are independently selected from the group

hydrogen,
 halo(F, Cl, Br, and I),
 cyano,
 amino,
 amido,
 nitro,
 hydroxy,
 C₁-C₆alkyl optionally substituted with 1-3 R⁶,
 C₂-C₆alkynyl optionally substituted with 1-3 R⁶,
 C₁-C₆alkyloxy optionally substituted with 1-3 R⁶,
 C₁-C₆acylamino optionally substituted with 1-3 R⁶,
 C₁-C₆alkylcarbonyl optionally substituted with 1-3 R⁶,
 C₁-C₆alkyloxycarbonyl optionally substituted with 1-3 R⁶,
 N-(C₁-C₆alkyl),N-(C₁-C₆acyl)amino optionally substituted with 1-
 3 R⁶,
 N-(C₁-C₆alkyl)carboxamido optionally substituted with 1-3 R⁶,
 N,N-di(C₀-C₆alkyl)amino optionally substituted with 1-3 R⁶,
 N,N-di(C₁-C₆alkyl)carboxamido optionally substituted with 1-3
 R⁶,
 C₁-C₄perfluoroalkyl, and
 C₁-C₃perfluoroalkoxy;

R⁶ is selected from the group

COOR²,
 CONR²R³,
 cyano,
 NR²R³,
 NR²COR³,
 azido,

nitro, and

hydroxy;

X is selected from the group

hydrogen,

COOR²,

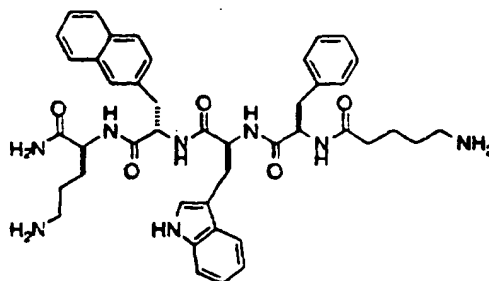
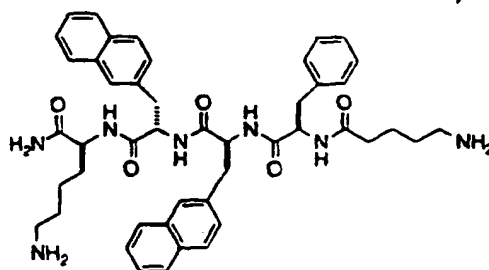
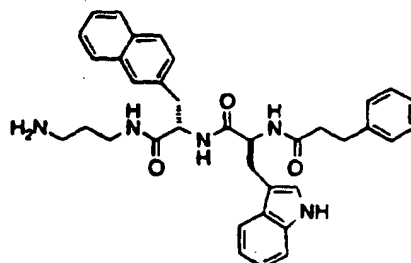
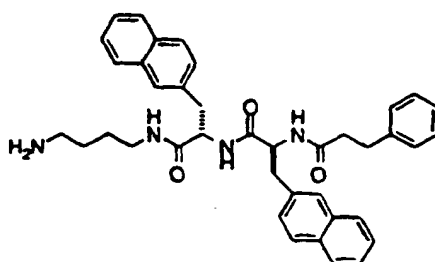
C₀-C₆alkyl-NR²R³,

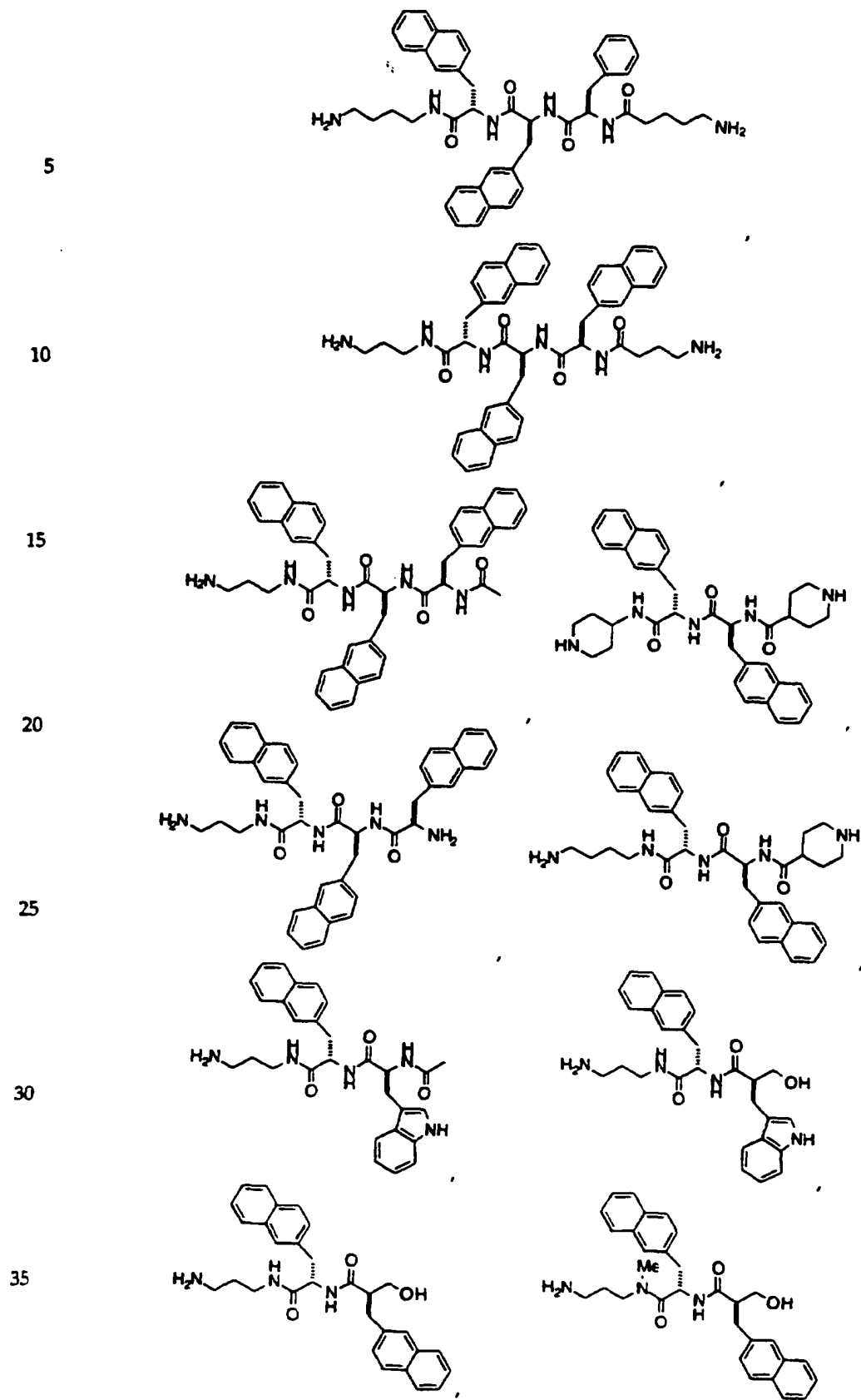
C₀-C₆alkyl-O-C₁-C₆alkyl optionally substituted with 1-2 R⁶, and

C₁-C₆alkyl optionally substituted with 1-2 R⁶; and

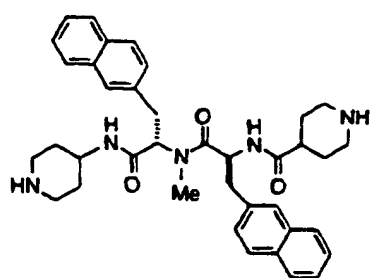
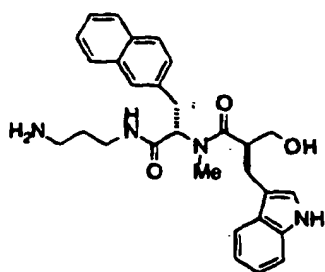
pharmaceutically acceptable salts thereof.

16. The compound of Claim 15 selected from the group

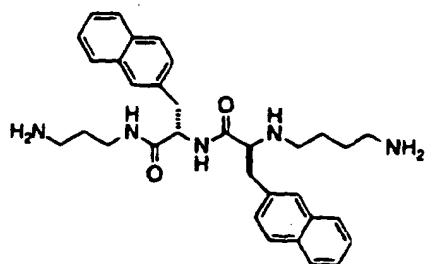
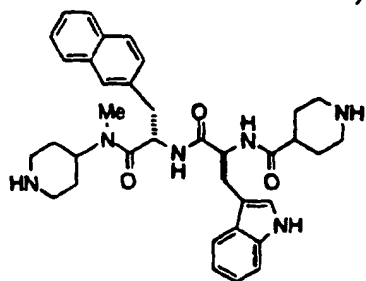




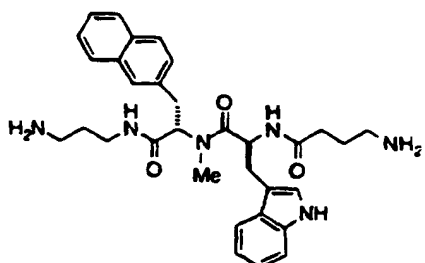
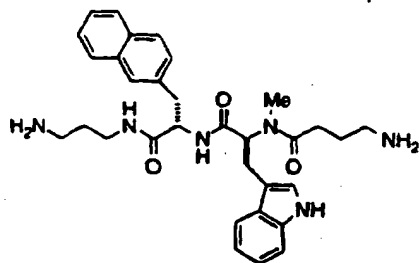
5



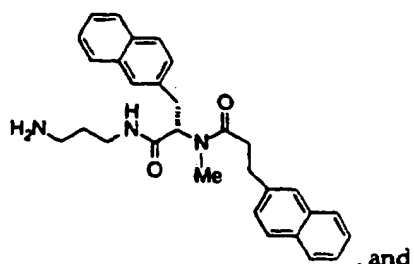
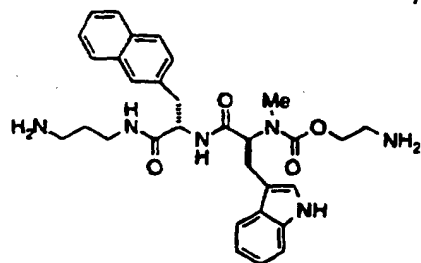
10



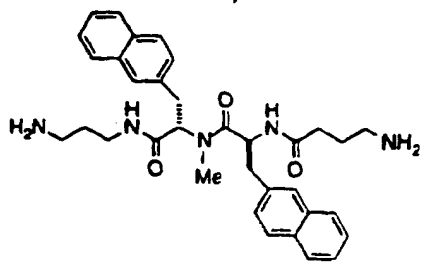
15



20



25



30

, and

pharmaceutically acceptable salts thereof.

17. A pharmaceutical composition comprising a pharmaceutically acceptable excipient and the compound of Claim 1.

35

18. A method for increasing the level of endogenous growth hormone in a mammal comprising administering to the mammal a pharmaceutically effective amount of the composition of Claim 17 to the mammal.
- 5 19. The method of Claim 18 further comprising administering the composition in combination with a growth factor selected from the group; growth hormone (GH), growth hormone releasing hormone (GHRH), insulin like growth factor-1 (IGF-1), and insulin like growth factor-2 (IGF-2).
- 10 20. A method for treating Type II diabetes in a mammal in need of such treatment comprising administering to the mammal a pharmaceutically effective amount of the composition of Claim 17 to the mammal.

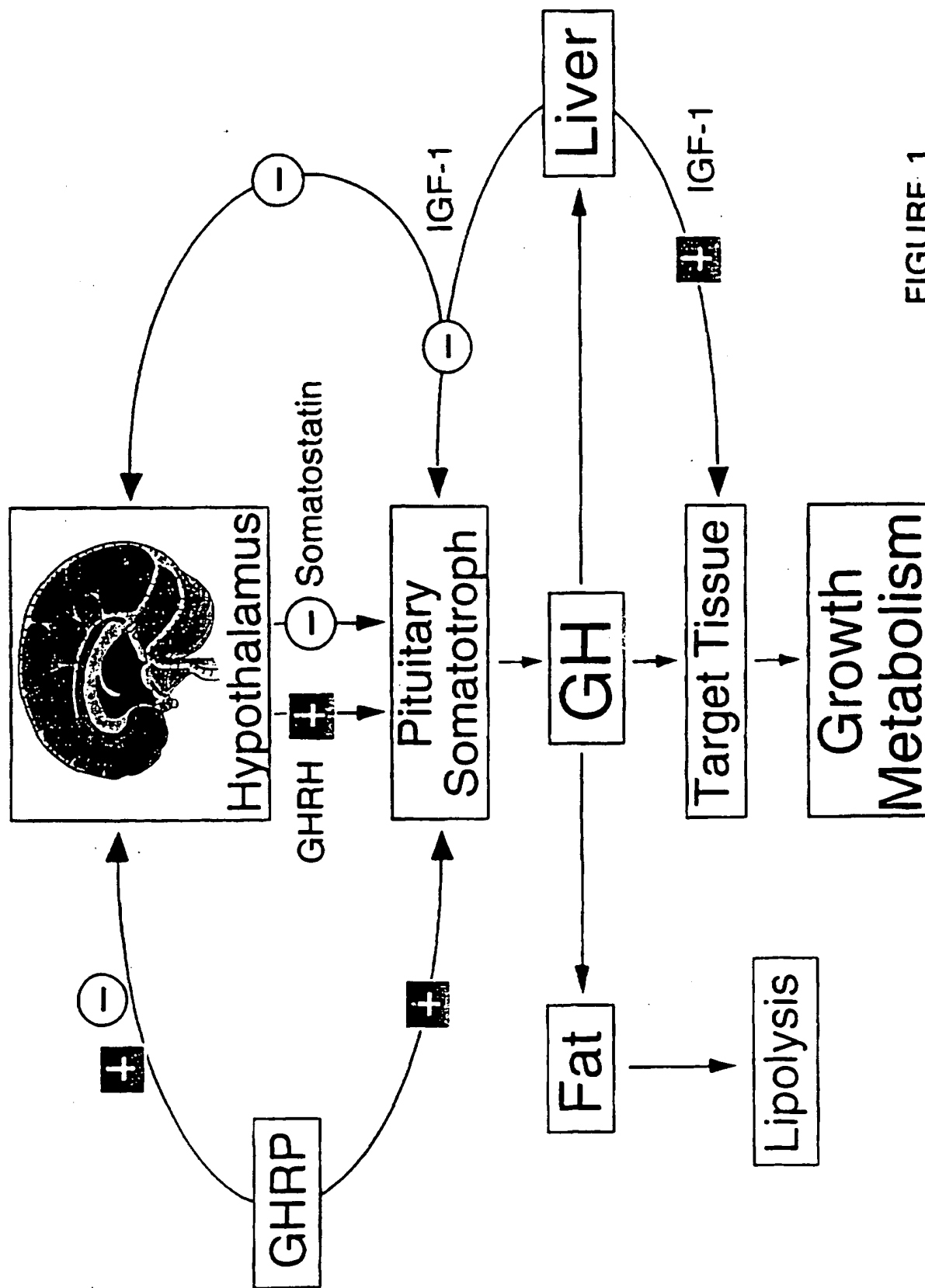
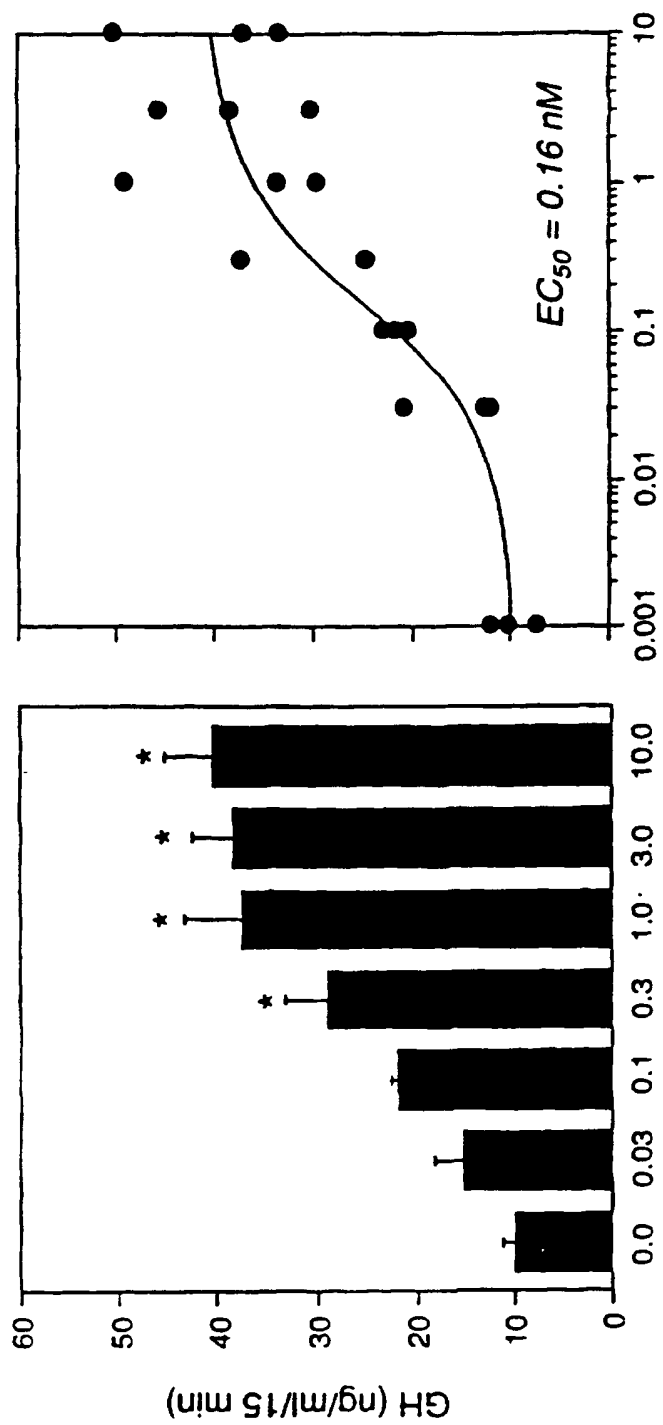


FIGURE 1



Concentration (nM)
(inip) bbFK-NH₂

FIGURE 2

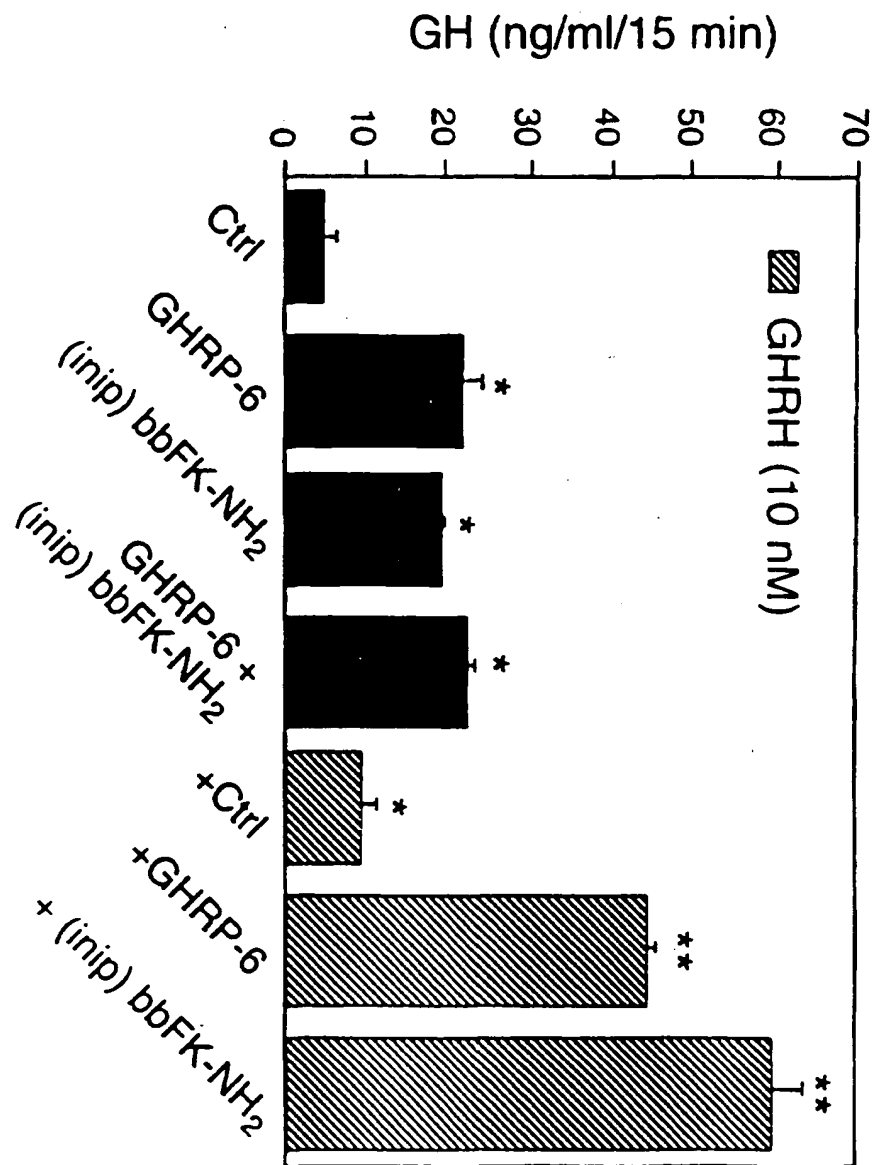


FIGURE 3

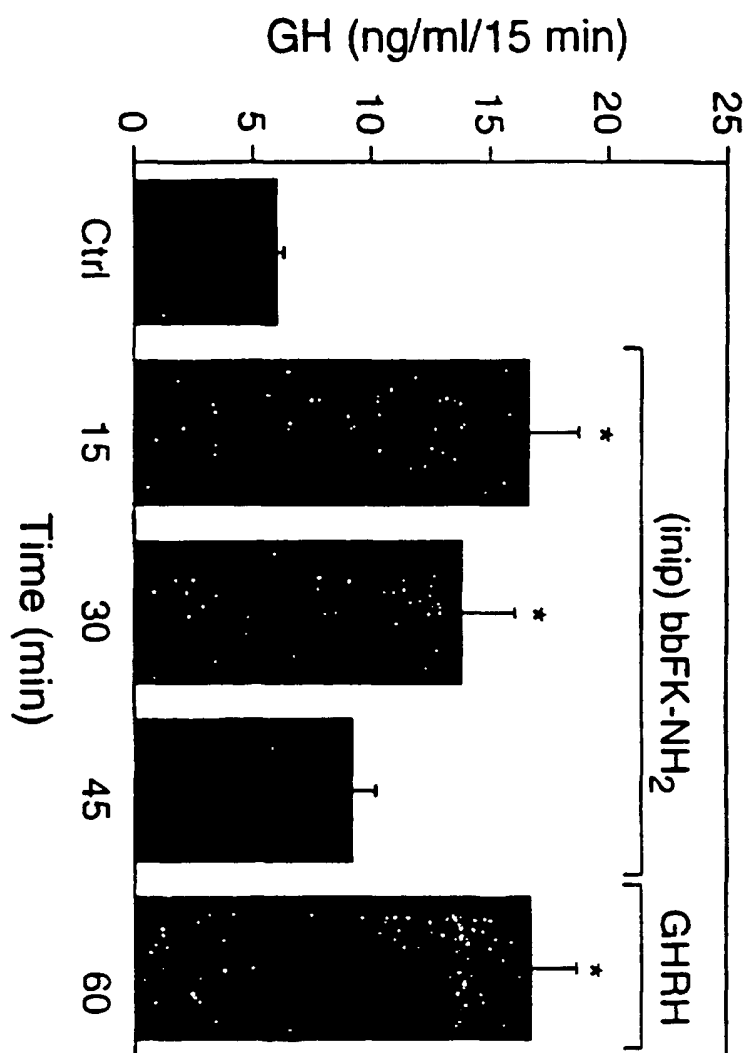


FIGURE 4

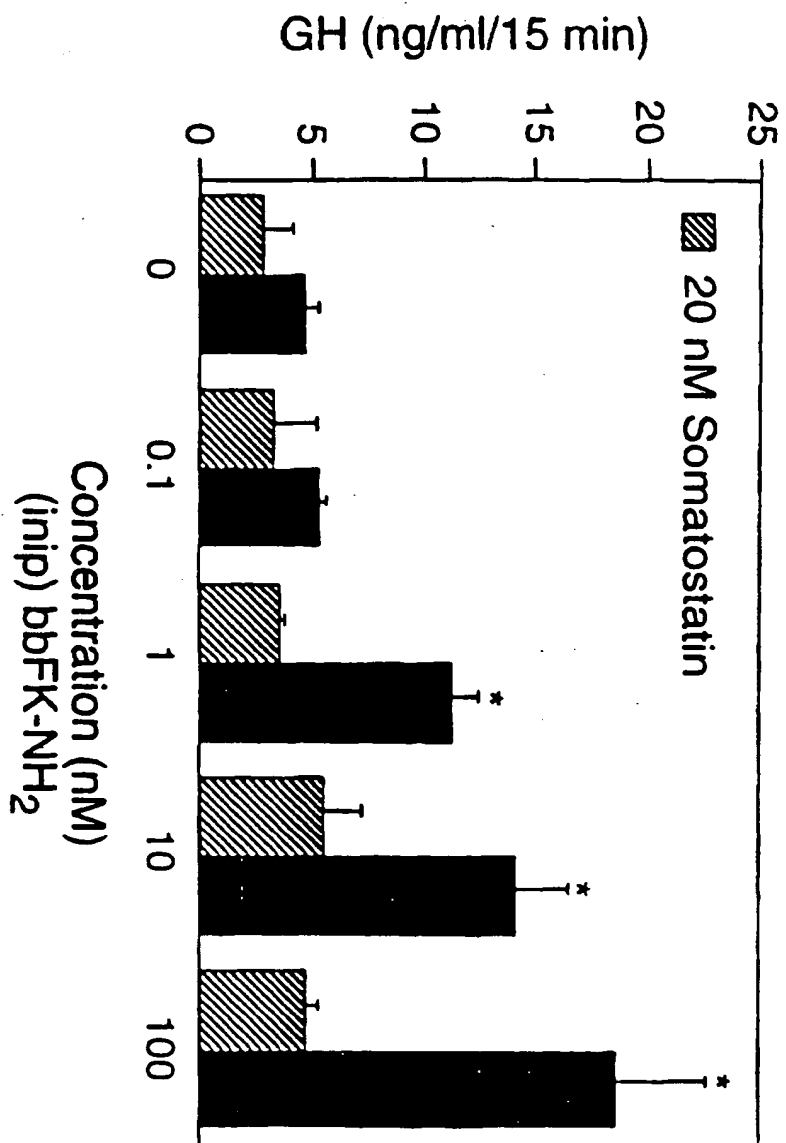


FIGURE 5

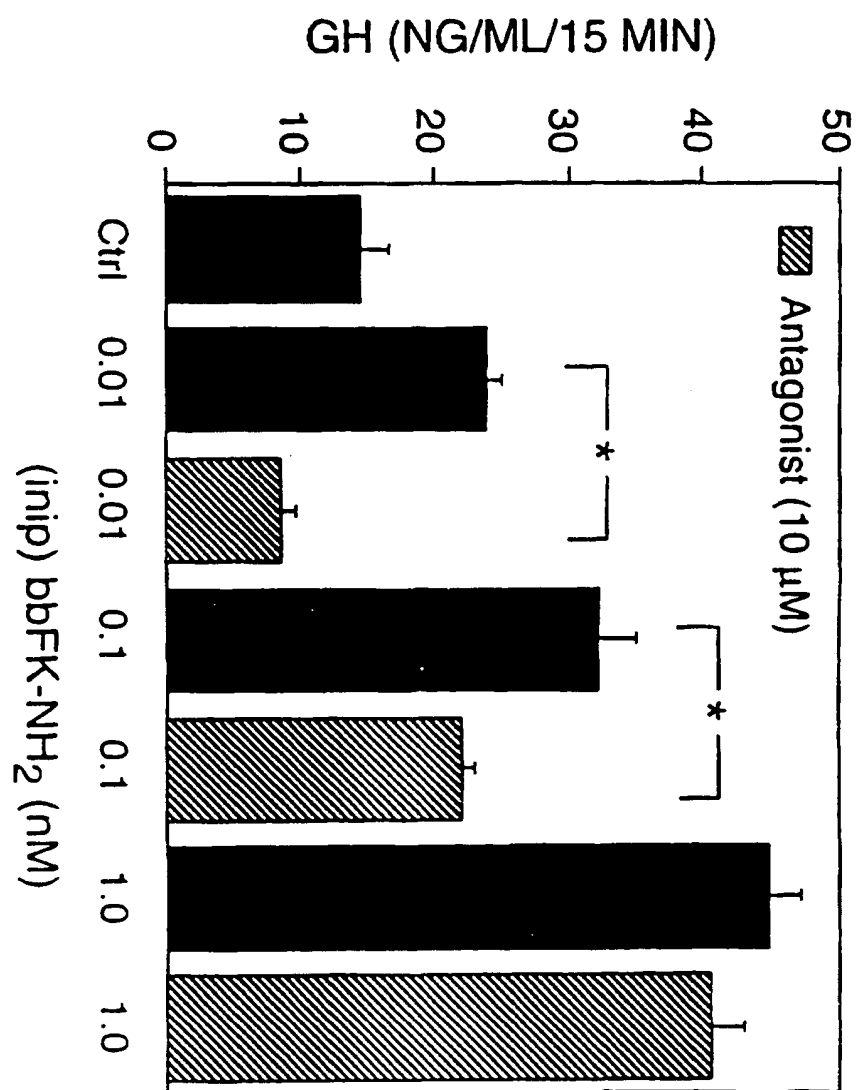


FIGURE 6

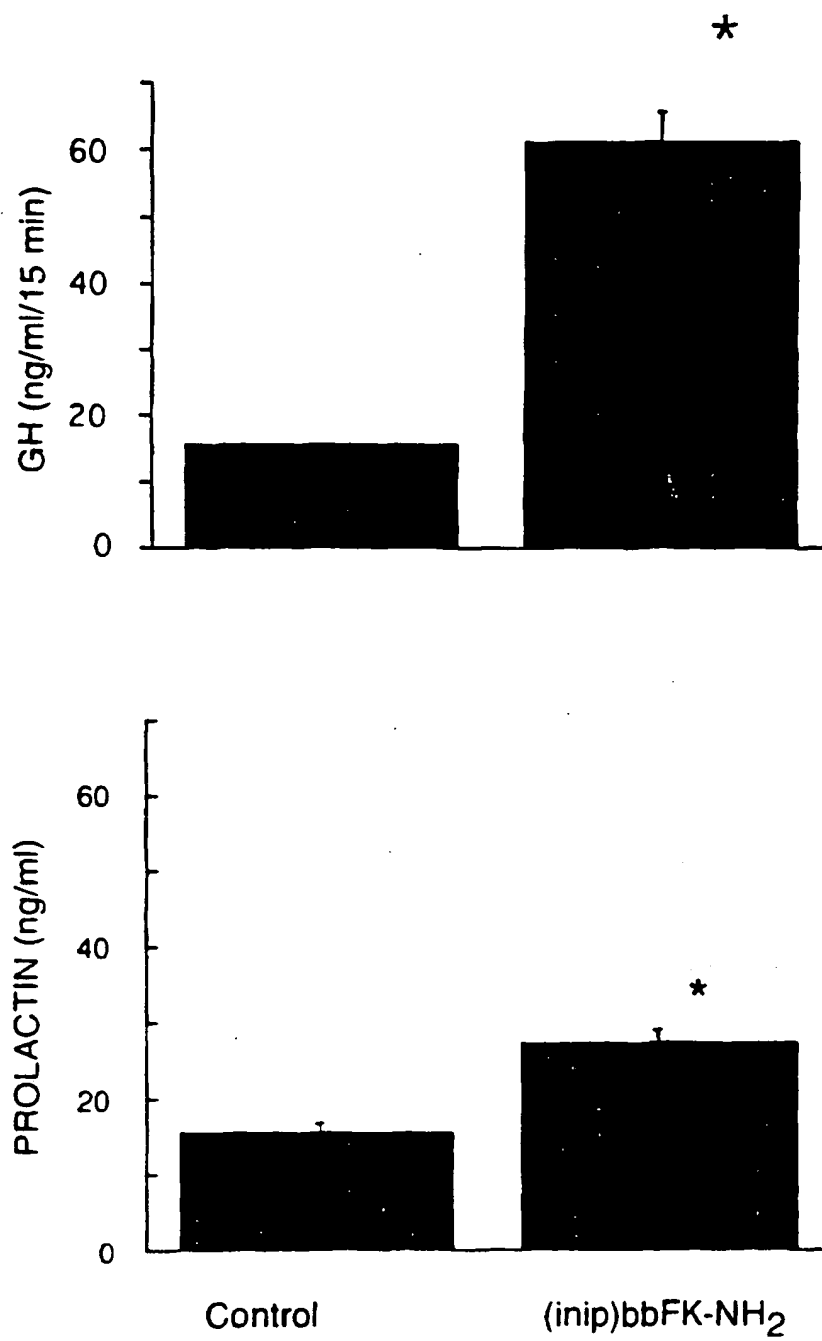


FIGURE 7

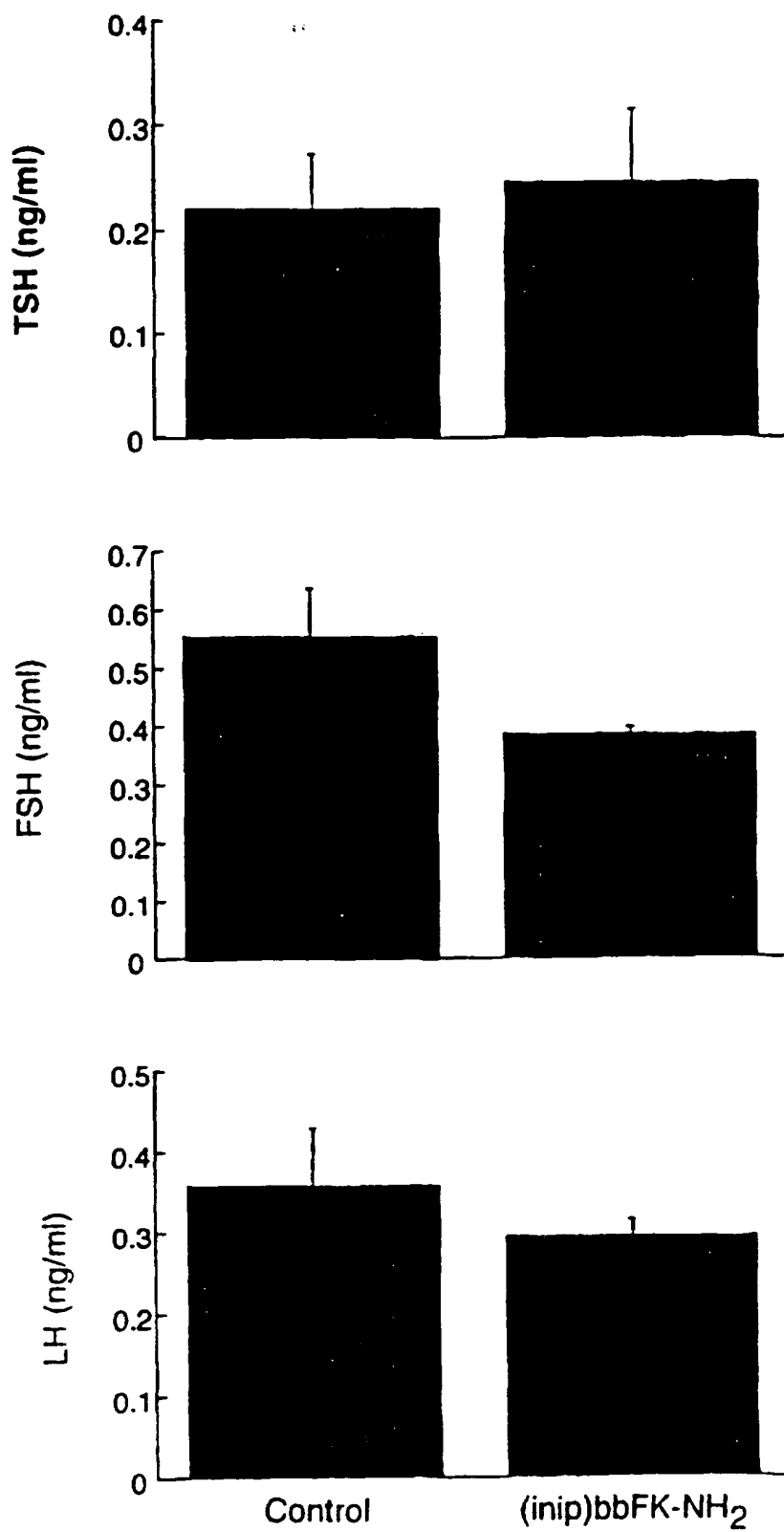


FIGURE 8

8/27

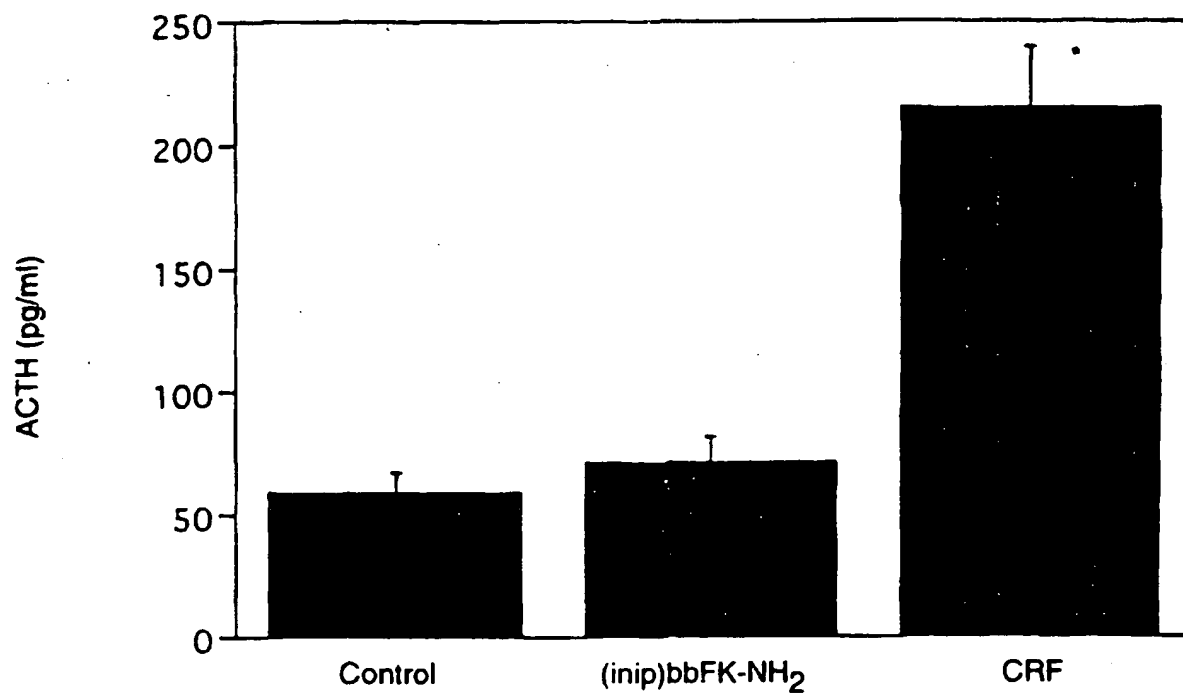


FIGURE 9

FIGURE 10B



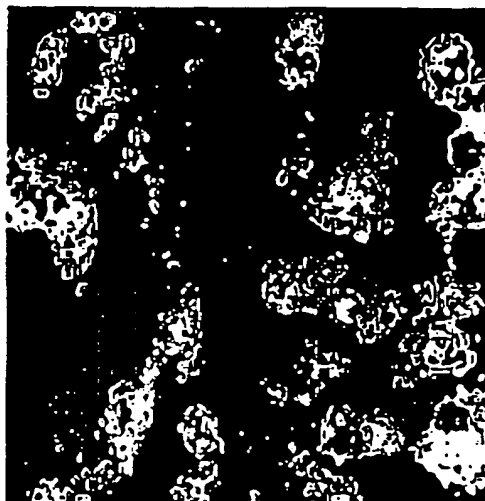
FIGURE 10D



FIGURE 10A



FIGURE 10C



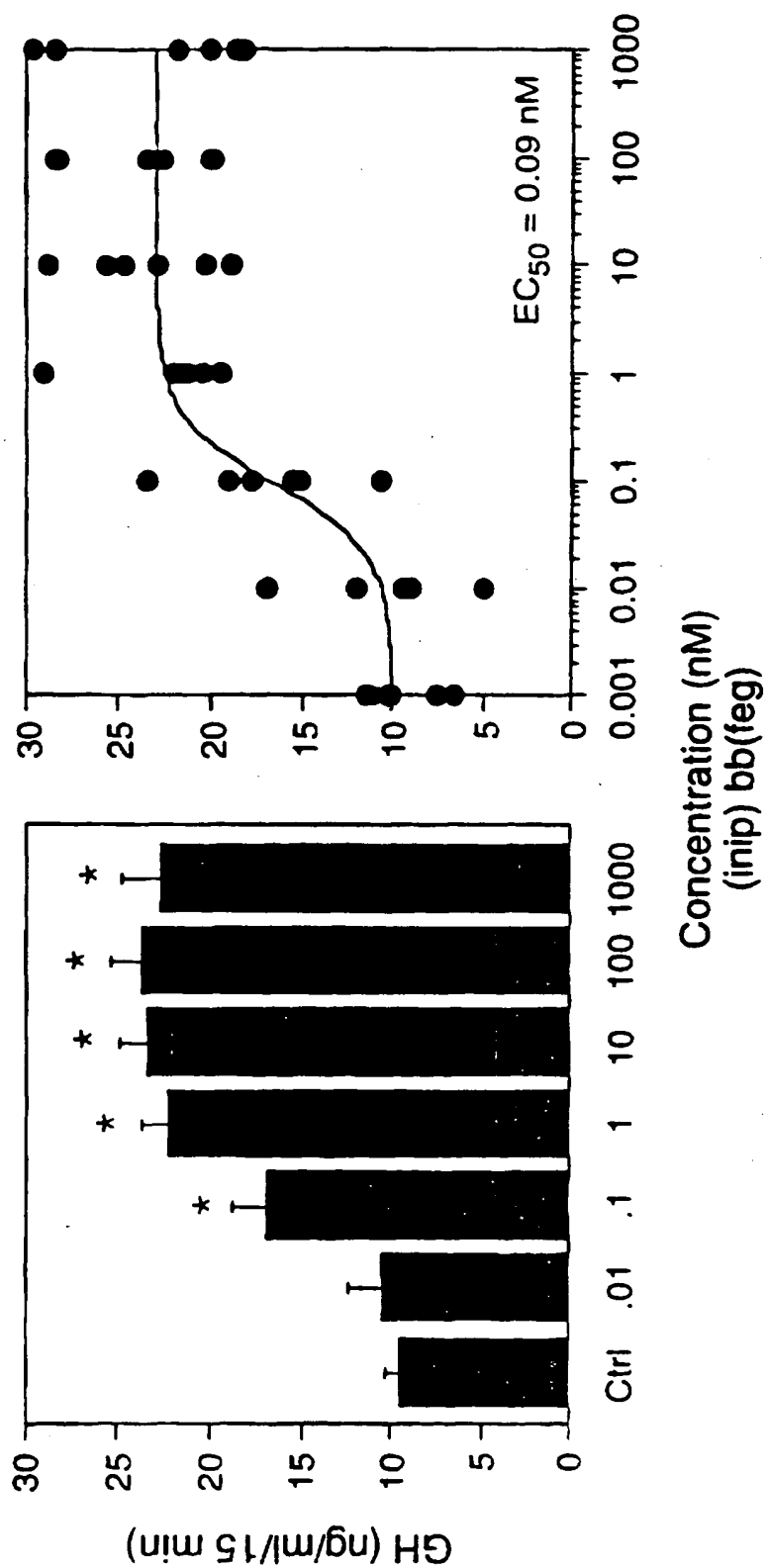


FIGURE 11

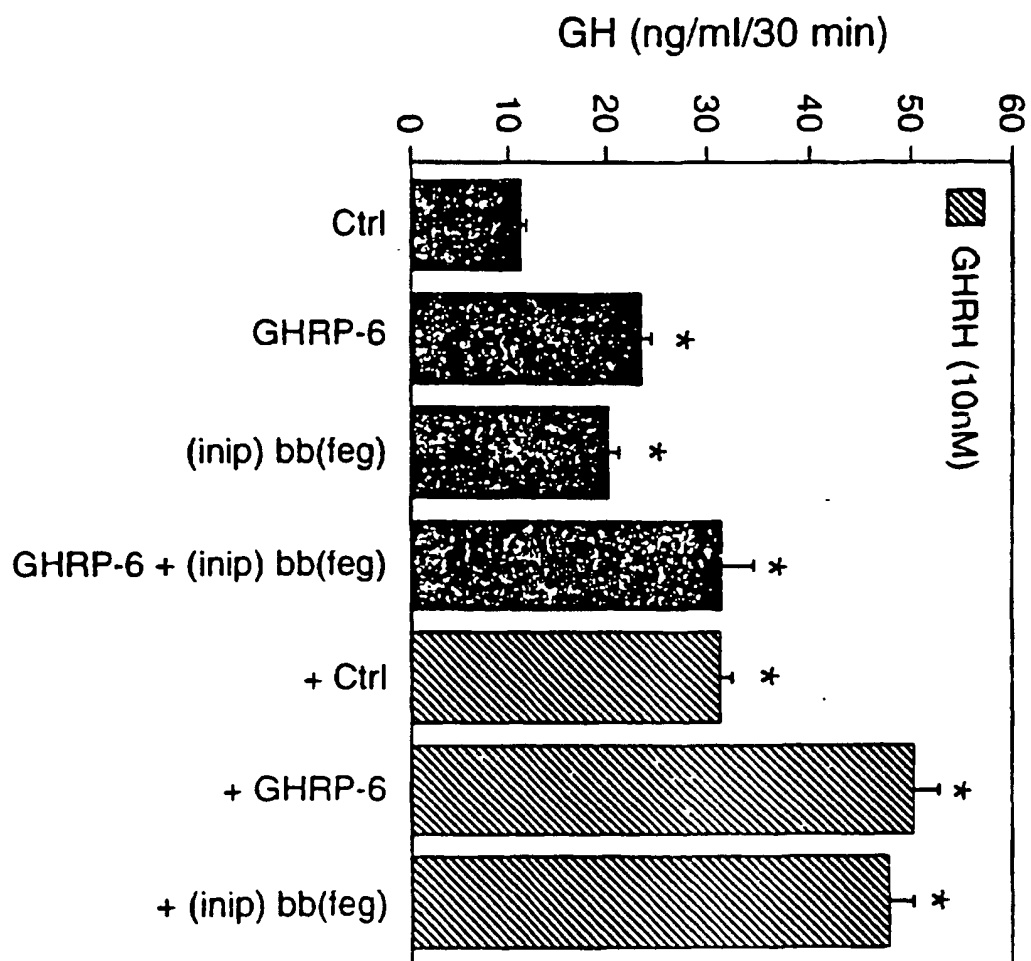


FIGURE 12

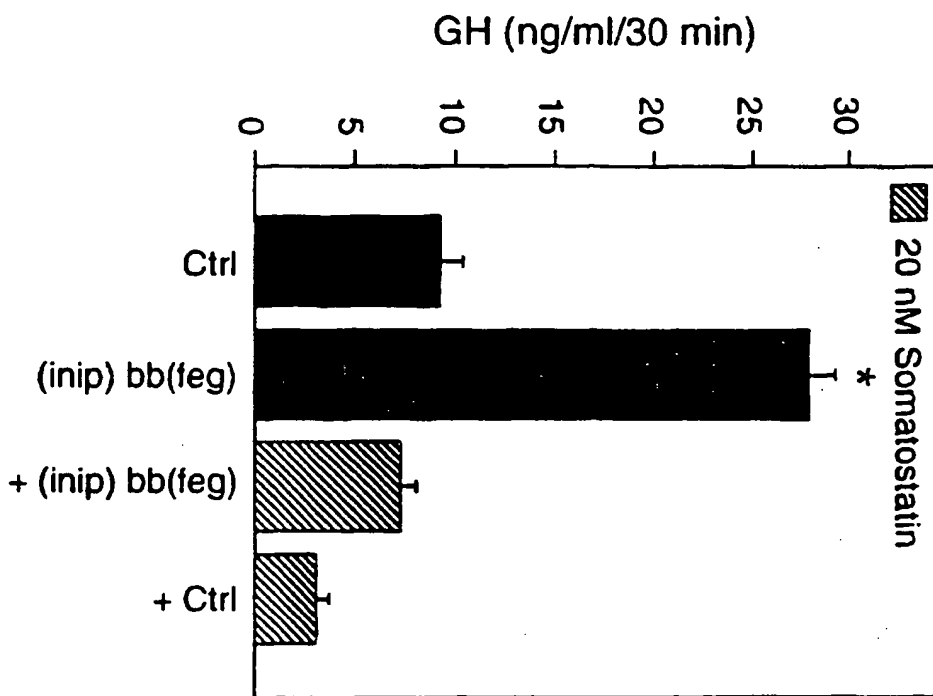


FIGURE 13

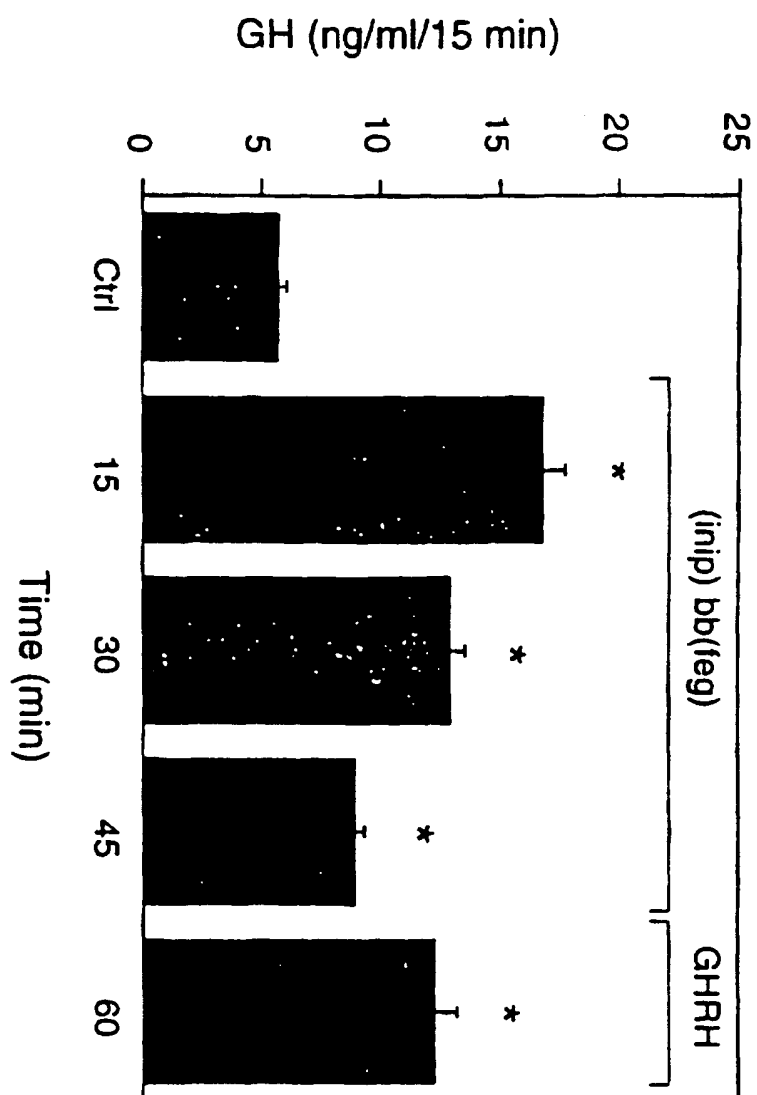


FIGURE 14

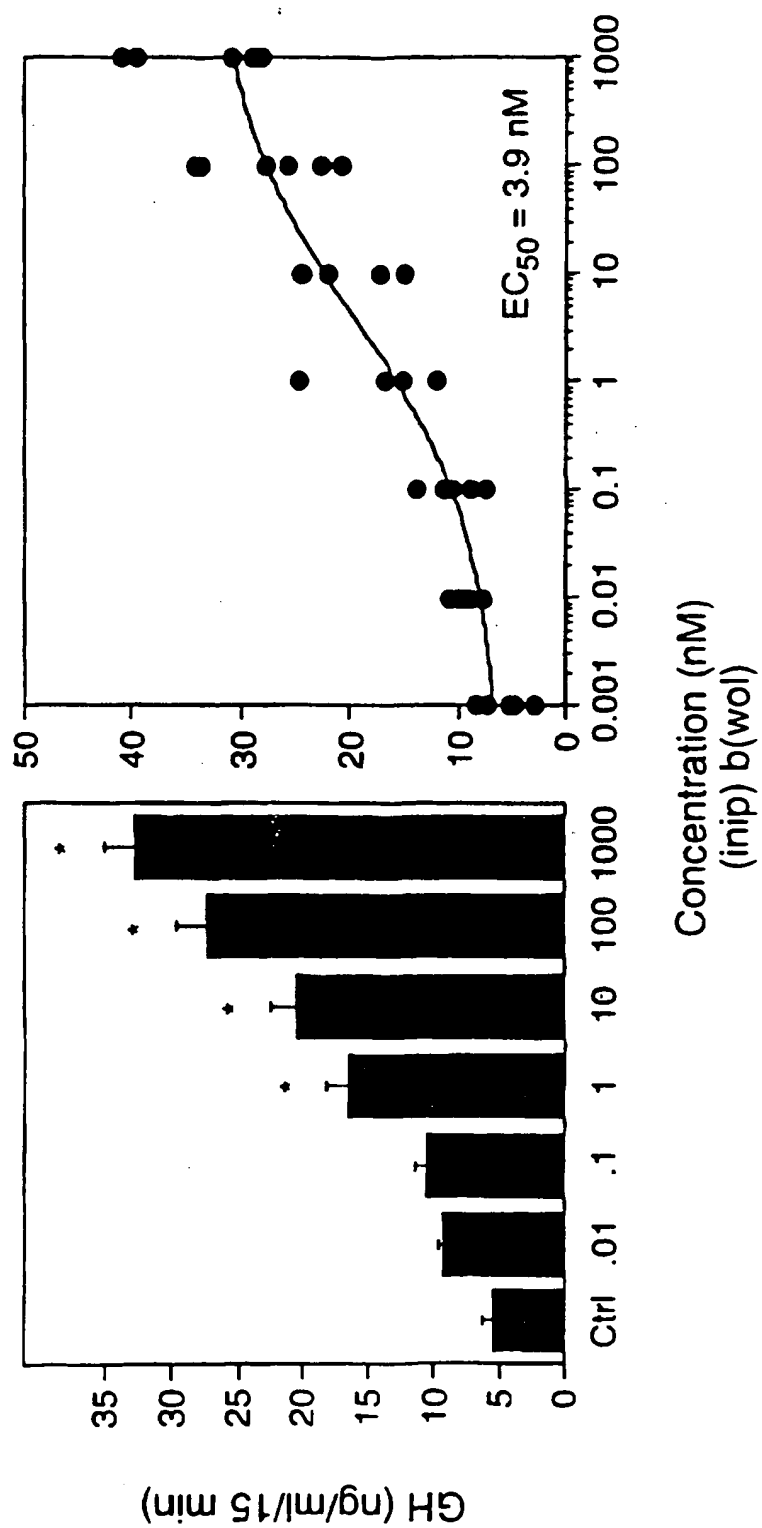


FIGURE 15

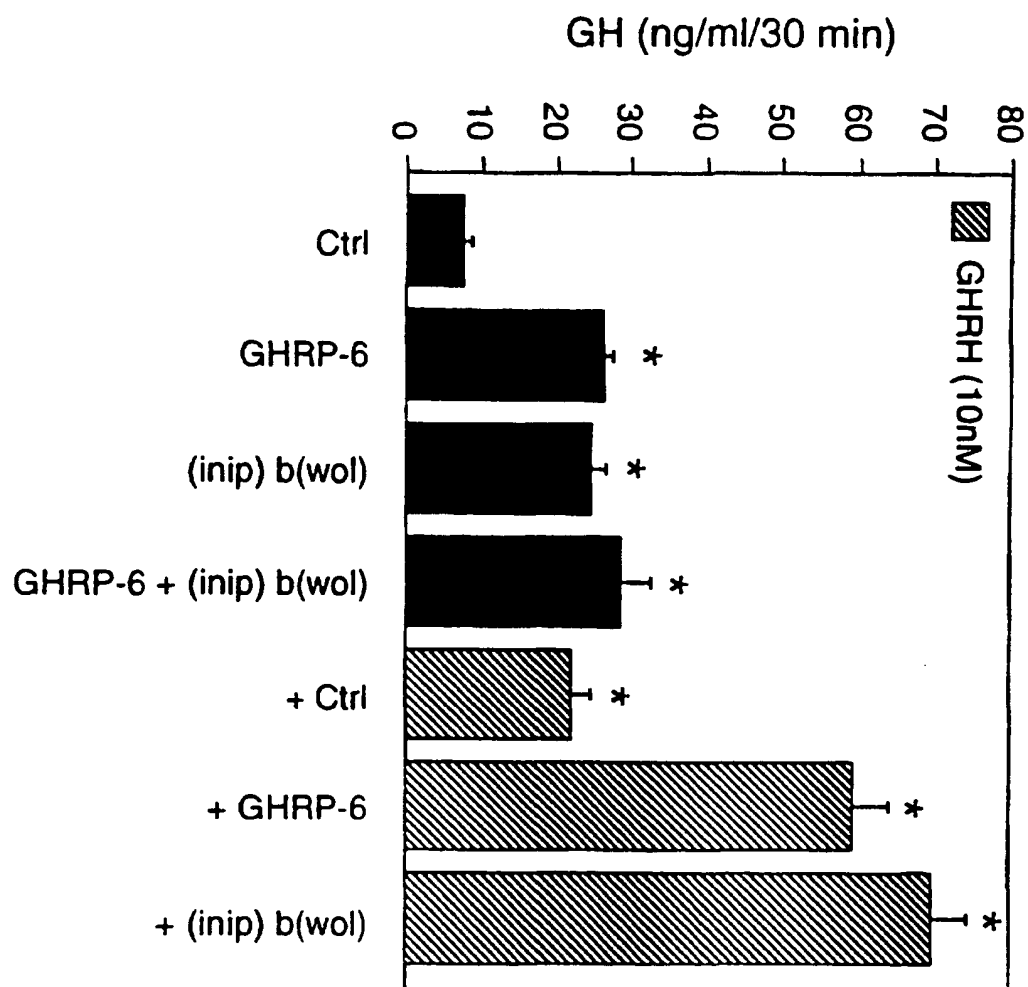


FIGURE 16

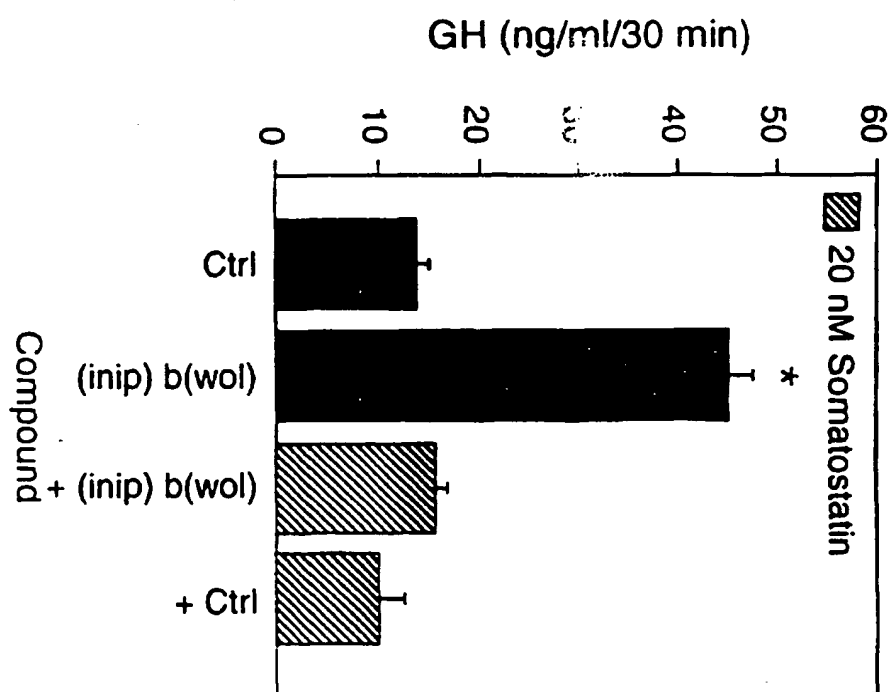


FIGURE 17

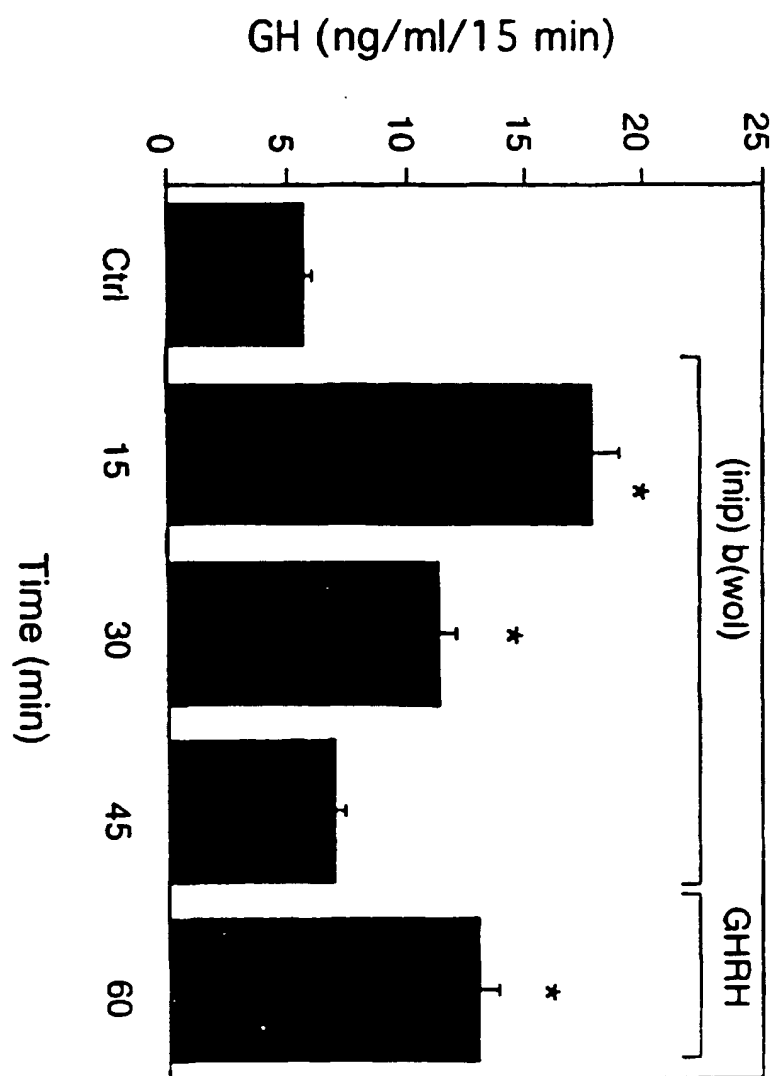
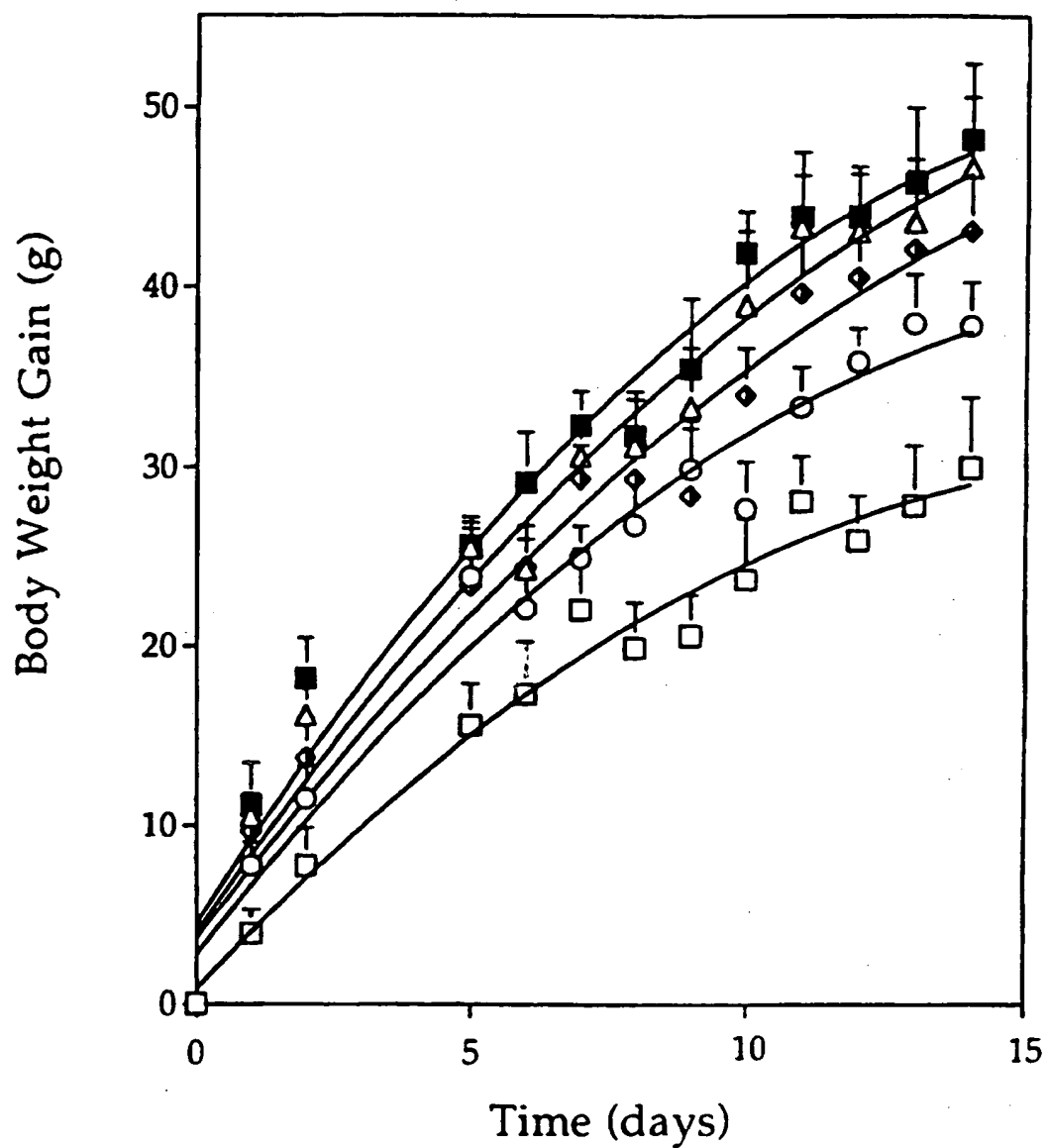


FIGURE 18



- Excipient
- (inip) b b F K-NH2 (4 µg)
- △ (inip) b b F K-NH2 (20 µg)
- ◆ (inip) b b F K-NH2 (100 µg)
- GHRH (600 µg)

FIGURE 19

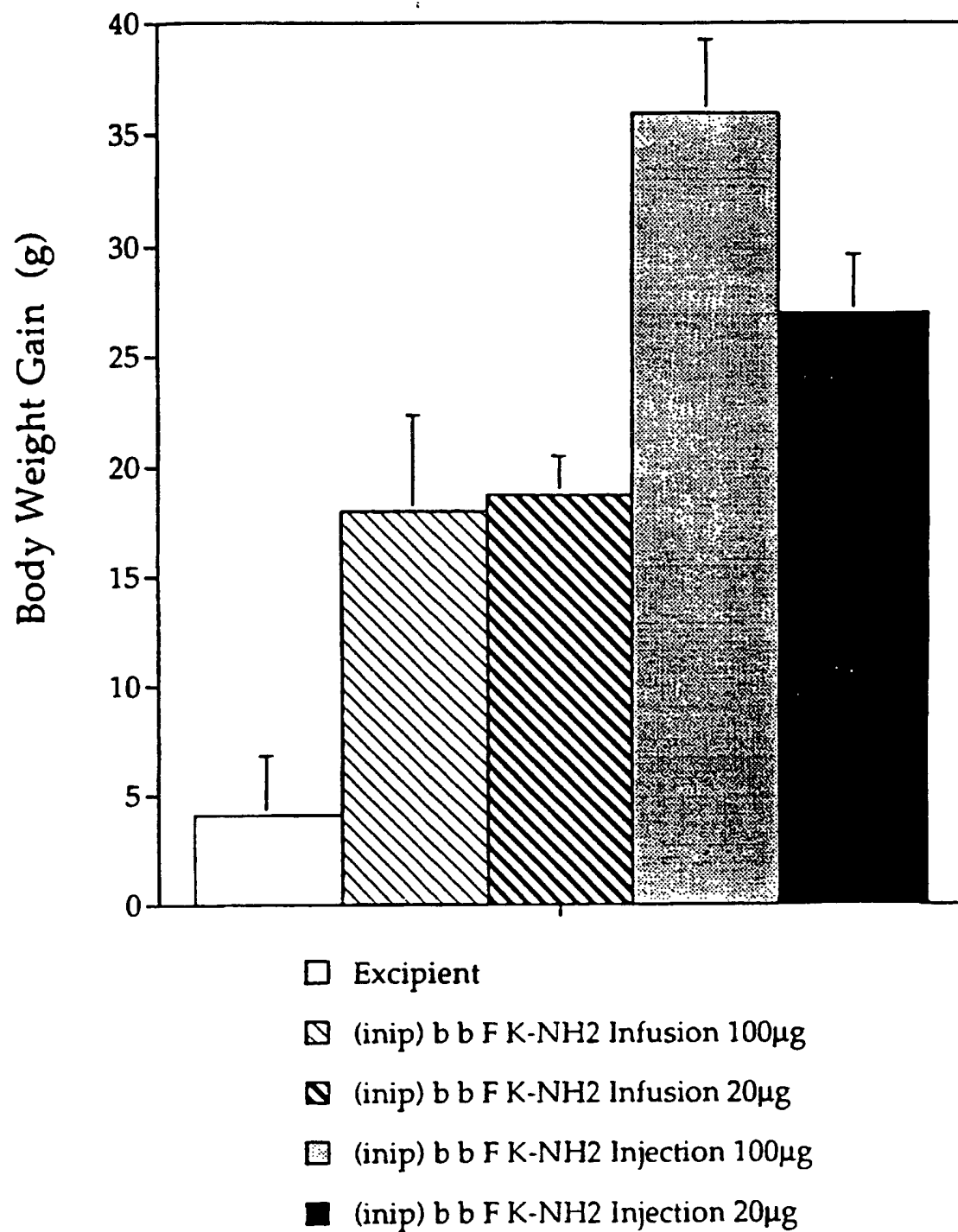


FIGURE 20

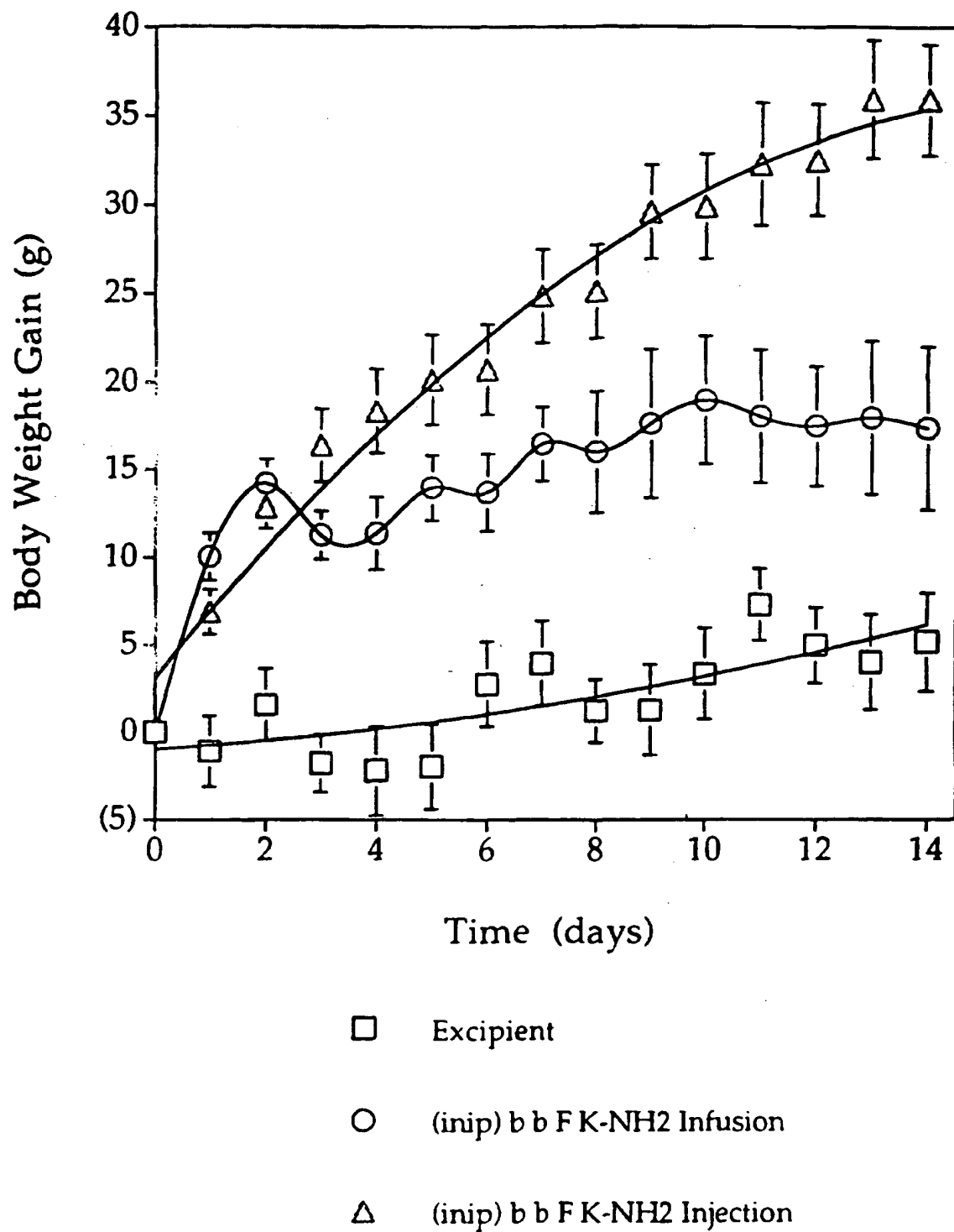


FIGURE 21

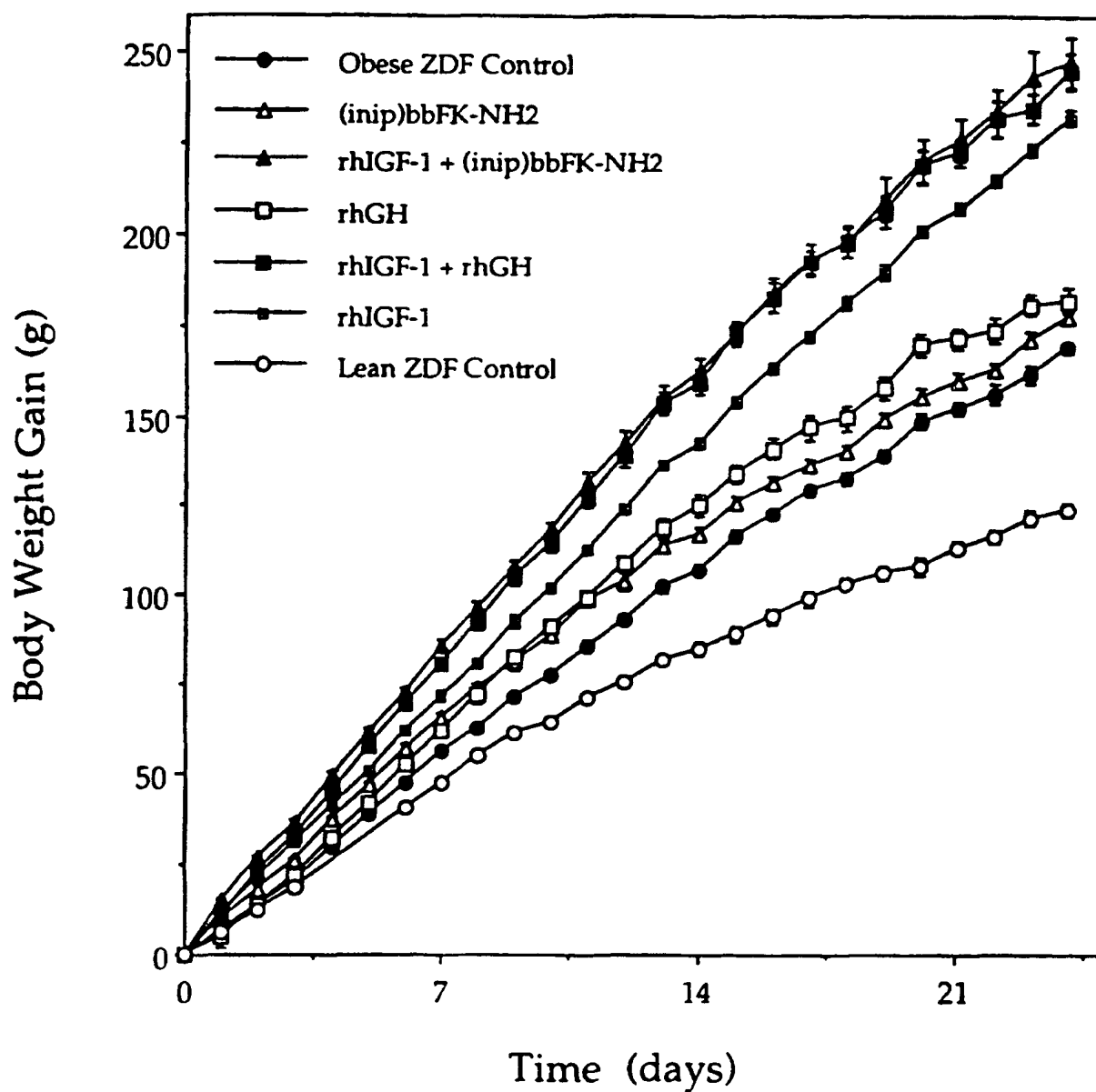


FIGURE 22

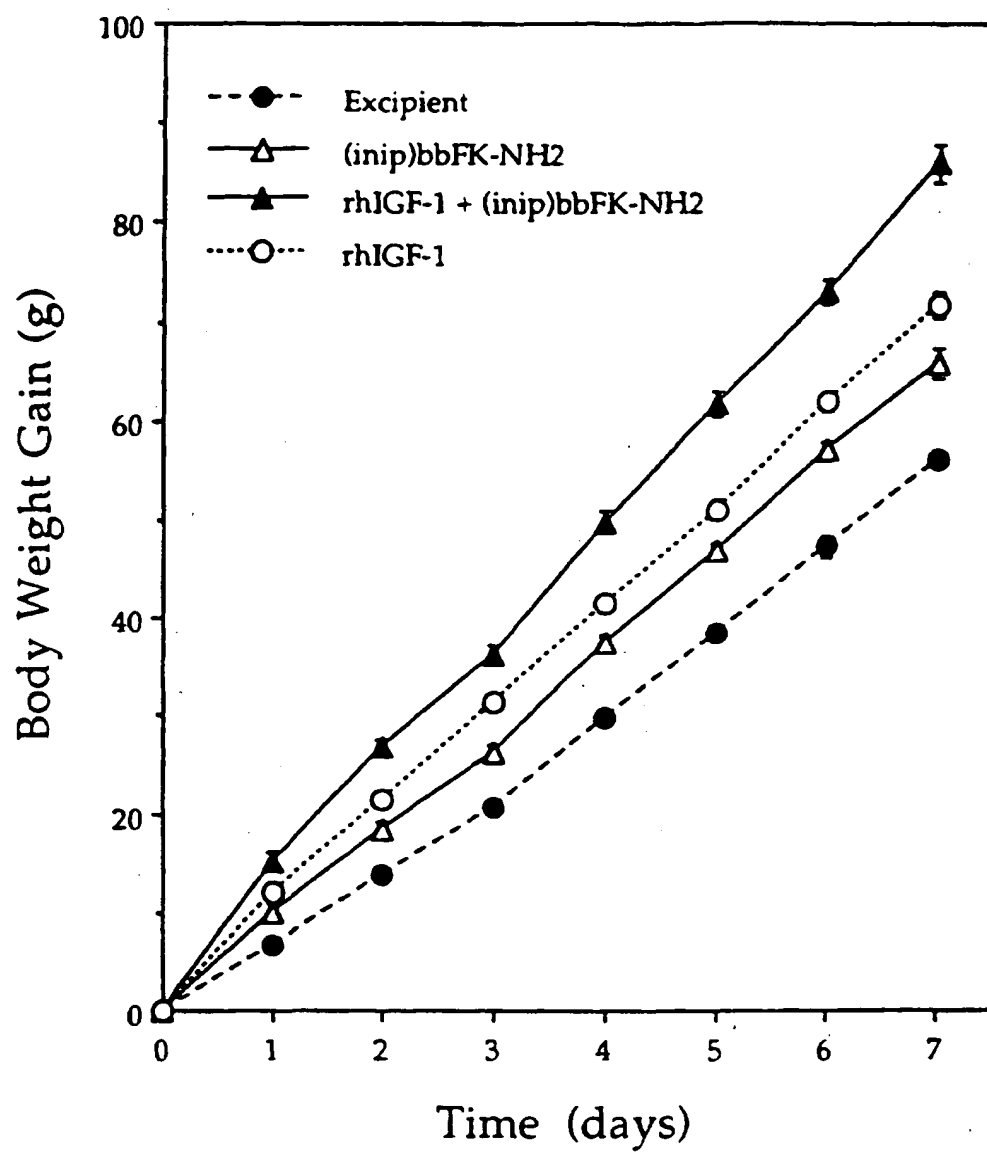


FIGURE 23

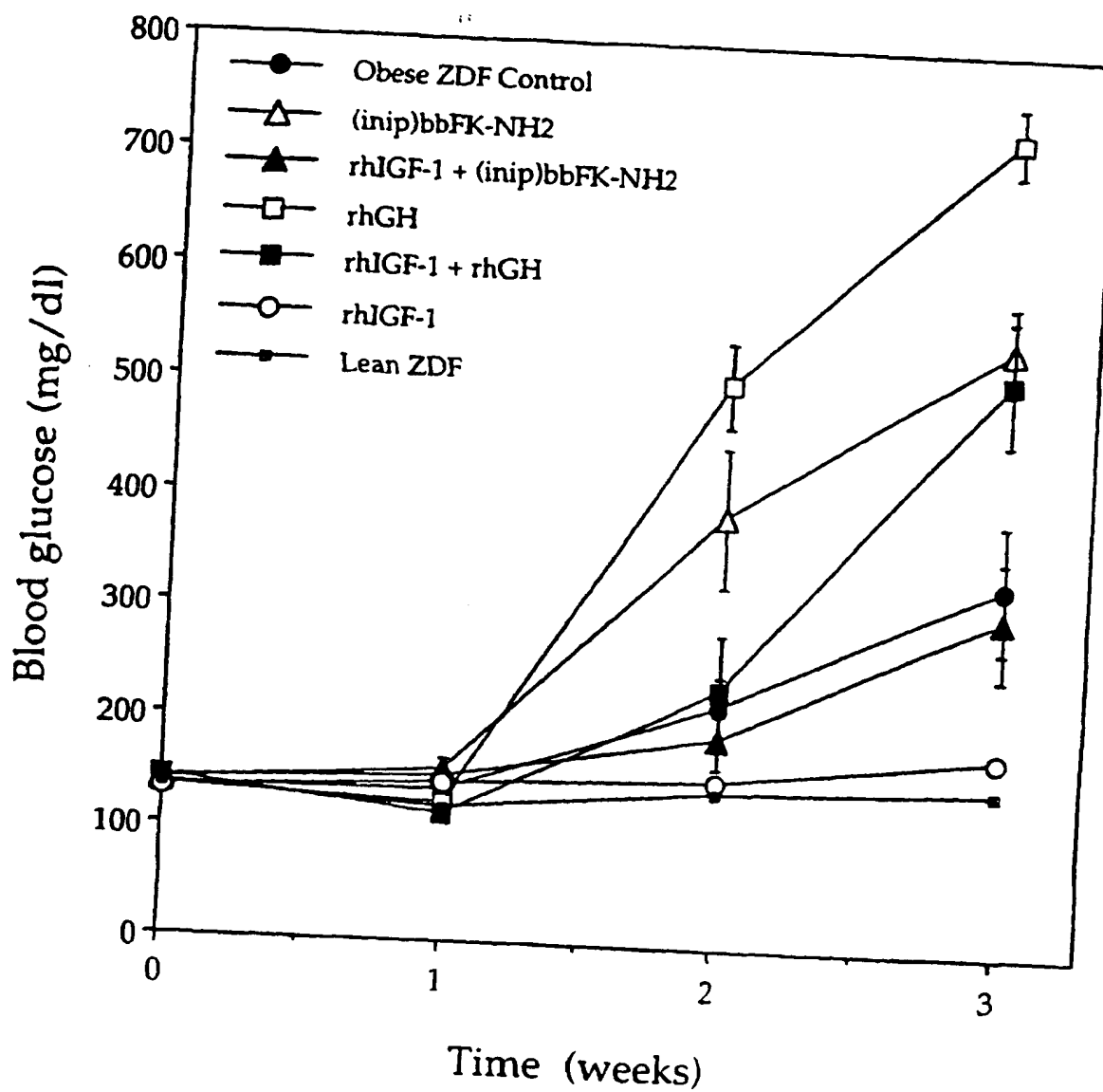


FIGURE 24

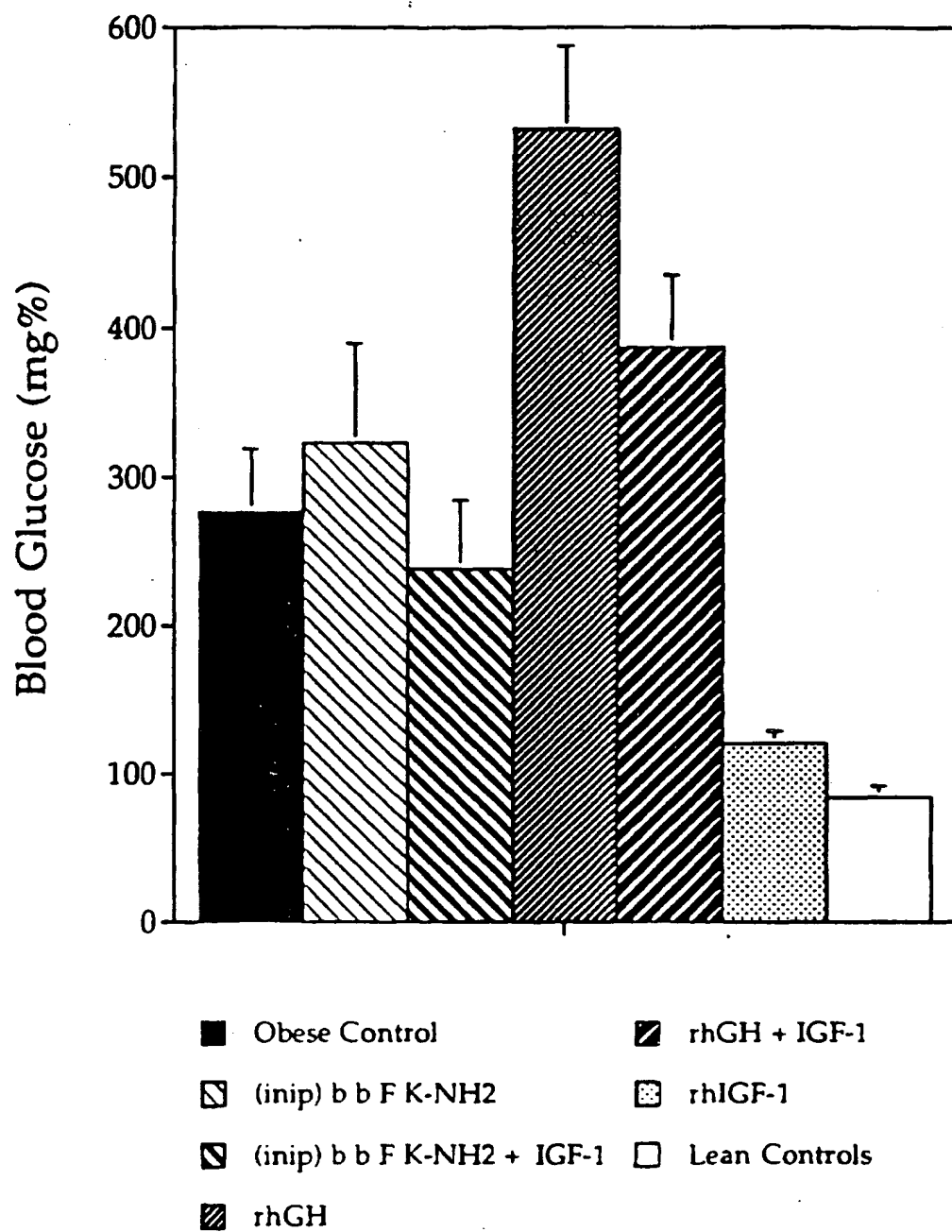


FIGURE 25

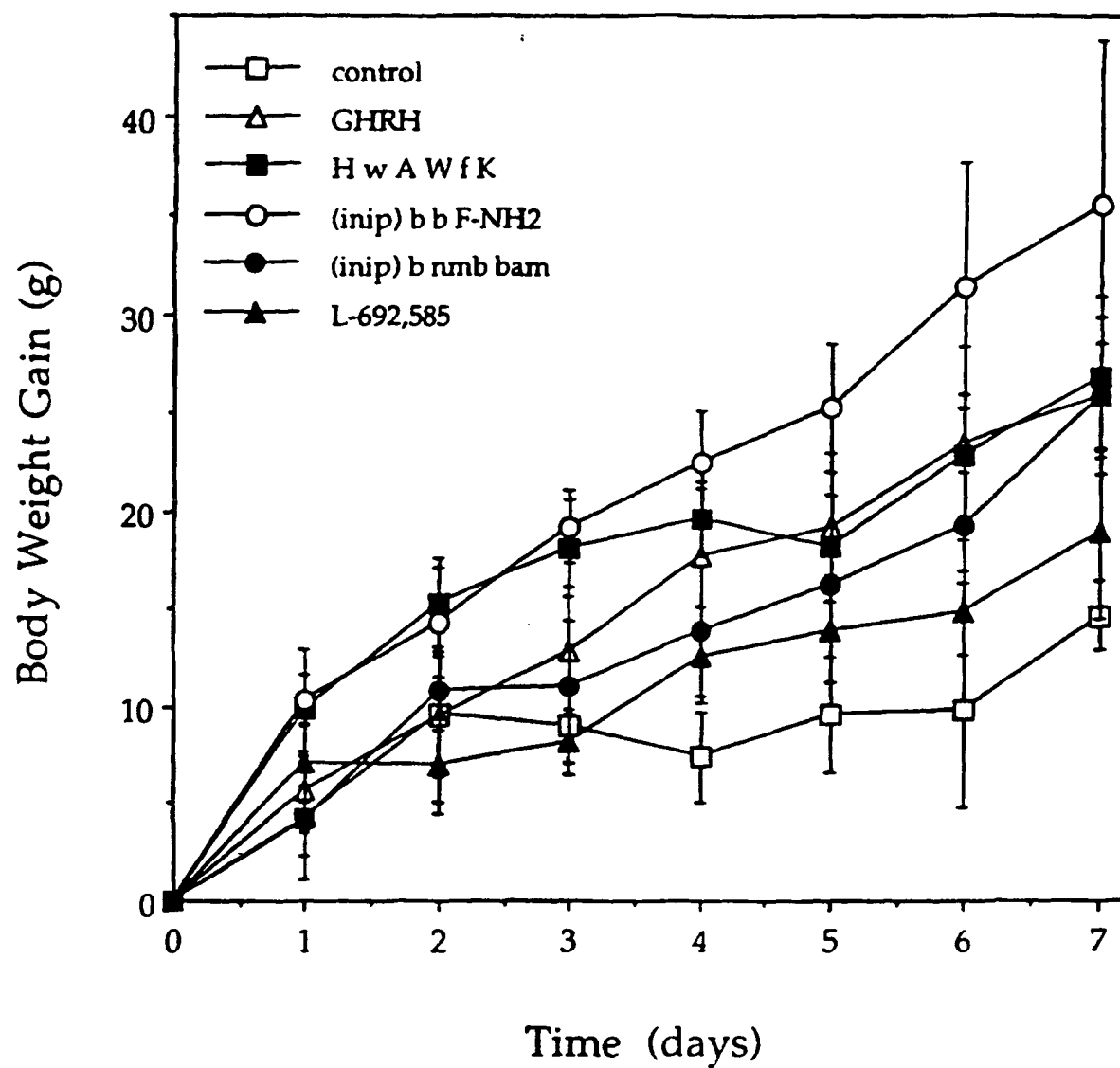


FIGURE 26

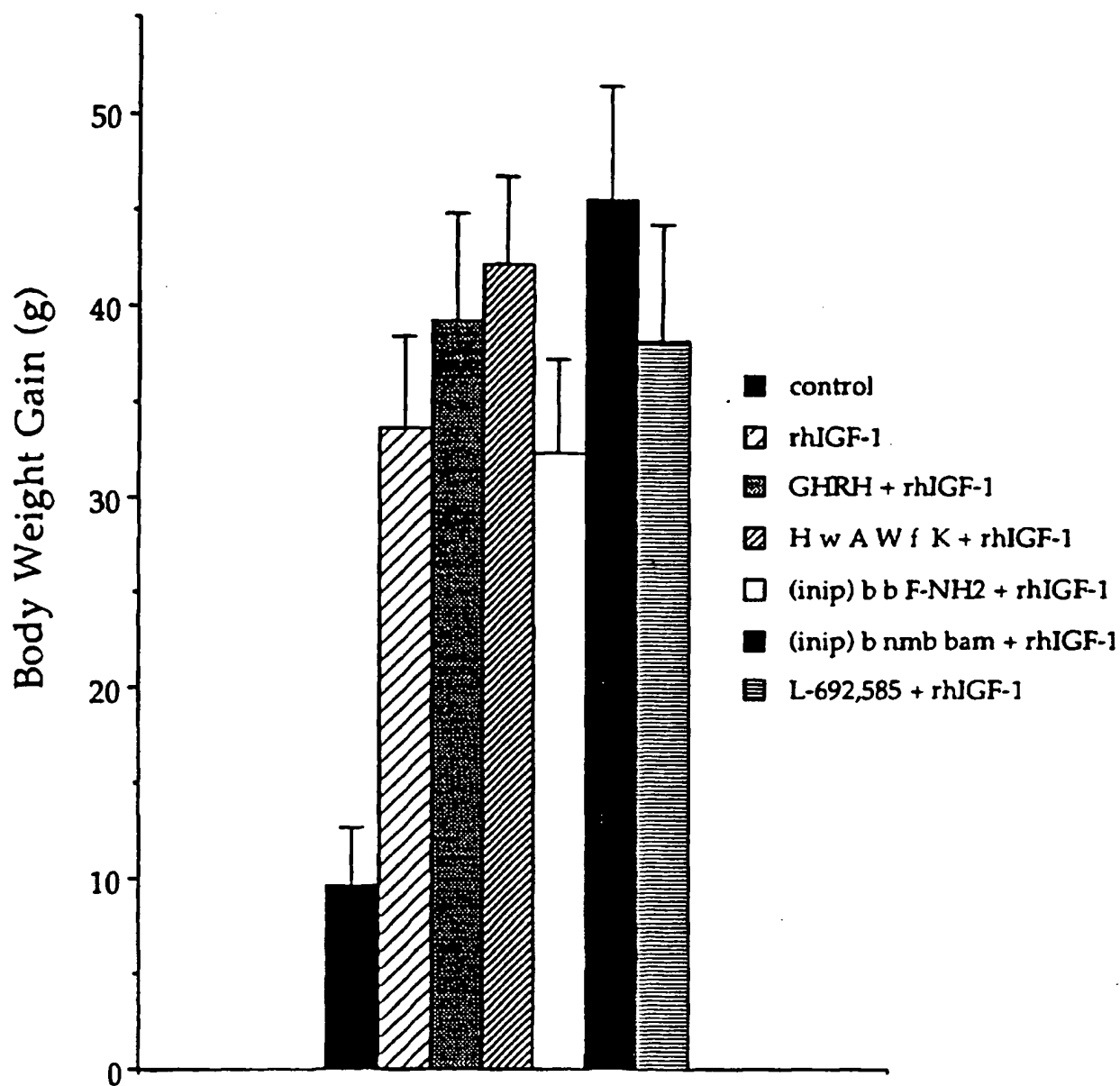


FIGURE 27

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau



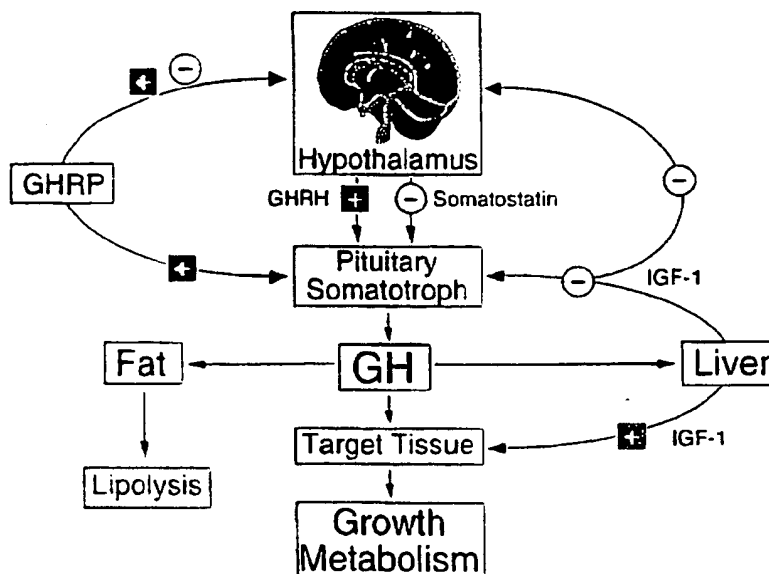
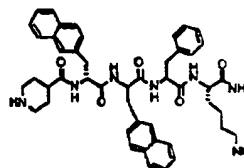
INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C07K 14/60, 5/023, 5/027, 5/03, 5/087, 5/107, 5/117, C07D 211/62, 401/12, A61K 38/18, 38/05, 38/06, 38/07, 31/445		A3	(11) International Publication Number: WO 96/15148
			(43) International Publication Date: 23 May 1996 (23.05.96)
(21) International Application Number: PCT/US95/14968		(81) Designated States: AL, AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TT, UA, UG, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, LS, MW, SD, SZ, UG).	
(22) International Filing Date: 16 November 1995 (16.11.95)		<p>Published With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</p> <p>(88) Date of publication of the international search report: 14 November 1996 (14.11.96)</p>	
(30) Priority Data: 08/340,767 16 November 1994 (16.11.94) US			
(71) Applicant: GENENTECH, INC. [US/US]; 460 Point San Bruno Boulevard, South San Francisco, CA 94080-4990 (US).			
(72) Inventors: SOMERS, Todd, C.; 706 Bounty Drive #610, Foster City, CA 94404 (US). ELIAS, Kathleen, A.; 2006 16th Avenue, San Francisco, CA 94116 (US). CLARK, Ross, G.; 711 Ursula Avenue, Pacifica, CA 94044 (US). McDOWELL, Robert, S.; 1264 Church Street, San Francisco, CA 94114 (US). STANLEY, Mark, S.; 284 Lauren Avenue, Pacifica, CA 94044 (US). BURNIER, John, P.; 202 Sterling Avenue, Pacifica, CA 94044 (US). RAWSON, Thomas, E.; 140 Hamilton Avenue, Mountain View, CA 94043 (US).			
(74) Agents: WINTER, Daryl, B. et al.; Genentech, Inc., 460 Point San Bruno Boulevard, South San Francisco, CA 94080-4990 (US).			

(54) Title: LOW MOLECULAR WEIGHT PEPTIDOMIMETIC GROWTH HORMONE SECRETAGOGUES

(57) Abstract

The present invention comprises growth hormone releasing peptides/peptidomimetics (GHRP) capable of causing release of growth hormone from the pituitary. Compositions containing the GHRP's of this invention are used to promote growth in mammals either alone or in combination with other growth promoting compounds, especially IGF-1. In a method of this invention GHRP's in combination with IGF-1 are used to treat Type II diabetes. An exemplary compound (1) of this invention is provided above.



FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	GB	United Kingdom	MR	Mauritania
AU	Australia	GE	Georgia	MW	Malawi
BB	Barbados	GN	Guinea	NE	Niger
BE	Belgium	GR	Greece	NL	Netherlands
BF	Burkina Faso	HU	Hungary	NO	Norway
BG	Bulgaria	IE	Ireland	NZ	New Zealand
BJ	Benin	IT	Italy	PL	Poland
BR	Brazil	JP	Japan	PT	Portugal
BY	Belarus	KE	Kenya	RO	Romania
CA	Canada	KG	Kyrgyzstan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo	KR	Republic of Korea	SE	Sweden
CH	Switzerland	KZ	Kazakhstan	SI	Slovenia
CI	Côte d'Ivoire	LI	Liechtenstein	SK	Slovakia
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CN	China	LU	Luxembourg	TD	Chad
CS	Czechoslovakia	LV	Latvia	TG	Togo
CZ	Czech Republic	MC	Monaco	TJ	Tajikistan
DE	Germany	MD	Republic of Moldova	TT	Trinidad and Tobago
DK	Denmark	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	US	United States of America
FI	Finland	MN	Mongolia	UZ	Uzbekistan
FR	France			VN	Viet Nam
GA	Gabon				

INTERNATIONAL SEARCH REPORT

International Application No.

PLI/US 95/14968

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C07K14/60 C07K5/023 C07K5/027 C07K5/03 C07K5/087
 C07K5/107 C07K5/117 C07D211/62 C07D401/12 A61K38/18
 A61K38/05 A61K38/06 A61K38/07 A61K31/445

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C07K C07D A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P, X	WO, A, 95 17423 (NOVONORDISK AS ; JOHANSEN NILS LANGELAND (DK); LAU JESPER (DK); MAD) 29 June 1995 see claims; examples 11, 25, 28-30 ---	1-18
X	WO, A, 93 04081 (UNIV TULANE) 4 March 1993 see page 31, last paragraph; claims ---	1-18
A	EP, A, 0 018 072 (BECKMAN INSTRUMENTS INC) 29 October 1980 see claims; examples ---	1-18
	-/--	

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- *A* document member of the same patent family

Date of the actual completion of the international search

24 September 1996

Date of mailing of the international search report

03.10.96

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentaan 2
 NL - 2280 HV Rijswijk
 Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,
 Fax: (+ 31-70) 340-3016

Authorized officer

Fuhr, C

INTERNATIONAL SEARCH REPORT

Inv. No. Application No

PCT/US 95/14968

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	LIFE SCIENCES, vol. 54, no. 18, US, pages 1321-1328, XP002013882 R. DEGHENGHI ET AL.: "GH-Releasing Activity of Hexarelin, a New Growth Hormone Releasing Peptide, in Infant and Adult Rats" see table 1	1-18
X	--- EP,A,0 274 259 (SANKYO CO) 13 July 1988 see table 6, page 52, compounds 6-183 -- 6-190; page 23, formula I-6; see page 19, line 46 - page 20, line 29; claims	1
P,X	--- INT. J. PEPT. PROTEIN RES. (1995), 45(6), 508-13 CODEN: IJPPC3;ISSN: 0367-8377, XP002013883 STAVROPOULOS, GEORGE ET AL: "Synthesis of potent agonists of substance P by replacement of Met11 with Glu(OBzl) and N-terminal glutamine with Gln of the C-terminal hexapeptide and heptapeptide of substance P" see page 511, right-hand column, paragraph 4 - page 512, left-hand column, paragraph 1	1
X	--- CHEMICAL ABSTRACTS, vol. 116, no. 11, 16 March 1992 Columbus, Ohio, US; abstract no. 106743, EGOROVA, S. V. ET AL: "Synthesis of shortened analogs of substance P with modification of the glutamine residue at position 6" XP002013884 see abstract & VESTSI AKAD. NAVUK BSSR, SER. KHIM. NAVUK (1991), (5), 61-7 CODEN: VBSKAK;ISSN: 0002-3590,	1
X	--- CHEMICAL ABSTRACTS, vol. 92, no. 1, 7 January 1980 Columbus, Ohio, US; abstract no. 2280, LIN, TSAU-YEN ET AL: "Inhibition of cathepsin D by synthetic oligopeptides" XP002013885 see abstract & J. BIOL. CHEM. (1979), 254(23), 11875-83 CODEN: JBCHA3;ISSN: 0021-9258, ---	1
	--- -/--	

3

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 95/14968

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
E	WO,A,95 34311 (MERCK & CO INC ;NARGUND RAVI (US); BARAKAT KHALED J (US); CHEN MEN) 21 December 1995 see claims; examples ---	1
P,X	WO,A,95 13069 (MERCK & CO INC ;MORRIELLO GREGORI J (US); PATCHETT ARTHUR A (US);) 18 May 1995 see claims; examples ---	1
X	WO,A,94 13696 (MERCK & CO INC ;CHEN MENG HSIN (US); JOHNSTON DAVID B R (US); NARG) 23 June 1994 see claims; examples -----	1

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 95/ 14968

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 18-20
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although these claims are directed to methods of treatment of the human and/or animal body, the search has been carried out and based on the alleged effects of the compound and/or composition.
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. claims 1-20 partially
2. claims 1-20 partially

- see continuation-sheet PCT/ISA/210 -

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-20 partially (invention 1).

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

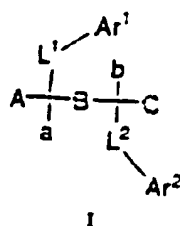
INTERNATIONAL SEARCH REPORT

International Application No. PCT/US95/ 14968

FURTHER INFORMATION CONTINUED FROM PCT/ISA/210

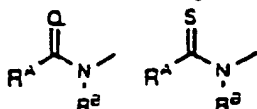
1. The group of compounds of formula I

Formula (I)



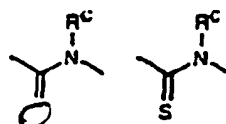
where

A is selected from the group



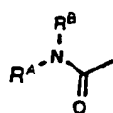
and

B is selected from the group



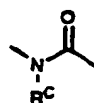
in combination with each other,
or where

A is



and

B is



in combination with each other, and their use.

2. Group of compounds of formula I not covered by the first invention.

- The compounds of the claims 2-16 are not completely dependent of claim 1 due to the fact that substituent X in formulae II to V is not completely covered by substituent C in formula I. The search for those compounds is limited to the ones covered by claim 1.
- In view of the extremely large number of compounds still falling under the first invention, and of the absence of any technical support for these compounds in the description, the ISA considers that it is not economically reasonable to draw a search report covering the entire subject matter characterized above. The search has therefore been limited to the real examples given in the description and closely related ones which comply with the first invention.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 95/14968

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-9517423	29-06-95	AU-A- 1272495	10-07-95
		CA-A- 2179597	29-06-95
		HU-A- 73497	28-08-96
		ZA-A- 9410261	23-06-95

WO-A-9304081	04-03-93	WO-A- 9220767	26-11-92
		AU-B- 666673	22-02-96
		AU-A- 2541692	16-03-93
		BG-A- 98489	28-02-95
		BR-A- 9206398	27-12-94
		CA-A- 2116120	23-02-93
		CN-A- 1073684	30-06-93
		CZ-A- 9400400	16-11-94
		EP-A- 0605484	13-07-94
		FI-A- 940807	21-02-94
		HU-A- 69178	28-08-95
		JP-T- 7507039	03-08-95
		NO-A- 940592	14-04-94
		NZ-A- 244034	28-08-95
		PL-B- 169562	30-08-96
		SK-A- 20494	05-10-94
		ZA-A- 9206337	22-04-93

EP-A-0018072	29-10-80	US-A- 4223019	16-09-80
		US-A- 4228156	14-10-80
		US-A- 4228157	14-10-80
		US-A- 4228155	14-10-80
		US-A- 4224316	23-09-80
		US-A- 4226857	07-10-80
		US-A- 4228158	14-10-80
		US-A- 4223020	16-09-80
		US-A- 4223021	16-09-80
		AU-A- 5680180	02-10-80
		CA-A- 1175810	09-10-84
		JP-C- 1513250	24-08-89
		JP-A- 61289098	19-12-86
		JP-B- 63060039	22-11-88
		JP-C- 1385850	26-06-87
		JP-A- 55133344	17-10-80
		JP-B- 61046000	11-10-86

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 95/14968

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP-A-0018072		JP-B- 1049359	24-10-89
		JP-C- 1566147	25-06-90
		JP-A- 59216860	06-12-84

EP-A-0274259	13-07-88	DE-D- 3789101	24-03-94
		DE-T- 3789101	21-07-94
		ES-T- 2062988	01-01-95
		JP-A- 1199994	11-08-89
		US-A- 5378690	03-01-95
		JP-A- 1063559	09-03-89
		JP-B- 7116118	13-12-95

WO-A-9534311	21-12-95	AU-A- 2695795	05-01-96

WO-A-9513069	18-05-95	US-A- 5494919	27-02-96
		US-A- 5492916	20-02-96
		US-A- 5492920	20-02-96
		AU-A- 1172995	29-05-95
		CA-A- 2175218	18-05-95
		NO-A- 961865	08-07-96

WO-A-9413696	23-06-94	US-A- 5536716	16-07-96
		AU-A- 5232093	23-06-94
		BG-A- 99710	31-01-96
		CA-A- 2110670	12-06-94
		CN-A- 1092071	14-09-94
		CZ-A- 9501514	13-12-95
		EP-A- 0615977	21-09-94
		FI-A- 952863	09-06-95
		HU-A- 72076	28-03-96
		JP-B- 2509530	19-06-96
		JP-A- 6263737	20-09-94
		NO-A- 952295	10-08-95
		PL-A- 309331	02-10-95
		SI-A- 9300646	30-09-94
		SK-A- 75995	08-11-95
		AU-A- 5232193	23-06-94
		CA-A- 2110672	12-06-94
		CN-A- 1092767	28-09-94
		CZ-A- 9501515	13-12-95

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 95/14968

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-9413696		EP-A- 0662481	12-07-95
		FI-A- 952862	09-06-95
		HR-A- 931484	30-04-96
		HU-A- 73228	29-07-96
		JP-B- 2509147	19-06-96
		JP-A- 7097367	11-04-95
		NO-A- 952294	10-08-95
		PL-A- 309332	02-10-95
		SI-A- 9300647	30-09-94
		WO-A- 9419367	01-09-94
		ZA-A- 9309272	08-08-94
		ZA-A- 9309274	08-08-94

THIS PAGE BLANK (USPTO)

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☐ BLACK BORDERS
- ☐ IMAGE CUT OFF AT TOP, BOTTOM OR SIDES
- ☐ FADED TEXT OR DRAWING
- ☒ BLURRED OR ILLEGIBLE TEXT OR DRAWING
- ☐ SKEWED/SLANTED IMAGES
- ☐ COLOR OR BLACK AND WHITE PHOTOGRAPHS
- ☐ GRAY SCALE DOCUMENTS
- ☐ LINES OR MARKS ON ORIGINAL DOCUMENT
- ☐ REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY
- ☐ OTHER: _____

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.

THIS PAGE BLANK (USPTO)